



Herbal Medicines in the Treatment of Type 2 Diabetes Mellitus.

Sorimuthu Pillai Subramanian¹, Subramanian Iyyam Pillai², Renuka Krishnamoorthy¹ and Rajendran Rajitha¹

¹Department of Biochemistry, University of Madras, Guindy Campus, Chennai-600 025.

²Post-Graduate and Research Department of Chemistry, Pachaiyappa's College, Chennai- 600 030.

(Received: 14 June 2024

Revised: 01 July 2024

Accepted: 18 August 2024)

KEYWORDS

Diabetes mellitus,
Herbal medicine,
Phytochemicals,
Trace elements,
Reactive oxygen
species.

ABSTRACT:

Various reports have shown that chronic low-grade inflammation is associated with the risk of developing type 2 diabetes mellitus and that sub-clinical inflammation contributes to insulin resistance and is linked to the characteristics of metabolic syndrome which primarily include chronic hyperglycemia. Oxidative stress stimulates the generation of inflammatory mediators and inflammation which in turn enhances the production of reactive oxygen species. The interaction between chronic hyperglycemia, oxidative stress, systemic inflammation and the factors promoting prevalence of diabetes mellitus, with particular focus on type 2 diabetes and its secondary complications are the main motivation for the compilation of this article. Phytochemicals, the secondary metabolites produced by the plants, are reported to have both significant beneficial and pharmacological properties in alleviating most of the human ailments especially diabetes mellitus. Herbal medicines are often preferred for the treatment of human ailments because of their availability, accessibility, affordability and non-toxicity. Based on our previous studies on the pharmacological and beneficial properties of various medicinal plants, phytochemicals and the mineral contents, the present article adjudicates the essential role of herbal medicines in the treatment of diabetes and its secondary complications.

INTRODUCTION

Diabetes mellitus (DM) has been recognized as one of the four major non-communicable diseases, namely cardiovascular diseases, cancer, chronic respiratory diseases, and diabetes, that insists on imperative awareness from all key shareholders worldwide in an effort to tackle its increasing prevalence and its associated secondary complications [1, 2]. Additionally, it is included as one of the top 10 causes of premature mortality worldwide. Further, DM is established as the third-highest risk factor for worldwide early mortality due to chronic hyperglycemia or severe hypoglycemia [3, 4]. According to the International Diabetes Federation (IDF) report for the year 2022, currently 537 million people are living with DM, and it is projected that by 2045, about 783 million people will be living with DM [5, 6]. This number is lower because, for each diagnosed case, there will be one undiagnosed case in first-world and eight in third-world countries [7, 8]. Factors driving this striking rise in the incidence of diabetes, especially in developing countries, are flaring urbanization and lifestyle changes, including increasing sedentary lifestyles, less physical activity, and the global nutrition

transition from nutrient-rich to energy-directed foods [9, 10]. Diabetes mellitus is a multifactorial, multisystemic and heterogeneous group of disorders essentially characterized by variable degrees of impaired secretion (T1DM) and/or action of insulin (T2DM), leading to persistent elevation in blood glucose levels [11, 12].

ETIOLOGY, CLASSIFICATION AND DIAGNOSIS OF DIABETES MELLITUS

Type 1 diabetes (T1DM) arises due to selective, progressive, and irreversible autoimmune destruction of a critical mass of pancreatic beta cells, varying from rapid to slow decline of insulin synthesis and secretion [13, 14]. Previously, T1DM was termed juvenile diabetes or insulin-dependent diabetes mellitus (IDDM) as it developed principally in adolescence [15-17]. However, the view has changed over the past decade, as T1DM may occur at any age and T2DM patients also require insulin treatment when all the oral hypoglycemic drugs are unsuccessful to control the persistent hyperglycemia [18]. T1DM can develop quickly over weeks or even days. Weight loss is common in people with type 1 diabetes when it first develops and before it is treated,



but it is less common in people with T2DM. The characteristic features of T1DM include.

1. T1DM accounts for nearly 90% of childhood-onset diabetes and 10% of adult-onset diabetes mellitus [19].
2. There is an absolute deficiency of insulin secretion, increased glucagon secretion, and/or failure to respond to oral therapy [20, 21].
3. More prone to diabetic ketoacidosis, a grave pathological condition due to excessive breakdown of muscle proteins and fats [22].
4. The chief contributors to T1DM include autoimmunity, genetics, and environmental factors [23, 24].
5. Nearly 90% of T1DM patients have one or more islet cell auto antibodies such as cytoplasmic islet cell antibodies (ICA), anti-insulin antibodies, glutamate decarboxylase (GADA), and zinc transporter 8 (ZnT8). The presence of two or more auto antibodies is an almost certain predictor of clinical hyperglycemia and T1DM. T1DM patients who are negative for auto antibodies are termed "idiopathic" (Type 1b) [25-28].
6. Occasionally, it occurs in association with other autoimmune disorders.
7. Equal frequency in males and females; obesity is rare; and more than 50% concordance in identical twins [29, 30].
8. The most robust association with viruses and T1DM involves enterovirus species such as Coxsackievirus, Rubella virus, and Hepatitis C virus [31].
9. Increased incidence and prevalence rates of severe hypoglycemic episodes (blood glucose levels below 54 mg/dl) may occur in T1DM patients at least once in two years due to therapeutic hyperinsulinemia, which in turn may arise due to a high dose of insulin administration, incorrect timing related to meals, an increase in glucose utilization during or after physical exercise, an increase in insulin sensitivity due to weight loss, a fall in insulin excretion under pathological conditions such as renal dysfunction or hepatic malfunction, etc [32, 33].

Type 2 diabetes mellitus is the most prevalent form due to numerous causes, with a frequency of 90–95% of

all known cases in the population [34-36]. It characterizes a whole spectrum of clinical syndromes, ranging from insulin secretion defects to impaired action on vital tissues. Age, obesity, and mental stress are also known to play a vital role in the development of T2DM. In the initial stages, the pancreas secretes more insulin (hyperinsulinemia) in order to maintain the blood glucose levels within the physiological range [37]. However, subsequent to the onset of T2DM, the secretion of insulin gradually decreased. The incidence of T2DM at earlier stages recurrently goes undiagnosed because the classic symptoms such as polyuria, polydipsia, and polyphagia develop gradually as hyperglycemia develops, and the insulin levels in such patients are often within the physiological range or elevated [38].

According to the American Diabetes Association (ADA), diabetes mellitus can be diagnosed clinically in three ways, and each, in the absence of unequal hyperglycemia, must be confirmed on a subsequent day, by any of the three methods, namely the levels of fasting plasma glucose (FPG) less than 120 mg/dl, two-hour postprandial glucose less than 140 mg/dl, and random glucose less than 180 mg/dl [39-42]. Nowadays, the levels of glycosylated haemoglobin (HbA1c), a measure of the percentage of haemoglobin that has been bound to glucose for the last three months, are used as a reliable and reproducible index of both diagnostic and prognostic value in the field of diagnosis and treatment of diabetes [43]. HbA1c is defined as specific glycosylated haemoglobin formed by non-enzymatic ketamine reaction between the carbonyl group of glucose and the N-terminal of the amino acid, valine in one or both beta chains of haemoglobin [44].

Recently, HbA1c percentages of less than 7 are regarded as good glycaemic control in diabetic individuals [45-47]. The classical signs (experienced only by the individual) of DM include polyuria, polydipsia, polyphagia, headache, body pain, whole body itching, discomfort, inactivity, and shortness of breath. The conventional symptoms (detected by someone) include unusual weight loss, psoriasis, frequent infections, foot ulcers, and delays in wound healing. The chronic complications of DM include retinopathy, neuropathy, nephropathy, cardiovascular complications, gastrointestinal disorders, skin infections, genital disorders, impotency and bone deformities. There is a



strong link between T2DM, chronic hyperglycemia-induced oxidative stress, and inflammation [48].

ROLE OF OXIDATIVE STRESSES IN DIABETES MELLITUS

Free radicals may be defined as an atom or a diatomic or polyatomic molecule that possesses one or more unpaired electron in its outer shell valency orbit but is neither positive nor negatively charged [49, 50]. Due to their electronic instability, free radicals are inherently highly unstable and extremely active. They always tend to acquire or lose an electron from the adjoining molecule through bonding to gain electronic stability and form a stable compound, acting as an oxidizing or reducing agent [51]. However, during this process of accepting or donating an electron, the neighboring contributor or beneficiary molecule becomes a free radical [52, 53]. Thus, the generation of free radicals is a cascade of reactions. Free radicals have acquired growing importance in the fields of biology and medicine, with particular reference to their role in the etiology of several dreadful diseases [54].

Most of the free radicals are derived from oxygen, and hence they are frequently termed as “Reactive Oxygen Species” (ROS). The gift of using oxygen has enabled humans and animals to metabolize carbohydrates, fats, and proteins to generate energy in the form of ATP. However, there are negative aspects, since breathing pure oxygen (100%) instead of 20% air is harmful to aerobic organisms [55, 56]. Free radicals are the products of the partial reduction of oxygen. When an oxygen molecule splits into single atoms, they have an unpaired electron and become free radicals. It has been estimated that more than 5% of consumed oxygen forms free radicals in our human body [57]. Four-electron reduction of molecular oxygen leads to the generation of water without the generation of ROS, whereas one-electron reduction results in the creation of ROS. The nitrogen-derived free radicals are known as “reactive nitrogen species” [58, 59].

Reactive oxygen and nitrogen species may be radical or non-radical compounds. The non-radical species are not free radicals, but they are voluntarily activating free radical reactions in living organisms [60]. The primary oxygen-free radicals include superoxide and hydroxyl radicals, whereas the non-radical species mainly include hydrogen peroxide, hypochlorous acid, ozone, organic peroxides, aldehydes, peroxy nitrile, and

singlet oxygen. RNS mainly include radical species such as nitric oxide, peroxy nitrile, and nitrogen dioxide, as well as non-radical species such as nitrous acid and dinitrogen tetroxide [61, 62].

Free radicals are generated during several endogenous and exogenous processes. Free radical formation occurs continuously in the cells as a consequence of both enzymatic and non-enzymatic reactions [63, 64]. Enzymatic reactions, which serve as sources of free radicals, include those involved in the respiratory chain, phagocytosis, prostaglandin synthesis, and the cytochrome P-450 system [65, 66]. Free radicals can also be formed in non-enzymatic reactions of oxygen with organic compounds as well as those initiated by ionizing reactions. The mitochondrial electron transport chain (ETC) in the course of oxidative phosphorylation, which generates energy in the form of ATP, is considered to be the primary source of free radicals in the system [67]. Mitochondria combine oxygen and glucose to produce carbon dioxide, water, and ATP. Free radicals are formed in the body either from normal essential metabolic processes in the human body or from external sources such as X-rays, ozone, cigarette smoking, air pollution, and industrial chemicals [68]. Free radical-generating substances can be found in the food we eat, the medicines we take, the air we breathe, and the water we drink. The other sources of free radical formation include alcohol, tobacco smoke, pesticides, and pollutants [69].

In addition to their harmful effects on health, ROS and RNS can have beneficial effects depending on their function, location, and level. For illustration, superoxide ($O_2^{\cdot-}$) and nitric oxide ($\cdot NO$) radicals at low or optimal concentrations are involved in cellular responses and contribute to signaling pathways [70, 71]. Free radicals play an indispensable role in numerous biological processes that are essential for life, such as the intracellular destruction of bacteria by phagocytes, especially by granulocytes and macrophages. Free radicals are also implicated in some cellular signaling processes, known as redox signaling [72]. At low to moderate levels, ROS are beneficial both in regulating processes involving the maintenance of homeostasis as well as a wide variety of cellular functions [73, 74].

Nitric oxide (NO), one of the principal oxides of nitrogen, mediates physiological processes in essentially every organ and tissue. Nitric oxide is an endothelium-



derived relaxing factor (EDRF) biosynthesized endogenously from L-arginine, oxygen, and NADPH by three known NO synthase isoforms: constitutively expressed endothelial NO synthase (eNOS or type 3 NOS) [51], neuronal NO synthase (type 1 NOS), and inducible NO synthase (iNOS or type 2 NOS). Reduction of inorganic nitrate may also yield nitric oxide. Nasal breathing produces nitric oxide within the body. NO acts as a signaling molecule in many physiological and pathological processes [75]. It was proclaimed as the "Molecule of the Year" in 1992. The 1998 Nobel Prize in Physiology or Medicine was awarded for discovering the role of nitric acid as a cardiovascular signaling molecule [76, 77]. Nitric oxide is highly reactive (having a lifetime of a few seconds), yet it diffuses profusely across the cell membranes. These characteristics make nitric oxide ideal for a transient paracrine and autocrine signaling molecule. It exhibits an extremely broad spectrum of functions, including prevention of blood clotting, regulation of blood pressure, roles in neurotransmission and memory formation, and mediation of the antibacterial and anti-tumour activity of macrophages [78, 79].

The inner lining, or endothelium, of blood vessels uses nitric oxide to signal the surrounding smooth muscle to relax, resulting in vasodilation, which is essential for the prevention of cardiovascular diseases [80, 81]. It is also involved in the regulation of blood flow to skeletal muscles, helping to deliver oxygen and other nutrients to the muscles during exercise. Nitric oxide is an important component of the immune response and a powerful neurotransmitter in the central nervous system in the prevention of neurodegenerative diseases [82, 83]. In fact, Sildenafil (Viagra) is a drug that exercises the nitric oxide pathway by enhancing the signals that are downstream of the nitric oxide pathway [84, 85].

On the other hand, excessive production of free radicals provokes structural modification of vital cellular proteins and the alteration of their functions, leading to cellular dysfunction and distraction of vital cellular processes [86]. Elevated levels of free radicals cause lipid, protein, and DNA damage. In particular, free radicals can break the lipid membrane, which contains polyunsaturated fatty acids, and increase membrane fluidity and permeability. Protein damage involves site-specific amino acid modification, peptide chain fragmentation, cross-linked reaction product aggregation,

electric charge alteration, enzymatic inactivation, and proteolysis susceptibility [87]. Additionally, ROS can damage DNA through oxidizing deoxyribose, breaking strands, removing nucleotides, modifying bases, and cross-linking DNA- proteins [88-91]. Disproportionate amounts of free radical generation often lead to cell damage and apoptosis, contributing to many diseases such as cancer, stroke [92], myocardial infarction, diabetes, and other significant conditions [51]. Many cancers are thought to be the result of interactions between free radicals and DNA that lead to mutations that affect the cell cycle, which then lead to neoplasia [93]. Because free radicals are necessary for life, the body has several enzymatic mechanisms to minimize radically induced damage and protect against excessive production of free radicals.

The free radicals vary widely in their reactivity; for example, the reactivity of ROS in decreasing order is: $\text{OH} > \text{O}_2 > \text{H}_2\text{O}_2 > \text{NO}$. Hydroxyl radicals are much more reactive than all other known free radicals, with a half-life period of about 10–10 seconds, and it is estimated that a cell produces around 50 hydroxyl radicals per second [94,95]. It has been reported that the hydroxyl radicals attack any molecule less than a few nanometers from where they are generated [96]. The hydroxyl radical reacts strongly with most organic and inorganic molecules, such as DNA, proteins, lipids, amino acids, sugars, vitamins, and metals, faster than its speed of generation. All mitochondrial enzyme proteins are susceptible to inactivation by hydroxyl radicals, while all amino acid residues of proteins can be oxidized by hydroxyl radicals. It has been estimated that hydroxyl radicals are responsible for more than 60–70% of tissue damage caused by ionizing radiation. Atherosclerosis and cancer, two major causes of death, are salient "free radical" diseases. In atherosclerosis, radical reactions involving diet-derived lipids in the arterial wall and serum to yield peroxides and other substances were reported [97].

Superoxide is a highly reactive free radical generated by the immune system to kill invading microorganisms; phagocytes, such as neutrophils, monocytes, macrophages, mast cells, and dendritic cells, are mobilised by chemotaxis to the site of bacterial infection and mediate damage through their surface receptors. It is involved in extensive damage to DNA, oxidation of cholesterol, proteins with electron density amino acid residues, such as cysteine, methionine,



tryptophan, tyrosine, and histidine [98], and lipids. Even though the human brain weighs a mere ~1400 g, it consumes 20% of the total basal oxygen (O₂) resources to sustain ATP-intensive neuronal activity to power its ~86 billion neurones and their unfathomably complex networks across trillions of synapses supported by ~250–300 billion glia. Depriving the brain of O₂ for just 30 minutes in an ischaemic stroke elicits a devastating toll: every minute, ~1.9 million neurones and ~14 million synapses expire because, without sufficient O₂, mitochondria are unable to reduce O₂ to H₂O to support ATP synthesis, such that even transitory ischaemia is neurodegenerative. The brain is susceptible to oxidative stress because it harnesses chemically diverse reactive species to perform heterogeneous signaling functions. The human brain consumes ~25% of circulating glucose to support neuronal activity [99], corresponding to ~5.6 mg of glucose per 100 g of brain tissue per minute. The fate of glucose in the brain is complex and involves neuronal-glia metabolic coupling. The brain is more susceptible to oxidative stress due to chronic hyperglycemia coupled with low levels of antioxidants.

The liver and brain are the most vulnerable organs next to brain to free radical attack [100]. Improper metabolism of ROS results in the expression of hypoxia-inducible factor-1 alpha, which increases TNF secretion, leading to an immune response that intensifies liver injury and liver dysfunction. Free radicals can damage cellular macromolecules and, therefore, may participate in hepatocellular injury when produced in excess. The chemical reactivity of nitric oxide (NO) is relatively low, and consequently, its direct toxicity is lower than that of ROS. However, it readily reacts with oxygen, producing the peroxy nitrite anion, a very damaging species for DNA, proteins, and lipids [101,102]. Though nitric oxide is a crucial compound involved in many aspects of health, including blood pressure regulation, athletic performance, and brain function, it is toxic at high exposures. The National Institute for Occupational Safety and Health (NIOSH) has set a permissible exposure limit of 25 ppm (30 mg/m³) over an 8-hour work day and at levels of 100 ppm, nitric oxide causes immediate death. However, nitric oxide is readily converted to nitrates and nitrites by oxygen and water, and its cell signaling is deactivated.

ANTIOXIDANTS AND DIABETES

In order to counteract the deleterious effects of free radicals, nature has gifted us with an array of pharmacologically active molecules collectively termed "antioxidants" [103-105]. These antioxidants can be generally classified in many different ways: (i) endogenous and exogenous; (ii) natural and synthetic; (iii) enzymatic and non-enzymatic; (iv) polar and non-polar; and (v) by the mechanisms in which they are involved. Endogenous antioxidants mainly constitute enzymes such as superoxide dismutase (SOD), catalase (CAT), glutathione reductase (GR), and glutathione peroxidase (GPx). Dietary non-enzymatic antioxidants such as vitamin C, vitamin E, carotenoids, polyphenols, flavonoids, and bioflavonoids are exogenous antioxidants that possess *in vivo* activity and are products of the body's metabolism [106, 107].

The first-line defense antioxidants (enzymatic) convert reactive superoxide and hydrogen peroxide into water and oxygen. Non-enzymatic antioxidants can act as a second-line defense against ROS by rapidly inactivating excessive radicals and oxidants. The enzymatic antioxidants further act as the third-line defense involved in detoxification and removal. Antioxidants principally display activities based on three mechanisms: hydrogen atom transfer, single electron transfer, and metal chelation. The antioxidants exert their pharmacological activity through three different pathways: (i) preventive: prevention of free radical formation and its derivatives (ii) interruption: interrupt radical oxidation reaction; and (iii) inactivation: inactivate free radical or radical derivative reaction products [108].

The natural biological processes in our system, such as breathing, digestion of food, detoxification, and utilizing fats for energy generation, produce destructive compounds called free radicals. Oxidative stress is a pathophysiological condition wherein the generation of ROS and RNS devastates the cell's inherent antioxidant defenses, leading to impairment of cellular architecture and metabolic functions [109, 110]. Oxidative stress has been implicated in the initiation, progression, and onset of almost all metabolic, chronic, or cancer disorders, wherein excessive free radicals destroy the cell membrane, inactivate the activity of major enzymes, prevent normal cell division, interfere with energy generation, and cause genetic mutation [111]. Most of the established drugs prescribed for the treatment of



dreadful diseases possess antioxidant properties in addition to their pharmacological efficacy [112].

ROLE OF HERBAL MEDICINES IN THE TREATMENT OF DIABETES

Throughout the world, several medicinal plants have been found successful for the treatment of numerous primary and secondary health complications, and their demand is increasing exponentially due to the growing recognition of natural products as being non-toxic, more efficacious, and easily available at an affordable price. Phytochemicals produced by plants serve as a priceless chemical library for drug discovery in the pharmaceutical industry. In fact, more than 60% of the marketed medicines are distillations, combinations, reproductions, or variations of substances that exist in nature [113]. Our ancestors recommended some of the natural products, which are profusely found in nature, long before their medicinal values were established and attested to by scientific validation.

Herbal medicines are often preferred because of their availability, accessibility, and affordability. Phytochemicals are ecologically derived secondary metabolites synthesized by plants to protect themselves against damage due to environmental stress and microbial attacks [114,115]. However, these phytochemicals are reported to have both beneficial and pharmacological properties in alleviating most human ailments [116,117]. Compared to humans, plants have highly developed protective systems due to their sessile (stationary) nature, and hence, plants serve as a rich source of novel substances for therapeutic applications [118, 119]. However, to ensure the safety and efficacy of herbal medicines, standardization and the development of processing aspects for "herbal medicine" are extremely essential.

We have been actively engaged in the identification of medicinal plants with antidiabetic properties for the past two decades and reported the antidiabetic [120], antilipidemic [121,122] antiulcerogenic [123], and antioxidant [124] properties of *Aloe vera* leaf gel extract in streptozotocin-induced experimental type 2 diabetes in rats. The seeds of *Eugenia jambolana* [125,126] *Momordica Charantia* [127,128] *Terminalia chebula* [129,130], *Nelumbo nucifera* [131,132], *Areca catechu* [133], *Cassia auriculata* [134], *Artocarpus heterophyllus* [135], and *Strychnos Potatorum* [136] are reported to possess

significant antidiabetic and antilipidemic properties. The leaves of *Murraya Koenigii* [137,138] *Pisidium guajava* [139], and *Annona Squamosa* [140, 141] *Euphorbia hirta* [142], *Lippia nodiflora* [143], and *Piper betel* [144-146]. *Sesbania grandiflora* [147] is reported to possess significant beneficial and pharmacological properties in the treatment of diabetes. The fruits such as *Morinda citrifolia* [148], *Ficus benegalensis*[149],and *Pithecellobium dulce* [150,151] *Pracitrullus fistulosus* [152], *Physalis peruviana* [153], *Immature palm fruits* [154,155] exhibit prominent antidiabetic properties. Similarly, biologically active phytochemicals such as *Resveratrol* [156-158] and *fisetin* [159, 160] *gossypin* [161,162] Rosmarinic acid [163,164] *Tang eretin* [165, 166] *Gymnemic acid*, *Trigonelline*, *Ferulic acid* [167, 170] *Silibinin* [171], *Morin* [172, 173] *Sinapic acid* [174,175], *D-Pinitol* [176], *Lupeol* [177], *syringin* [178] *Diosmin* [179, 180] 3- Hydroxyflavone [181], zinc mixed ligand complex [182] and *Swertiamarin* [183,184], *Avicularin* [185, 186] are reported to have significant properties in controlling chronic hyperglycemia induced hyperlipidemia and oxidative stress in experimental type 2 diabetes in rats. The results of the above earlier reports indicate that the observed pharmacological properties of the medicinal plants indicate the synergetic effect of all phytochemicals present in them. Likewise, the observed antidiabetic and antilipidemic properties of the individual phytochemicals may be due to their significant anti-oxidant properties. In traditional medicine system, the medicinal plants used are very often powder or paste forms of the crude herbs, which often contain both the organic and in-organic constituents. However, experimental studies performed so far on pharmacological properties were mostly with the organic active principals. Some of the in-organic elements such as zinc, vanadium, manganese, chromium and potassium can reduce the blood sugar level and their indirect role in the management of diabetes mellitus is being increasingly recognized. These in-organic trace elements are nutritionally essential for healthy life. Hence, we have studied the role of role of vanadium [187-190] in ameliorating chronic hypoglycemia and its secondary complications. Further detailed studies are in progress to elucidate the possible mechanisms by which these phyto-ingredients regulate metabolic processes to alleviate chronic hyperglycemia and its secondary complications.



CONCLUSIONS

In conclusion, the results of the present study provide substantial scientific evidence for the rationale that the phytochemicals and trace elements present in the medicinal plants elicit significant antidiabetic, antilipidemic and antioxidant properties. Isolation and identification of biologically active ingredients from the traditionally important medicinal plants will pave a new pathway in the treatment of diabetes mellitus and its secondary complications.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

REFERENCES

- [1] Farmer, L., 1952. Notes on the History of Diabetes Mellitus: Views Concerning Its Nature and Etiology up to the Discovery of the Role of the Pancreas. *Bulletin of the New York academy of medicine*. 28(6), 408.
- [2] Nissinen, A., Berrios, X., Puska, P., 2001. Community-based non-communicable disease interventions: lessons from developed countries for developing ones. *Bulletin of the world Health Organization*. 79(10), 963-70.
- [3] Nathan, D.M., 1993. Long-term complications of diabetes mellitus. *New England journal of medicine*. 328(23), 1676-85.
- [4] Deshmukh, C.D., Jain, A., Nahata, B., 2015. Diabetes mellitus: a review. *Int. J. Pure Appl. Biosci*. 3(3), 224-30.
- [5] Ong, K.L., Stafford, L.K., McLaughlin, S.A., 2023. Global, regional, and national burden of diabetes from 1990 to 2021, with projections of prevalence to 2050: a systematic analysis for the global burden of disease study 2021. *Lancet*. 402, 203-234.
- [6] Bergman, M., Manco, M., Satman, I., Chan, J., Schmidt, M.I., Sesti, G., Fiorentino, T.V., Abdul-Ghani, M., Jagannathan, R., Aravindakshan, P.K., Gabriel, R., 2024. International Diabetes Federation Position Statement on the 1-hour post-load plasma glucose for the diagnosis of intermediate hyperglycaemia and type 2 diabetes. *Diabetes research and clinical practice*. 209, 111589.
- [7] Sun, H., Saeedi, P., Karuranga, S., 2022. IDF diabetes Atlas: global, regional and country-level diabetes prevalence estimates for 2021 and projections for 2045. *Diabetes Res. Clin.Pract.* 183,109119.
- [8] Kumar, A., Gangwar, R., Ahmad Zargar, A., Kumar, R., Sharma, A., 2024. Prevalence of diabetes in India: A review of IDF diabetes atlas 10th edition. *Current diabetes reviews*. 20(1), 105-14.
- [9] Hu, F.B., Manson, J.E., Stampfer, M.J., 2001. Diet, lifestyle, and the risk of type 2 diabetes mellitus in women. *N Engl J Med*. 345, 790-797.
- [10] Pretorius, B., Ambuko, J., Papargyropoulou, E., Schönfeldt, H.C., 2021. Guiding nutritious food choices and diets along food systems. *Sustainability*.13(17), 9501.
- [11] National Diabetes Data Group., 1979. Classification and diagnosis of diabetes mellitus and other categories of glucose intolerance. *Diabetes*. 28, 1039-1057.
- [12] Kumar, R., Saha, P., Kumar, Y., Sahana, S., Dubey, A., Prakash, O., 2020. A review on diabetes mellitus: type1 & Type2. *World Journal of Pharmacy and Pharmaceutical Sciences*. 9(10), 838-50.
- [13] Gepts, W., 1965. Pathologic anatomy of the pancreas in juvenile diabetes mellitus. *Diabetes*.14, 619-33.
- [14] Roep, B.O., Thomaidou, S., van Tienhoven, R., Zaldumbide, A., 2021. Type 1 diabetes mellitus as a disease of the β -cell (do not blame the immune system?).*Nat. Rev. Endocrinol*. 17(3), 150-161.
- [15] Scott, C.R., Smith, J.M., Craddock, M.M., Pihoker, C., 1997. Characteristics of youth-onset noninsulin-dependent diabetes mellitus and insulin-dependent diabetes mellitus at diagnosis. *Pediatrics*. 100(1), 84-91.
- [16] Fowler, M.J., 2007. Classification of diabetes: not all hyperglycemia is the same. *Clinical diabetes*. 25(2), 74-7. <https://doi.org/10.2337/diaclin.25.2.74>
- [17] Katsarou, A., Gudbjörnsdottir, S., Rawshani, A., Dabelea, D., Bonifacio, E., Anderson, B.J., Jacobsen, L.M., Schatz, D.A., Lernmark, A., 2017. Type 1 diabetes mellitus. *Nature reviews Disease primers*. 3(1), 1-7.
- [18] Hay, L.C., Wilmshurst, E.G., Fulcher, G., 2003. Unrecognized hypo-and hyperglycemia in well-controlled patients with type 2 diabetes mellitus:



- the results of continuous glucose monitoring. *Diabetes technology & therapeutics*. 5(1), 19-26.
- [19] Diagnosis and classification of diabetes mellitus. 2011. *Diabetes Care*, 34(1). doi: 10.2337/dc11-S062.
- [20] Kimball, C.P., Murlin, J.R., 1923. Aqueous extracts of pancreas, III: some precipitation reactions of insulin. *J. Biol Chem*. 58,337-346.
- [21] Muller, T.D., Finan, B., Clemmensen, C., DiMarchi, R.D., Tschop, M.H., 2017. The new biology and pharmacology of glucagon. *Physiol. Rev.* 97(2),721-766.
- [22] Glaser, N., Barnett P., McCaslin, I., 2001. Pediatric emergency medicine collaborative research committee of the american academy of pediatrics: risk factors for cerebral edema in children with diabetic ketoacidosis. *N.Engl. J. Med.* 25, 264–269.
- [23] Eisenbarth, G.S., 1986. Type I diabetes mellitus: a chronic autoimmune disease. *N. Engl. J. Med.* 314, 1360–1368.
- [24] Syed, F.Z., 2022. Type 1 diabetes mellitus. *Annals of internal medicine*. 175(3), 33-48.
- [25] Kanungo, A., Sanjeevi, C.B., 2003. IA-2 autoantibodies are predominant in latent autoimmune diabetes in adults' patients from eastern India. *Ann N Y Acad Sci*. 1005, 390–4.
- [26] Unnikrishnan, A.G., Singh, S.K., Sanjeevi, C.B., 2004. Prevalence of GAD65 antibodies in lean subjects with type 2 diabetes. *Ann N Y Acad Sci*. 1037, 118–21.
- [27] Redondo, M.J., Jeffrey, J., Fain, P.R., 2008. Concordance for islet autoimmunity among monozygotic twins. *N Engl J Med*. 359, 2849–50.
- [28] Patel, S.K., Ma, C.S., Fournalanos, S., Greenfield, J.R., 2021. Autoantibody-negative type 1 diabetes: a neglected subtype. *Trends in Endocrinology & Metabolism*. 32(5), 295-305.
- [29] Matsuda, A., Kuzuya, T., 1994. Relationship between obesity and concordance rate for type 2 (non-insulin-dependent) diabetes mellitus among twins. *Diabetes research and clinical practice*. 26(2), 137-43.
- [30] Bouchard, C., 2021. Genetics of obesity: what we have learned over decades of research. *Obesity*. 29(5), 802-20.
- [31] Jun, H.S., Yoon, J.W., 2004. A new look at viruses in type 1 diabetes. *ILAR journal*. 45(3), 349-74
- [32] Cryer, P.E., Fisher, J.N., Shamon, H., 1994. Hypoglycemia. *Diabetes care*. 17(7), 734-55.
- [33] Nakhleh, A., Shehadeh, N., 2021. Hypoglycemia in diabetes: An update on pathophysiology, treatment, and prevention. *World journal of diabetes*. 12(12), 2036. DOI: 10.4239/wjd.v12.i12.2036.
- [34] Ganda OP., 1995. Prevalence and incidence of secondary and other types of diabetes. National Diabetes Data Group, National Institute of Diabetes and Digestive and Kidney Disease, National Institutes of Health. *Diabetes in America*. 2nd ed. Bethesda, MD: NIH Publication (95-1468).
- [35] Deshpande, A.D., Harris-Hayes, M., Schootman, M., 2008. Epidemiology of diabetes and diabetes-related complications. *Physical therapy*. 88(11), 1254-64. doi: 10.2522/ptj.20080020.
- [36] DeFronzo, R.A., Ferrannini, E., Groop, L., Henry, R.R., Herman, W.H., Holst, J.J., Hu, F.B., Kahn, C.R., Raz, I., Shulman, G.I., Simonson, D.C., 2015. Type 2 diabetes mellitus. *Nature reviews Disease primers*. 1(1), 1-22.
- [37] Zimmet P. Type 2 (non-insulin-dependent) diabetes—an epidemiological overview. *Diabetologia*. 1982 Jun;22(6):399-411.
- [38] Ta, S., 2014. Diagnosis and classification of diabetes mellitus. *Diabetes care*. 37(1), 81-90.
- [39] Wilkerson, H.L., Butler, F.K., Francis, J.O., 1960. The effect of prior carbohydrate intake on the oral glucose tolerance test. *Diabetes*. 9, 386–391.
- [40] Tuomilehto, J., Lindstrom, J., Eriksson, J.G., 2001. Finnish Diabetes Prevention Study Group. Prevention of type 2 diabetes mellitus by changes in lifestyle among subjects with impaired glucose tolerance. *N Engl J Med*. 344, 1343–1350.
- [41] Knowler, W.C., Barrett-Conno,r E., Fowler, S.E., 2002. Diabetes Prevention Program Research Group. Reduction in the incidence of type 2 diabetes with lifestyle intervention or metformin. *N Engl J Med*. 346, 393–403.
- [42] Punthakee, Z., Goldenberg, R. Katz, P., 2018. Definition, classification and diagnosis of diabetes, prediabetes and metabolic syndrome. *Canadian journal of diabetes*. 42, S10-S15.
- [43] Gruev, T., Janikevik Ivanovska, D., Jovcevaska-Mecevska, J., Dimitrov, S., 2011. Differences in hematology and biochemical parameters in diabetic patients. 0457.



- [44] Bookchin, R.M., Gallop, P.M., 1968. Structure of haemoglobin A1c: nature of the N-terminal beta chain blocking group. *Biochem Biophys Res Commun.* 32, 86–93.
- [45] Gavin, L.A., Barth, J., Arnold, D., Shaw, R., 2000. Troglitazone add-on therapy to a combination of sulfonylureas plus metformin achieved and sustained effective diabetes control. *Endocrine practice.* 6(4), 305-10.
- [46] Kilpatrick, E.S., Rigby, A.S., Atkin, S.L., 2007. Variability in the relationship between mean plasma glucose and HbA1c: implications for the assessment of glycemic control. *Clinical chemistry.* 53(5), 897-901.
- [47] Abera RG, Demesse ES, Boko WD. Evaluation of glycemic control and related factors among outpatients with type 2 diabetes at Tikur Anbessa Specialized Hospital, Addis Ababa, Ethiopia: a cross-sectional study. *BMC endocrine disorders.* 2022 Mar 7; 22(1):54.
- [48] Greene, D. A., 1986. Acute and chronic complications of diabetes mellitus in older patients. *The American Journal of Medicine.* 80(5), 39–53. doi:10.1016/0002-9343(86)90536-x.
- [49] Commoner, B., Townsend, J., Pake, G.E., 1954. Free radicals in biological materials. *Nature.* 174(4432), 689–91.
- [50] Sinclair, A.J. and Lunec, J., 1995. Free radicals, oxidative stress and diabetes mellitus. In *Immunopharmacology of free radical species.* pp. 183-198. Academic Press.
- [51] Gutowski, M., Kowalczyk, S., 2013. A study of free radical chemistry: their role and pathophysiological significance. *ActaBiochimicaPolonica.* 60(1), 1-6.
- [52] Mukherji, S.M. and Singh, S.P., 1984. Reaction mechanism in organic chemistry. Macmillan.
- [53] Recknagel, R.O., Glende, E.A., Britton, R.S., 2020. Free radical damage and lipid peroxidation. In *Hepatotoxicology.* 401-436.
- [54] Bajaj, S., Khan, A., 2012. Antioxidants and diabetes. *Indian journal of endocrinology and metabolism.* 16(2), S267-71.
- [55] Fridovich, I., 1999. Fundamental aspects of reactive oxygen species, or what's the matter with oxygen?. *Annals of the New York Academy of Sciences.* 893(1), 13-8.
- [56] Verenich, S., 2010. Role of Oxidative Reactive Species and Antioxidants in Metabolism and Transport of Therapeutic Drugs.
- [57] Younes, M., 1999. Free radicals and reactive oxygen species. In *Toxicology* 111-125. Academic Press.
- [58] Pham-Huy, L.A., Hua He., Pham-Huy, C., 2008. Free Radicals, Antioxidants in Disease and Health. *Int J Biomed Sci.* 4(2), 89–96.
- [59] Ozcan, A., Ogun, M., 2015. Biochemistry of reactive oxygen and nitrogen species. *Basic principles and clinical significance of oxidative stress.* 3, 37-58.
- [60] Boveris, A., 1998. Biochemistry of free radicals: from electrons to tissues. *MEDICINA-BUENOS AIRES.* 58, 350-6.
- [61] Beckman, J.S., Koppenol, W.H., 1996. Nitric oxide, superoxide, and peroxynitrite: the good, the bad, and ugly. *Am. J Physiol.* 271, C1424–37.
- [62] Pourova, J., Kottova, M., Voprsalov, a M., Pour, M., 2010. Reactive oxygen and nitrogen species in normal physiological processes. *Actaphysiologica.* 198(1), 15-35.
- [63] Andrew, P.J., Mayer, B., 1999. Enzymatic function of nitric oxide synthases. *Cardiovasc Res.* 43(3), 521–31.
- [64] Ifeanyi, O.E., 2018. A review on free radicals and antioxidants. *Int. J. Curr. Res. Med. Sci.* 4(2), 123-33.
- [65] Droge, W., 2002. Free radicals in the physiological control of cell function. *Physiological reviews.*
- [66] Martemucci, G., Costagliola, C., Mariano, M., D'andrea, L., Napolitano, P., D'Alessandro, A.G., 2022. Free radical properties, source and targets, antioxidant consumption and health. *Oxygen.* 2(2), 48-78. <https://doi.org/10.3390/oxygen2020006>.
- [67] Pospisil, P., 2009. Production of reactive oxygen species by photosystem II. *BiochimicaetBiophysicaActa (BBA)-Bioenergetics.* 1787(10), 1151-60
- [68] Bartosz, G., 2003. Generation of reactive oxygen species in biological systems. *Comments on Toxicology.* 9(1), 5-21.
- [69] Phaniendra, A., Jestadi, D.B., Periyasamy, L., 2015. Free radicals: properties, sources, targets, and their implication in various diseases. *Indian journal of clinical biochemistry.* 30, 11-26.



- [70] Darley-usmar, V.M., Hogg, N., O'leary, V.J., Wilson, M.T., Moncada, S., 1992. The simultaneous generation of superoxide and nitric oxide can initiate lipid peroxidation in human low density lipoprotein. *Free radical research communications*. 17(1), 9-20.
- [71] Kalyanaraman, B., Darley-USmar, V., Davies, K.J., Dennery, P.A., Forman, H.J., Grisham, M.B., Mann, G.E., Moore, K., Roberts II, L.J., Ischiropoulos, H., 2012. Measuring reactive oxygen and nitrogen species with fluorescent probes: challenges and limitations. *Free radical biology and medicine*. 52(1), 1-6.
- [72] Finkel, T., Holbrook, N.J., 2000. Oxidants, oxidative stress and the biology of ageing. *Nature*. 408(6809), 239-47.
- [73] Vetrovsky, P., Entlicher, G., 1997. Nitrogen (II) oxide (nitric oxide, NO): its origin, fate and physiological significance. A review. *Collection of Czechoslovak chemical communications*. 62(9), 1355-83.
- [74] Bhattacharyya, A., Chattopadhyay, R., Mitra, S., Crowe, S.E., 2014. Oxidative stress: an essential factor in the pathogenesis of gastrointestinal mucosal diseases. *Physiological reviews*. 94(2), 329-54.
- [75] Murad, F., 2004. Discovery of some of the biological effects of nitric oxide and its role in cell signaling. *Bioscience reports*. 24(4-5), 452-74.
- [76] Ignarro, L.J., 2002. After 130 years, the molecular mechanism of action of nitroglycerin is revealed. *Proc. Nat. Acad. Sci. USA*. 99, 7816-7817. doi: 10.1073/pnas.132271799.
- [77] Cary, S.P., Winger, J.A., Derbyshire, E.R. and Marletta, M.A., 2006. Nitric oxide signaling: no longer simply on or off. *Trends in biochemical sciences*. 31(4), 231-239.
- [78] Michel, T., Feron, O., 1997. Nitric oxide synthases: which, where, how, and why? *J Clin. Invest*. 100, 2146-52.
- [79] He, W., Kwesiga, M.P., Gebreyesus, E. and Liu, S., 2019. Nitric Oxide and oxidative stress-mediated cardiovascular functionality: from molecular mechanism to cardiovascular disease. In *Vascular Biology-Selection of Mechanisms and Clinical Applications*. IntechOpen.
- [80] Furchgott, R.F., Zawadzki, J.V., 1980. The obligatory role of endothelial cells in the relaxation of arterial smooth muscle by acetylcholine. *Nature*. 288(5789), 373-376.
- [81] Lobo, V., Patil, A., Phatak, A., Chandra, N., 2010. Free radicals, antioxidants and functional foods: Impact on human health. *Pharmacognosy reviews*. 4(8), 118.
- [82] Prast, H., Philippu, A., 2001. Nitric oxide as modulator of neuronal function. *Progress Neurobiol*. 64, 51-68.
- [83] Volodyaev, I., Vladimirov, Y.A., 2023. Enzymatic Sources of Free Radicals. In *Ultra-Weak Photon Emission from Biological Systems: Endogenous Biophotonics and Intrinsic Bioluminescence*. 219-261. Cham: Springer International Publishing.
- [84] Glossmann, H., Petrischor, G., Bartsch, G., 1999. Molecular mechanisms of the effects of sildenafil (VIAGRA®). *Experimental gerontology*. 34(3), 305-318.
- [85] Dishy, V., Sofowora, G., Harris, P.A., Kandcer, M., Zhan, F., Wood, A.J. Stein, C.M., 2001. The effect of sildenafil on nitric oxide-mediated vasodilation in healthy men. *Clinical Pharmacology & Therapeutics*. 70(3), 270-279. doi: 10.1067/mcp.2001.117995.
- [86] Ayala, D., Ullastres, A., González, J., 2014. Adaptation through chromosomal inversions in *Anopheles*. *Frontiers in Genetics*. 5, 129.
- [87] Sharma, P., Jha, A.B., Dubey, R.S., Pessaraki, M., 2012. Reactive oxygen species, oxidative damage, and antioxidative defense mechanism in plants under stressful conditions. *Journal of botany*. 1, 217037.
- [88] Cadet, J., Wagner, J.R., 2013. DNA base damage by reactive oxygen species, oxidizing agents, and UV radiation. *Cold Spring Harbor perspectives in biology*. 5(2), a012559.
- [89] Tsatsakis, A., Docea, A.O., Constantin, C., Calina, D., Zlatian, O., Nikolouzakis, T.K., Stivaktakis, P.D., Kalogeraki, A., Liesivuori, J., Tzanakakis, G., Neagu, M., 2019. Genotoxic, cytotoxic, and cytopathological effects in rats exposed for 18 months to a mixture of 13 chemicals in doses below NOAEL levels. *Toxicology letters*. 316, 154-170.
- [90] Cadet, J., Angelov, D., Wagner, J.R., 2022. Hydroxyl radical is predominantly involved in oxidatively generated base damage to cellular



- DNA exposed to ionizing radiation. *International Journal of Radiation Biology*. 98(11), 1684-1690.
- [91] Liang, W., Guan, W., Chen, R., Wang, W., Li, J., Xu, K., Li, C., Ai, Q., Lu, W., Liang, H., Li, S., 2020. Cancer patients in SARS-CoV-2 infection: a nationwide analysis in China. *The lancet oncology*. 21(3), 335-337.
- [92] Padureanu, R., Albu, C.V., Mititelu, R.R., Bacanoiu, M.V., Docea, A.O., Calina, D., Padureanu, V., Olaru, G., Sandu, R.E., Malin, R.D., Buga, A.M., 2019. Oxidative stress and inflammation interdependence in multiple sclerosis. *Journal of clinical medicine*. 8(11), 1815.
- [93] Cadenas, E., Davies, K.J., 2000. Mitochondrial free radical generation, oxidative stress, and aging. *Free radical biology and medicine*. 29(3-4), 222-230.
- [94] Fenton HJH. Oxidation of tartaric acid in presence of iron. *Journal of the Chemical Society*. 1894;65(41):899-910
- [95] Gligorovski, S., Strekowski, R., Barbati, S., Vione, D., 2015. Environmental implications of hydroxyl radicals (\cdot OH). *Chemical reviews*. 115(24), 13051-13092.
- [96] Singh, R., Devi, S., Gollen, R., 2015. Role of free radical in atherosclerosis, diabetes and dyslipidaemia: larger-than-life. *Diabetes/metabolism research and reviews*. 31(2), 113-126.
- [97] Afanas' ev, I.B., 2007. Signaling functions of free radicals superoxide & nitric oxide under physiological & pathological conditions. *Molecular biotechnology*. 37(1), 2-4.
- [98] Pellerin, L., 2010. Food for thought: the importance of glucose and other energy substrates for sustaining brain function under varying levels of activity. *Diabetes & metabolism*. 36, S59-S63.
- [99] Vertuani, S., Angusti, A., Manfredini, S., 2004. The antioxidants and pro-antioxidants network: an overview. *Current pharmaceutical design*. 10(14), 1677-1694.
- [100] Vendemiale, G., Grattagliano, I., Altomare, E., 1999. An update on the role of free radicals and antioxidant defense in human disease. *International Journal of Clinical and Laboratory Research*. 29, 49-55.
- [101] Lubos, E., Handy, D.E., Loscalzo, J., 2008. Role of oxidative stress and nitric oxide in atherothrombosis. *Frontiers in bioscience: a journal and virtual library*. 13, 5323.
- [102] Radi, R., 2018. Oxygen radicals, nitric oxide, and peroxynitrite: Redox pathways in molecular medicine. *Proceedings of the National Academy of Sciences*. 115(23), 5839-5848.
- [103] Olcott, H.S., Mattill, H.A., 1936. Antioxidants and the Autoxidation of Fats. VI. Inhibitors. *Journal of the American Chemical Society*. 58(9), 1627-1630.
- [104] Hamid, A.A., Aiyelaagbe, O.O., Usman, L.A., Ameen, O.M., Lawal, A., 2010. Antioxidants: Its medicinal and pharmacological applications. *African Journal of pure and applied chemistry*. 4(8), 142-151.
- [105] Chaudhary, P., Janmeda, P., Docea, A.O., Yeskaliyeva, B., Abdull Razis, A.F., Modu, B., Calina, D., Sharifi-Rad, J., 2023. Oxidative stress, free radicals and antioxidants: Potential crosstalk in the pathophysiology of human diseases. *Frontiers in chemistry*. 11, 1158198. <https://doi.org/10.3389/fchem.2023.1158198>.
- [106] Ratnam, D.V., Ankola, D.D., Bhardwaj, V., Sahana, D.K. Kumar, M.R., 2006. Role of antioxidants in prophylaxis and therapy: A pharmaceutical perspective. *Journal of controlled release*. 113(3), 189-207.
- [107] Tyuryaeva, I., Lyublinskaya, O., 2023. Expected and unexpected effects of pharmacological antioxidants. *International Journal of Molecular Sciences*. 24(11), 9303.
- [108] Forman, H.J., Fisher, A.B., 1981. Antioxidant defenses. In *Oxygen and living processes: an interdisciplinary approach*. 235-249. New York, NY: Springer New York.
- [109] Girotti, A.W., 1985. Mechanisms of lipid peroxidation. *J. Free Radic. Biol. Med.* 1, 87-95.
- [110] Harischandra, D.S., Jin, H., Ghosh, A., Anantharam, V., Kanthasamy, A., Kanthasamy, A.G., 2016. Antioxidants and Redox-Based Therapeutics in Parkinson's Disease. *Inflammation, Aging, and Oxidative Stress*. 261-276.
- [111] Kujoth, G.C., Hiona, A., Pugh, T.D., Someya, S., Panzer, K., Wohlgemuth, S.E., Hofer, T., Seo, A.Y., Sullivan, R., Jobling, W.A., Morrow, J.D., 2005. Mitochondrial DNA mutations, oxidative stress, and apoptosis in mammalian aging. *Science*. 309(5733), 481-484.



- [112] Wondrak, G.T., 2009. Redox-directed cancer therapeutics: molecular mechanisms and opportunities. *Antioxidants & redox signaling*. 11(12), 3013-3069.
- [113] Farnsworth, N.R., Akerele, O., Bingel, A.S., Soejarto, D.D., Guo, Z., 1985. Medicinal plants in therapy. *Bulletin of the World Health Organization*. 63: 965-981.
- [114] Velu, G., Palanichamy, V., Rajan A.P., 2018. Phytochemical and pharmacological importance of plant secondary metabolites in modern medicine. *Bioorganic phase in natural food: an overview*. 135-56.
- [115] Kumar, S., Korra, T., Thakur, R., Arutselvan, R., Kashyap, A.S., Nehela, Y., Chaplygin, V., Minkina, T., Keswani, C., 2023. Role of plant secondary metabolites in defence and transcriptional regulation in response to biotic stress. *Plant Stress*. 8, 100154. <https://doi.org/10.1016/j.stress.2023.100154>.
- [116] Dillard, C.J., German, J.B., 2000. Phytochemicals: nutraceuticals and human health. *Journal of the Science of Food and Agriculture*. 80(12), 1744-56.
- [117] Rabizadeh, F., Mirian, M.S., Doosti, R., Kiani-Anbouhi, R., Eftekhari, E., 2022. Phytochemical classification of medicinal plants used in the treatment of kidney disease based on traditional persian medicine. *Evidence-Based Complementary and Alternative Medicine*. 2022(1), 8022599.
- [118] Ahmad, I., Husain, F.M., Maheshwari, M., Zahin, M., 2014. Medicinal plants and phytochemicals: a potential source of novel antibiofilm agents. *Antibiofilm Agents: From Diagnosis to Treatment and Prevention*. 205-232.
- [119] Llauro Maury, G., Mendez Rodriguez, D., Hendrix, S., Escalona Arranz, J.C., Fung Boix, Y., Pacheco, A.O., Garcia Díaz, J., Morris-Quevedo, H.J., Ferrer Dubois, A., Aleman, E.I., Beenaerts, N., 2020. Antioxidants in plants: A valorization potential emphasizing the need for the conservation of plant biodiversity in Cuba. *Antioxidants*. 9(11), 1048.
- [120] Rajasekaran, S., Sivagnanam, K., Narayanan, V., Subramanian, S., 2001. Hypoglycemic and hypolipidemic effects of *Aloe vera* on experimental Rabbits. *Biomedicine*. 21(4), 40-45.
- [121] Rajasekaran, S., Sivagnanam, K., Ravi, K., Subramanian, S., 2004. Hypoglycemic effect of *Aloe vera* gel on streptozotocin-induced diabetes in experimental rats. *Journal of Medicinal food*. 7(1):61-66.
- [122] Subramanian, S., Sathish Kumar, D., Arulselvan, P., 2006. Wound healing potential of *Aloe vera* leaf gel studied in experimental rabbits. *Asian Journal of Biochemistry*. 1 (2), 178- 185,
- [123] Subramanian, S., Sathish Kumar, D., Arulselvan, P., Senthilkumar, G.P., U. S. Mahadeva Rao., 2007. Evaluation of Anti-ulcerogenic potential of *Aloe vera* leaf gel extract studied in experimental rats. *Journal of Pharmacology and Toxicology*. 2(1), 85-97.
- [124] Rajasekaran, S., Sivagnanam, K., Subramanian, S., 2005. Antioxidant effect of *Aloe vera* gel extract in streptozotocin-induced diabetes in rats. *Pharmacological Reports*. 57, 90-96.
- [125] Ravi, K., Rajasekaran, S., Subramanian, S., 2003. Hypoglycemic effect of *Eugenia jambolana* seed kernels on Streptozotocin-induced diabetes in rats. *Pharmaceutical biology*. 41(8), 598-603.
- [126] Ravi, K., Sathishsekar, D., Subramanian, S., 2004. Hypoglycemic activity of inorganic constituents in *Eugenia jambolana* seed on Streptozotocin-induced diabetes in rats. *Biological Trace Element Research*. 99(1-3), 145-55.
- [127] Sathish Sekar, D., Sivagnanam, K., Subramanian, S., 2005. Antidiabetic activity of *Momordica charantia* seeds on Streptozotocin induced diabetic rats. *Die Pharmazie-An international journal of pharmaceutical sciences*. 60(5), 383-7.
- [128] Sathishsekar, D., Subramanian, S., 2005. Antioxidant properties of *Momordica Charantia* (bitter gourd) seeds on Streptozotocin induced diabetic rats. *Asia Pacific journal of clinical nutrition*. 14(2), 153-158.
- [129] Senthilkumar, G.P., Arulselvan, P., Sathishkumar, D., Subramanian, S., 2006. Anti-diabetic activity of fruits of *Terminalia chebula* on Streptozotocin induced diabetic rats. *Journal of health sciences*. 52(3), 283-91.
- [130] Senthilkumar, G.P., Subramanian, S., 2007. Evaluation of antioxidant potential of *Terminalia chebula* Fruits studied in Streptozotocin-induced diabetic rats. *Pharmaceutical biology*. 45(6), 511-8.



- [131] Subramanian, S., Sivashankari, M., 2010. Antidiabetic and antioxidant potential of *Nelumbo nucifera* seeds in Streptozotocin induced experimental diabetes in rats. *Biomedicine*. 30 (2), 183-191.
- [132] Sivasankari, M., Iyyam Pillai, S., Subramanian, S., Kandaswami, M., 2010. Evaluation of hypoglycemic activity of inorganic constituents in *Nelumbo nucifera* seeds on Streptozotocin-induced diabetes in rats. *Biological trace element research*. 138, 226-237.
- [133] Kavitha, L., Kumaravel, B., Sriram Prasath, G., Subramanian, S., 2013. Beneficial role of *Areca catechu nut* extract in Alloxan-induced Diabetic Rats. *Research Journal of Pharmacognosy and Phytochemistry*. 5(2), 100-108.
- [134] Sriram Prasath, G., Aravind, C., Subramanian, S., 2019. Antidiabetic and Antioxidant Properties of *Cassia auriculata* Flower Extract: An *in vitro* Study. *International Journal of Pharmaceutical Sciences Review and Research*, 55(1), 91-96.
- [135] Suchithra, E.R., Subramanian, S., 2014. Antidiabetic activity of *Artocarpus heterophyllus* rag extract studied in high fat fed-low dose STZ induced experimental type 2 diabetic rats. *Der Pharmacia Lettre*. 6 (3), 102-109.
- [136] Sharmila, C., Subramanian, S.P., 2019. Studies on the Defluoridization Competency of a Mixture of Raw Vermiculite and *Strychnos potatorum linn.* Seeds. *Der Pharma Chemica*. 11(4), 38-43.
- [137] Arulselvan, P., Senthilkumar, G.P., Sathish Kumar, D., Subramanian, S., 2006. Anti-diabetic effect of *Murraya koenigii* leaves on streptozotocin induced diabetic rats. *Die Pharmazie-An International Journal of Pharmaceutical Sciences*. 61(10), 874-7.
- [138] Arulselvan, P., Subramanian, S.P., 2007. Beneficial effects of *Murraya koenigii* leaves on antioxidant defense system and ultra structural changes of pancreatic β -cells in experimental diabetes in rats. *Chemico-biological interactions*. 165(2):155-64.
- [139] Subramanian, S., Haseena Banu, H., Mookambika Ramya Bai, R., Shanmugavalli, R., 2009. Biochemical evaluation of antihyperglycemic and antioxidant nature of *Psidium guajava* leaves extract in Streptozotocin-induced experimental diabetes in rats. *Pharmaceutical biology*. 47(4), 298-303.
- [140] Hayath Basha, S.K., Subramanian, S., 2011. Biochemical evaluation of antidiabetic and antioxidant potentials of *Annona squamosa* leaves extracts studied in STZ induced diabetic rats. *International Journal of Pharmaceutical Sciences and Research*. 2(3), 643-655.
- [141] Hayath Basha, S.K., Subramanian, S., 2012. Antidyslipidemic Property of *Annona Squamosa* Leaves Extract Studied in Streptozotocin-Induced Experimental Diabetes in Rats. *Asian Journal of Research in Chemistry*. 5(2), 234-8.
- [142] Subramanian, S., Bhuvaneshwari, S., Sriram Prasath, G., 2011. Antidiabetic and antioxidant potentials of *Euphorbia hirta* leaves extract studied in streptozotocin-induced experimental diabetes in rats. *General physiology and biophysics*. 30(3), 278-85.
- [143] Subramanian, S., Priya, N., Thamizhinyan, V., 2011. Biochemical evaluation of hypoglycemic, hypolipidemic and antioxidant properties of *Lippia nodiflora* leaves studied in alloxan-induced experimental diabetes in rats. *Research Journal of Pharmacology and Pharmacodynamics*. 3(6), 299-304.
- [144] Radhika, K., Kumaravel, B., Thamizhiniyan, V., Subramanian, S., 2013. Biochemical evaluation of antidiabetic activity of *Piper betel* leaves extract in alloxan-induced diabetic rats. *Asian Journal of Research in Chemistry*. 6(1), 76-82.
- [145] Srividya, S., Roshana Devi, V., Subramanian, S., 2015. Hypoglycemic and hypolipidemic properties of hydroxychavicol, a major phenolic compound from the leaves of *Piper betle linn.* studied in high fat diet fed-low dose STZ induced experimental type 2 diabetes in rats. *Der Pharmacia Lettre*. 7(11), 130-40.
- [146] Srividya, S., Roshana Devi, V., Subramanian, S., 2017. Evaluation of Antioxidant Properties of Dihydroxychavicol, a Major Phenolic Compound Isolated from the Leaves of *Piper Betle. Linn* . *BAOJ Diabet*. 3(1), 018.
- [147] Sangeetha, A., Sriram Prasath, G., Subramanian, S., 2014. Antihyperglycemic and antioxidant potentials of *Sesbania grandiflora* leaves studied in STZ induced experimental diabetic rats. *International Journal of Pharmaceutical Science and Research*, 5(6), 2266-2275.
- [148] Mahadeva Rao, U.S., Subramanian, S., 2009. Biochemical evaluation of antihyperglycemic and



- antioxidative effects of *Morinda citrifolia* fruit extract studied in Streptozotocin-induced diabetic rats. *Medicinal Chemistry Research*. 18, 433-46.
- [149] Kavi Priya, S., Thamizhiniyan, V., Subramanian, S., 2013. Antidiabetic potential of *Ficus bengalensis* fruit extract studied in alloxan-induced experimental diabetes in rats. *Research Journal of Pharmacology and Pharmacodynamics*. 5(2), 110-118.
- [150] Pradeepa, S., Subramanian, S., Kaviyarasan, V., 2013. Biochemical evaluation of antidiabetic properties of *Pithecellobium dulce* fruits studied in Streptozotocin induced experimental diabetic rats. *International Journal of Herbal Medicine*. 1(4), 21-28.
- [151] Pradeepa, S., Subramanian, S., Kaviyarasan, V., 2014. Antioxidant role of *Pithecellobium dulce* fruit pulp extract in ameliorating hyperglycemia induced oxidative stress studied in Streptozotocin induced experimental diabetic rats. *Journal of Pharmacy Research*. 8(3), 377-384.
- [152] Karandikar, A., Sriram Prasath, G., Subramanian, S., 2014. Evaluation of Antidiabetic and antioxidant activity of *Praecitrullus fistulosus* fruits in STZ Induced Diabetic Rats. *Research Journal of Pharmacy and Technology*. 7(2), 196-203.
- [153] Sathyadevi, M., Suchithra, E.R., Subramanian, S., 2014. *Physalis peruviana* Linn. fruit extract improves insulin sensitivity and ameliorates hyperglycemia in high-fat diet low dose STZ-induced type 2 diabetic rats. *Journal of Pharmacy Research*, 8(4), 625-632.
- [154] Renuka, K., Parvathi, N., Subramanian, S.P., 2020. Biochemical studies on the antidiabetic properties of *Immature Palmyra Palm* fruits studied in high fat diet fed-low dose Streptozotocin induced type 2 diabetes in rats. *GSC Biological and Pharmaceutical Sciences*. 12(3), 223-35.
- [155] Renuka, K., Gopalakrishnan, V., Subramanian, S., 2020. Evaluation of Antioxidant Properties of *Immature Palmyra Palm* Fruits Extract Studied in High Fat Diet Fed-Low Dose Streptozotocin Induced Experimental Diabetes in Rats. *International Journal of Recent Scientific Research*. 11(02) (A), 37216-37224.
- [156] Palsamy, P., Subramanian, S., 2008. Resveratrol, a natural phytoalexin, normalizes hyperglycemia in streptozotocin-nicotinamide induced experimental diabetic rats. *Biomedicine and Pharmacotherapy*. 62(9), 598-605.
- [157] Palsamy, P., Subramanian, S., 2009. Modulatory effects of resveratrol on attenuating the key enzymes activities of carbohydrate metabolism in Streptozotocin-nicotinamide-induced diabetic rats. *Chemico-biological interactions*. 179, 356-62.
- [158] Rajitha Rajendran., Iyyam Pillai Subramanian., Sorimuthu Pillai Subramanian., 2024. Investigations Using Molecular Docking and Simulation on the Antidiabetic Activity of Resveratrol Aldehyde Molecule. *Journal of Chemical Health Risks*. 12(4), 490-503.
- [159] Sriram Prasath, G., Subramanian, S., 2011. *Fisetin*, a bioflavonoid ameliorates hyperglycemia in STZ-induced experimental diabetes in rats. *International Journal of Pharmaceutical Sciences Review and Research*. 6(1), 68-74.
- [160] Sriram Prasath, G., Shanmuga Sundaram, C., Subramanian, S., 2013. Fisetin averts oxidative stress in pancreatic tissues of Streptozotocin-induced diabetic rats. *Endocrine*. 44, 359-68.
- [161] Thamizhiniyan, V., Subramanian, S., 2012. Antidiabetic activity of *gossypin*, a pentahydroxyflavone glucoside in Streptozotocin induced experimental diabetes in rats. *Journal of Diabetes*. 4, 41-46.
- [162] Thamizhiniyan, V., Vijayaraghavan, K., Subramanian, S.P., 2012. *Gossypin*, a flavonol glucoside protects pancreatic beta-cells from glucotoxicity in Streptozotocin-induced experimental diabetes in rats. *Biomedicine and Preventive Nutrition*. 2(4), 239-45.
- [163] Jayanthi, G., Subramanian, S., 2013. Extraction, isolation and characterization of rosmarinic acid, a major polyphenol in non-volatile constituent of mint leaves. *Asian Journal of Research in Chemistry*. 6(12), 1160-1165.
- [164] Jayanthi, G., Subramanian, S., 2014. Rosmarinic acid, a polyphenol, ameliorates hyperglycemia by regulating the key enzymes of carbohydrate metabolism in high fat diet-STZ induced experimental diabetes mellitus. *Biomedicine and Preventive Nutrition*. 4(3), 431-437.
- [165] Lakshmi, A., Subramanian, S., 2014. Chemotherapeutic effect of *tangeretin*, a polymethoxylated flavone studied in 7, 12-



- dimethylbenz (a) anthracene induced mammary carcinoma in experimental rats. *Biochimie*. 99, 96-109.
- [166] Lakshmi, A., Subramanian, S., 2015. *Tangeretin*, a citrus flavonoid attenuates oxidative stress and protects hepatocellular architecture in rats with 7, 12-dimethylbenz (a) anthracene induced experimental mammary carcinoma. *Journal of Functional Foods*. 15, 339–353.
- [167] Subramanian, S., Sriram Prasath, G., 2014. *Trigonelline* improves insulin sensitivity and modulates glucose homeostasis in high fat fed-Streptozotocin induced type 2 diabetic rats. *Journal of Pharmacy Research*. 8(4), 563-569.
- [168] Subramanian, S., Sriram Prasath, G., 2014. Antidiabetic and antidyslipidemic nature of *trigonelline*, a major alkaloid of fenugreek seeds studied in high-fat-fed and low-dose Streptozotocin-induced experimental diabetic rats. *Biomedicine and Preventive Nutrition*. 4(4), 475-80.
- [169] Roshana Devi, V., Sriram Prasath, G., Subramanian, S., 2015. Antidiabetic properties of GTF-231, an ayurvedic formulation studied in high fat diet fed-low dose STZ induced experimental type 2 diabetes in rats. *Der Pharmacia Lettre*. 7 (7), 113-123.
- [170] Roshana Devi, V., Subramanian, S., 2016. Biochemical evaluation of hypoglycemic effect of GTF-231, a polyherbal preparation in high fat diet fed-low dose STZ-induced experimental type 2 diabetes in rats. *Der Pharmacia Lettre*. 8(18), 121-132.
- [171] Uma Maheswari, J., Iyyam Pillai, S., Subramanian, S., 2015. Zinc-Silibinin complex: Synthesis, spectral characterization and biochemical evaluation of antidiabetic potential in high fat fed low dose STZ induced type 2 diabetic rats. *Journal of Chemical and Pharmaceutical Research*. 7(3), 2051-2064.
- [172] Sendrayaperumal, V., Iyyam Pillai, S., Subramanian, S., 2014. Design, synthesis and characterization of zinc–morin, a metal flavonol complex and evaluation of its antidiabetic potential in HFD–STZ induced type 2 diabetes in rats. *Chemico-biological interactions*. 219, 9-17.
- [173] Sendrayaperumal, V., Iyyam Pillai, S., Gowthaman, D., Subramanian, S., 2014. DNA binding, DNA cleavage and BSA interaction of Vanadium-Morin complex. *International Journal of Innovative Research in Science and Engineering*.
- [174] Nithya, R., Subramanian, S., 2015. Acid, a Naturally Occurring Carboxylic Acid Derivative Ameliorates Hyperglycemia in High Fat Diet-Low Dose STZ Induced Experimental Diabetic Rats. *International Journal of Scientific Engineering and Technology Research*, 4(30), 5746-5750.
- [175] Nithya, R., Roshana Devi, V., Selvam, R., Subramanian, S.P., 2017. Sinapic acid regulates glucose homeostasis by modulating the activities of carbohydrate metabolizing enzymes in high fat diet fed-low dose STZ induced experimental type 2 diabetes in rats. *Global Journal of Obesity, Diabetes and Metabolic Syndrome*. 4(2), 054-061.
- [176] Sivakumar, S., Subramanian, S., 2009. D-Pinitol attenuates the impaired activities of hepatic key enzymes in carbohydrate anglycogen metabolism of Streptozotocin-induced diabetic rats. *General Physiology and Biophysics*. 28, 233-241.
- [177] Saratha, V., Iyyam Pillai, S., Subramanian, S., 2011. Isolation and Characterization of *Lupeol*, a triterpenoid from *Calotropis gigantea* latex. *International Journal of Pharmaceutical Sciences Review and Research*. 10 (2), 54-57.
- [178] Shanmuga Sundaram, C., Iyyam Pillai, S., Subramanian, S., 2014. Isolation, characterization of syringin, phenylpropanoid glycoside from *Musa paradisiaca tepal* extract and evaluation of its antidiabetic effect in Streptozotocin-induced diabetic rats. *Biomedicine & Preventive Nutrition*. 4, 105–111.
- [179] Gopalakrishnan, V., Iyyam Pillai, S., Subramanian, S., 2015. Synthesis, Spectral Characterization, and Biochemical Evaluation of Antidiabetic Properties of a New Zinc-Diosmin Complex Studied in High Fat Diet Fed-Low Dose Streptozotocin Induced Experimental Type 2 Diabetes in Rats. *Biochemistry Research International*. 1, 350829.
- [180] Gopalakrishnan, V., Subramanian, S., 2016. Zinc-diosmin complex ameliorates oxidative stress in the pancreatic tissues of experimental rats with type 2 diabetes induced by high fat diet fed-low dose streptozotocin. *Der Pharmacia Lettre*. 8 (2), 398-407.



- [181] Jaiganesh, C., Roshana Devi, V., Iyyam Pillai, S., Subramanian, S., 2017. Synthesis, Characterization And Evaluation of Antidiabetic Properties of A New Metformin- 3-Hydroxyflavone Complex Studied In High Fat Diet Fed - Low Dose Streptozotocin Induced Experimental Type 2 Diabetes In Wistar Rats. *International Journal of Pharma and Bio Sciences*. 8(3), 1-15.
- [182] Muruganatham, K., Roshana Devi, V., Iyyam Pillai, S., Subramanian, S., 2018. Synthesis and evaluation of antidiabetic properties of a zinc mixed ligand complex in high-fat diet - low-dose streptozotocin-induced diabetic rats. *Asian Journal of Pharmaceutical and Clinical Research*. 11 (5), 429-438.
- [183] Selvam, R., Muruganatham, K., Subramanian, S., 2018. Biochemical evaluation of antidiabetic properties of swertiamarin, a secoiridoid glycoside of *enicostemma littorale* leaves, studied in high-fat diet-fed low-dose streptozotocin-induced type 2 diabetic rats. *Asian Journal of Pharmaceutical and Clinical Research*. 11(10), 486-492.
- [184] Selvam, R., Muruganatham, K., Subramanian, S., 2019. Antioxidant properties of swertiamarin, from *Enicostemma Littorale blume* leaves studied in high fat fed and low-dose streptozotocin induced diabetic rats. *Asian Journal of Pharmacy and Pharmacology*. 5(2), 344-352.
- [185] Neeli Parvathi., Subramanian Iyyam Pillai., Sorimuthu Pillai Subramanian., 2020. Design, synthesis and spectral characterization of a new Zinc-Avicularin, a metal flavonolcomplex and evaluation of its toxicity and antidiabetic efficacy in HFD- Low Dose Streptozotocin Induced Experimental Type 2 Diabetes in Rats. *Diabetes*. 6(2), 9-18.
- [186] Neeli Parvathy., Iyyam Pillai Subramanian., Sorimuthu Pillai Subramanian., 2023. Biochemical Evaluation of Antioxidant Properties of A Zn-Avicularin Complex Studied in High Fat Diet Fed-Low Dose Streptozotocin Induced Experimental Type2 Diabetes in Rats. *Research Journal of Pharmacy and Technology*. 16(1), 145-152,
- [187] Ramachandran, B., Sathishsekar, D., Kandaswamy, M., Narayanan, V., Subramanian, S., 2004. Hypoglycemic effect of macrocyclic binuclear oxovanadium (IV) complex on Streptozotocin-induced diabetic rats. *Experimental Diabetes Research*. 5, 137-142,
- [188] Iyyam Pillai, S., Subramanian, S., Kandaswamy, M., 2013. A novel insulin mimetic vanadium flavonol complex: Synthesis, characterization and in vivo evaluation in STZ-induced rats. *European Journal of Medicinal Chemistry*. 63, 109-117.
- [189] Kalavakunda Vijayaraghavan., Subramanian Iyyam Pillai., Gowthaman, D, Sorimuthu Pillai Subramanian., 2012. Design, synthesis and characterization of zinc-3 hydroxy flavone, a novel zinc metallo complex for the treatment of experimental diabetes in rats. *European Journal of Pharmacology*. 680, 122-129.
- [190] Shanthakumari, D., Srinivasalu, S., Subramanian, S., 2006. Antioxidant defense system in red blood cell lysates of men with dental fluorosis living in Tamilnadu, India. *Fluoride*. 39(3), 231-239.