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# **Analytical Validation and Development of Cefixime Drug Substances** in Pharmaceutical Formulations by Gas Chromatography

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#### **KEYWORDS**

#### Cefixime, Validating the Assay, Chromatography, ICH Q2 (R1)

#### **ABSTRACT:**

The present study was conducted to develop and validate an analytical procedure for the determination of cefixime in Pharmaceutical Formulations. The analytical test attributes and evaluated as per the guidelines of ICH Q2 (R1). The method was validated for the determination of Assay in finished products of cefixime and the method validation parameters were evaluated for the analytical test attribute cefixime meets the acceptance criteria. The results obtained were within the specified limits and the samples were analyzed for test item concentration by Chromatography.

#### INTRODUCTION

The improved controlled drug delivery system avoids dose dumping and results in the most therapeutic administration of a particular drug to a particular person with a particular ailment. pharmacokinetic advantages like, maintenance of constant therapeutic level over a prolonged period of time and thus reduction in fluctuation in therapeutic levels minimizing the risk of resistance especially in case of antibiotics. For the present study cefixime is selected as drug candidate, it fulfills the following characteristics which indicate its suitability for fabrication into the floating drug delivery system. Cefixime is a very poorly soluble in water after its oral administration; it is slowly and incompletely absorbed from the gastrointestinal tract, which resulting into the poor bioavailability i.e., 40-50%. (Aulton, et. al., 2000, Singh, e.t al., 2000, Atyabi, et. al., 1996). So, in order to improve the therapeutic effect of the drug by increasing its bioavailability, we are planning to formulate Cefixime gas powered systems for controlled release with increased gastric retention. The present development study of Cefixime in the form of tablet or capsule which provides a combination of spatial and temporal control of drug delivery to patients for effective therapeutic results (Okeke, et. al., 1998).

The formulation was composed of 69.9% ciprofloxacin base, 0.34% sodiumalginate, 1.03% xanthan gum, 13.7% sodium bicarbonate and 12.1% cross linked polyvinyl pyrrolidone. The viscolyzing agents initially and the gel forming polymer later formed a hydrated gel matrix that entrapped the gas, causing the tablet to float and be retained in the stomach or upper part of the small intestine (spatial control). The hydrated gel matrix created a tortuous diffusion path for the drug, resulting in sustained release of the drug. Baumgartner developed floating of ciprofloxacin tablets Hydrochloride offer a new possibility of treating the stomach infected with Helicobacter pylori (Baumgartner et. al., 2001). The objective of this study was to select suitable materials such as polymers hydroxyl ethyl cellulose (HEC), hydroxyl-propyl cellulose (HPC), HPMCK4M and obtained controlled drug release for more than 8hr from non disintegrated matrices plays an important role in prolonging gastric residence time.

Krogel developed floating-pulsatile drug delivery systems based on a reservoir system consisting of a drug-containing effervescent core and a polymeric coating (Krogel, et. al., 1999). Preliminary studies identified important core and coating properties for the two systems. For the floating system, a polymer coating

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with a high elongation value and high water-and low CO2 permeability's was selected (Eudragit® RL: acetyltributyl citrate 20%, w:w), while for the pulsatile DDS, a weak, semi permeable film, which ruptured after a certain lag time was best (ethyl cellulose: dibutylsebacate 20%, w:w).

Badve developed a hollow calcium pectinate bead by simple process of acid-base reaction during ionotropic cross linking (Badve, e.t al., 2006). Dave optimized a gastro retentive drug delivery system of ranitidine hydrochloride. Guar gum, xanthan gum and hydroxyl propyl methyl cellulose were evaluated for gel-forming properties (Dave, et. al., 2004) Sodium bicarbonate was in corporated as a gas generating agent. Xiaoqiang studied three floating matrix formulations of phenoporlamine hydrochloride based on gas forming agent. Hydroxy propyl methylcellulose K4M and Carbopol 971P NF was used in formulating the hydro gel drug delivery system (Xiaoqiang, et. al., 2006). Yang developed as well able asymmetric triple-layer tablet with Floating ability using hydroxyl propyl methyl cellulose (HPMC) and poly (ethylene oxide) (PEO) as the rate controlling polymeric membrane excipients (Yang, et. al., 1999). Nur developed floating tablets of captopril using HPMC (4000and 15000cps) and carbopol934P. Invitro studies revealed that buoyancy of the tablet is governed by both the swelling of the hydrocolloid particles and tablet porosity (Nur, et. al., 2000). Ozdemir developed floating bilayer tablets with controlled release for furosemide (Ozdemir, et. al., 2000). One layer contained the polymers HPMC 4000, HPMC 100 and CMC (for the control of the drug delivery) and the drug. Pentewar developed a floating drug delivery system of cefixime trihydrate matrix tablet using polymer blends of different viscosity grades of HPMC (Pentewar, et. al., 2010). Gas producing agent used was sodium bicarbonate. The effect of citric acid on the drug release was also investigated by dissolution studies.

#### **METHODOLOGY**

Name of the Product: Cefixime (Cephalosporin

Therapeutic Category: Antibacterial

**Storage condition**: Store in an air tight container below 25°C.

### **RESIDUAL SOLVENTS (By GC)Equipment:**

Gas chromatograph system with Flame Ionization Detector (Shimadzu 2010 or equivalent) Head space auto sampler (AOC 5000 or equivalent) Data handling system (GC solution)

#### **Reagents:**

Methanol :AR grade

Ethanol :AR grade
Ethyl acetate :AR grade
N, N-Dimethylformamide (DMF) :AR grade
Sodium Chloride :AR grade
Water :Mill-Q grade

**Note:** purity of N, N-Dimethylformamide (DMF) and water used in the analysis should be checked for any impurities eluting at the same relative retention times as that of different residual solvents analyzed by this method.

#### **Gas Chromatographic Conditions:**

Col DB-1701 [14% Cynopropylphenyl and 86% dimethyl polysiloxane] capillary column of 30m length, 0.32 mm I.D. And film thickness 1.0 □ m

Detector : FID
Attention : 0
Carrier gas : Nitrogen
Purge gas : Nitrogen

: Hydrogen - 60 kpa equivalent to 50ml.min Zero air- 50 kpa equivalent to 500ml /min Nitrogen - 100 kpa

equivalent to 40ml/min

Column Flow

(pressure) : 20 kPa equivalent to 0.53

ml/min.

Split : 1:10
Stop time : 32 minutes
Temperature : 40°c
Capillary Injector : 220°C
Detector : 260°C

15°C/min.

Column oven temp:  $40^{\circ}$ C (15 min.)  $\longrightarrow$  220°C

(5 min.)

# **EXPERIMENTAL Head Space Condition:**

Cycle : HS-inj
Syringe : 2.5 ml -HS.
Sample volume : 1ml

Sample volume
Incubation temperature: 80°C

Incubation time : 20 min.

Agitation speed : 500 rpm

Syringe temperature : 110°C.

Fill speed : 1 ml/sec.

Pull-up delay : 500 msec.

Inject to : GC-inj.

Inject speed : 1 ml/sec.

#### **Preparation of solutions:**

#### Blank solution:

Add 0.5 ml of dimethyl formamide (DMF) and 0.5 ml of water to the headspace vialcontaining about 0.5 g of sodium chloride and seal the vial immediately.

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#### Preparation of Standard solution:

Solution A: Weigh accurately and transfer about 0.3 g of Ethyl acetate, into a 10 ml volumetric flask containing 5 ml of DMF and make up to volume with the same DMF.

Solution B: Weigh accurately transfers about 0.3 g of Methanol, 0.5 g of Ethanol, into a 50 ml volumetric flask containing about 25 ml of DMF and make up to volume with the sameDMF.

Solution C: Take 0.5 ml of solution A and 0.25 ml solution B into a 25ml clean dry Volumetric flask contain about 15 ml DMF, mix and make up to volume with the same DMF.

Transfer 0.5 ml of the solution C to the headspace vial containing about 0.5 g of sodiumchloride. Add 0.5 ml of water and seal the vial immediately.

Sample solution: Transfer about 0.1 gm of Cefixime weighed accurately, to the headspace vial containing about 0.5 gm of sodium chloride. Add 0.5 ml of DMF and 0.5 ml of water and seal the vialimmediately.

#### **Evaluation Blank:**

Place the sealed vial of blank solution in the sample tray and run the headspace analyzer, record the chromatogram. No interfering peak should be observed at the retention time of analyte peaks.

#### **Evaluation of System Suitability:**

Place sealed vial of standard solution in the sample tray and run the headspace analyzer, record the chromatogram. No interfering peak should be observed at the retention time of analyte peaks.

#### **Procedure:**

Place the sealed vials of sample solution in duplicate in the sample tray and run the headspace analyzer. Record the chromatograms and measure the peak areas of analytes.

The retention time of N, N-Dimethylformide is about 27 min.

#### **Elution Order:**

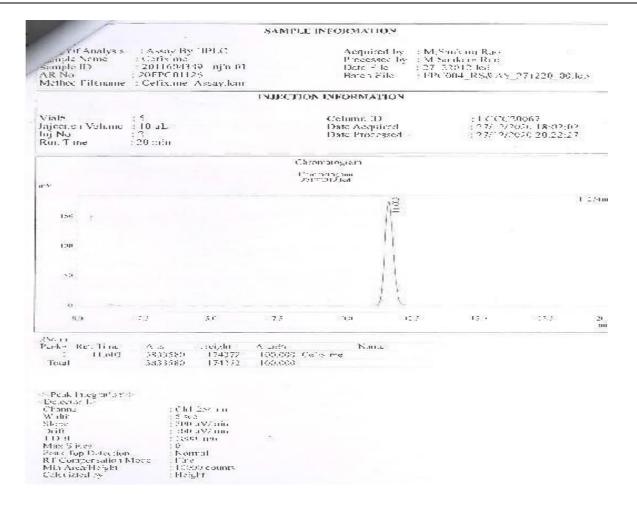
S. No.	Name	RRT
1	Methanol	~0.28
2	Ethanol	~0.36
3	Ethyl Acetate	~0.62
4	N.N-Dimethylformamide	=1.00

#### Chromatograms

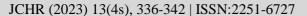
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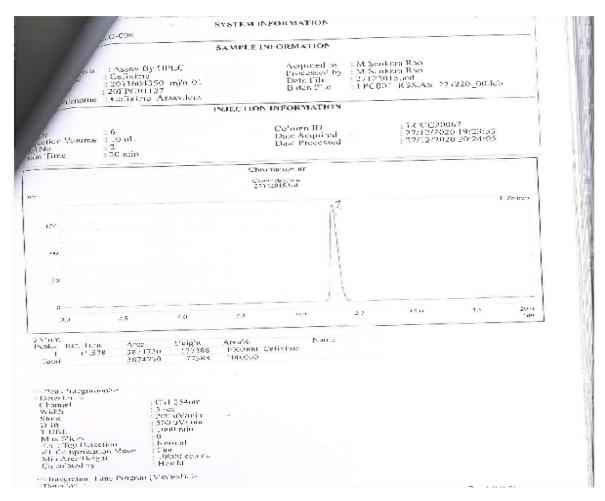




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# ANALYTICAL RESULTS

NALY HCAL RESULTS					
	Experimental Results		Results		
Test No	Test	Specification	Ist Batch	II <sup>nd</sup> Batch	III <sup>rd</sup> Batch
1.	Description	A white to light yellow,crystalline powder.	Light yellow crystalline powder	Light yellow crystalline powder	Light yellow crystalline powder
2.		Soluble in methanol and in propylene glycol; slightly soluble in alcohol, in acetone, and in glycerin; very slightly soluble in 70% sorbitol and in octanol; practically insoluble in ether, in ethyl acetate, in hexane, and in water.	Complies	Complies	Complies
3.	·	IR spectrum must exhibit maxima at the same wave numbers asthe Cefixime working standard spectrum.	Complies	Complies	Complies
4.	Specific rotation (°) (Testsolution 10 mg/ml, in sodium bicarbonate solution (2 in100ml) (on anhydrous basis)	Between -75 and -88	-82°	-83°	-83.0°
5.	Crystallinity	The particles should show birefringence and extinction positions	Complies	Complies	Complies
6.	pH (0.7 mg of cefixime per ml solution in water)	Between 2.6 and 4.1	3.192	3.186	3.095
7.	Water (% w/w)	Between 9.0 to 12.0	11.4	11.3	11.4

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Experimental Results			Results		
Test No					
	Test	Specification	Ist Batch	II <sup>nd</sup> Batch	III <sup>rd</sup> Batch
8.	Chromatographic purity(By HPLC, %	Not more than 1.0Not more	0.17	0.18	0.17
	w/w) Any Individual impurityTotal	than 2.0	0.88	0.88	0.79
	impurities				
9.	Assay as Cefixime (ByHPLC, µg/mg)	Not less than 950 andNot	996	987	992
	(Anhydrous basis)	more than 1030			
10.	Bulk Density* Untapped densityTapped	Report resultsReport results	0.81	0.81	0.77
	density		0.91	0.91	0.86
11.	Particle size*				
	By Sieve (% w/w) /By Malvern (μ)	Report results	#40(P)-100%	#40(P)-100%	#40(P)-100%

Experiment Results			Results			
S. No	Parameters	Limit	Ist Batch	II <sup>nd</sup> Batch	III <sup>rd</sup> Batch	
1.	Residual solvents (by GC) ppm					
	a) Methanol	Not more than 3000Not more	29	28	24	
	b) Ethanol	than 5000	81	86	84	
	c) Ethyl Acetate	Not more than 5000	691	698	823	

#### **CONCLUSION**

From the comprehensive reviewed data and its subsequent evaluation during the Cefixime (Cephalosporins) project work. It is an antibiotic drug, it is demonstrated that the process is robust as all the quality attributes are found within the acceptance criteria and statistical controls as shown in the experimental results. Then we can distribute the product to the formulators. This is considering that the process is adequate and capable of producing the product Cefixime (Cephalosporin antibiotic) to meet the predefined specification.

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