www.jchr.org

JCHR (2024) 14(2), 3718-3725 | ISSN:2251-6727



Comparative Evaluation of Antimicrobial Activity Associated with Transmucosal Components of Implants

Dr. Girish Nazirkar¹, Dr. Prabhakar Angadi², Dr. Ashlesha Marathe³, Dr. Riya More⁴, Dr. Palak Boob⁴, Dr. Hitendra R. Jain⁵

(Received: 07 January 2024 Revised: 12 February 2024

Accepted: 06 March 2024)

KEYWORDS

Antimicrobial activity, transmucosal components, prosthetic abutment

ABSTRACT:

Background: Recently, there has been a growing interest in mucointegration as the formation of an early and long-standing soft tissue barrier seems essential for both the initial healing and long-term implant survival. The mucointegration is of major importance to ensure good prognosis of implant and to prevent bacterial progression from the oral cavity on the implant surface. Another important parameter is the design and material composition of the trans-mucosal components because it can create a stabilizing ring of connective tissue to protect underlying structures. Transmucosal components include healing abutment and prosthetic abutment. Prosthetic abutments must have properties including biocompatibility, polishability, antimicrobial properties, etc. Few of various prosthetic abutments which are commonly available were studied for antimicrobial activity in this paper.

Aim: To evaluate and compare antimicrobial activity on prosthetic abutment of implant Material & Methods: Various implant prosthetic abutment materials which are available today include titanium, cobalt chromium, zirconia polished and zirconia glazed, peek etc. Out of these titanium, cobalt chromium, zirconia polished and glazed zirconia were studied for antimicrobial activity. The saliva samples were collected and depending upon prosthetic abutments selected, it was divided into various groups. The selected prosthetic abutments were immersed in saliva samples and cultured. After specific period of time microbial flora from the prosthetic abutments was scrapped and identified under a microscope for its type and quantity then the groups were compared.

Results: Results showed titanium abutment material showed less microbial activity as compared to other groups, while cobalt chromium abutment material showed high microbial activity. Glazed and polished zirconia material did not show any significant difference with each other and titanium abutment group.

Conclusion: Less the microbes present on the prosthetic abutment material, more was antimicrobial activity of the material and vice versa. Better the antimicrobial activity, better the soft tissue health and mucointegration and thus leading to long term success of implants.

INTRODUCTION:

Treatment with dental implants is a commonly recognized prosthodontic procedure. The need for long-term stability and efficient functional rehabilitation is

growing along with the aging of society.¹ Osseointegration has been regarded as a crucial and essential component of implant success.² Furthermore, one of the key elements affecting the long-term results of dental implant therapy is the stability of the soft tissue

¹Professor & HOD, SMBT Dental College and Hospital, Sangamner, Amrutnagar

²Professor, SMBT Dental College and Hospital, Sangamner, Amrutnagar

³Reader, SMBT Dental College and Hospital, Sangamner, Amrutnagar

⁴PG Resident, SMBT Dental College and Hospital, Sangamner, Amrutnagar

⁵ Reader & HOD, Dept. of Public Health Dentistry, S.M.B.T Dental College & Hospital, Sangamner, Amrutnagar

www.jchr.org

JCHR (2024) 14(2), 3718-3725 | ISSN:2251-6727



surrounding the implant.³ Gaining knowledge of the relationship between load distribution at the bone—implant contact and the intricate design of the implant and abutment is crucial.⁴ The mechanical stability of implant abutments can be impacted by the vastly varied features of the various materials and designs.⁵

Mucointegration has drawn more attention lately since it appears that the establishment of an early and durable soft tissues barrier is crucial for both the initial healing process and the long-term survival of implants. The mucointegration is of major importance to ensure good prognosis of implant and to prevent bacterial progression from the oral cavity on the implant surface. Another important parameter is the design and material components of the transmucosal components because it can create a stabilizing ring of connective tissue to protect underlying structures.

The surrounding soft tissues play a critical role in the long-term viability of dental implants by acting as a barrier against bacterial colonization and preventing peri-implant disease. A perfect transmucosal implant component should reduce plaque buildup and bacterial colonization while simultaneously promoting epithelial and connective tissue attachment and its maintenance over time. 8

Research has indicated that the surface characteristics of dental implants significantly influence the initial bacterial adherence. Bit is vitally critical for implant materials exposed to the oral cavity to impede early colonization because early colonizers, like as streptococci, establish an environment that also supports the accumulation of later colonists. Factors include the transmucosal parts of implants and the chemical makeup and surface roughness of abutments have a significant impact on plaque formation. But the surface roughness of abutments have a significant impact on plaque formation.

Transmucosal components include healing abutment and prosthetic abutment. Prosthetic abutments must have properties including biocompatibility, polishability, antimicrobial properties, etc. Various kinds of implant abutment materials such as titanium, cast gold alloy, zirconia, cobalt chromium, PEEK, are used.

Few of the prosthetic abutments which are commonly available will be studied for antimicrobial activity in this study.

METHODOLOGY:

SAMPLES:

Group A: Type and quantity of microbial flora on 3 titanium abutment.

Group B: Type and quantity of microbial flora on 3 cobalt chromium abutment

Group C: Type and quantity of microbial flora on 3 zirconia polised abutment

Group D: Type and quantity of microbial flora on 3 zirconia glazed abutment

The sample size came out to be 12.

PROCEDURE OF THE STUDY:

This study is an In-vitro study and is held in Department of Prosthodontic and Crown & Bridge. The saliva samples was collected post lunch and depending upon transmucosal components selected, it was divided into four A,B,C and D groups. The selected transmucosal components were immersed in saliva samples and was incubated for 72 hours in an incubator. After that, transmucosal components was cultured in three different culture mediums i.e Mac Conkey's agar medium, Blood agar medium, Nutrient agar medium for 24 hours. Then, colonies were scrapped from culture media with the help of nicrome wire loop.

Plates displaying growth of colonies were subjected to gram staining protocol. Then colonies were observed under microscope for quantity. Bacterial count was determined in Colony Forming Units. These were identified under a microscope for its quantity then the groups will be compared with each other.

STATISTICAL TESTS:

- Statistical analysis was performed using Statistical Product and Service Solution (SPSS) version 16 for windows (SPSS Inc., Chicago, IL).
- Descriptive quantitative data was expressed in mean and standard deviation respectively.
- Overall comparison between four groups will be done using Anova f test followed by Tukey post hoc test to find pair wise difference.

www.jchr.org

JCHR (2024) 14(2), 3718-3725 | ISSN:2251-6727



 Confidence interval is set at 95% and probability of alpha error set at 5%. Power of study set at 80%.

RESULTS:

Overall intergroup comparison was done of mean antimicrobial activity associated with transmucosal components of four different implant materials respectively using One way Anova F test. The results of Blood agar, Mac Conkey agar and Nutrient agar respectively showed 2.55,3.9,1.8 * 10³ cfu for Group A, 81.9,93.8,65.4 * 10³ cfu for Group B, 6.6,8.2,5.3 * 10³ cfu for Group C and 5.4,5.2,3.1 * 10³ cfu for Group D. Thus One way Anova F test showed values of Blood

agar as 37.8 and that of Mac Conkey agar 24.9 and Nutrient agar 20.1.

Pairwise intergroup comparison was done of mean antimicrobial activity associated with transmucosal components of four different implant materials respectively using Tukey's post hoc test. The comparison between Blood agar, Mac Conkey agar and Nutrient agar respectively showed Group A vs Group B p <0.001 for all mediums, Group A vs Group C p = 0.143,0.084,0.065, Group A vs Group D p = 0.195,0.472, 0,245 , Group B vs Group C p <0.001 for all mediums , Group B vs Group D p <0.001 for all mediums , Group C vs Group D p=0.872,0.751,0.617 (Table 1, 2)(Graph 1)

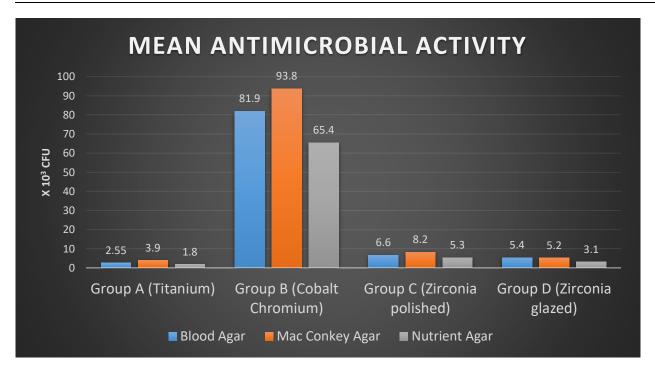
<u>Table 1: Overall intergroup comparison of mean antimicrobial activity associated with transmucosal components of four different implant materials respectively using One way Anova F test</u>

(x 10 ³ CFU)	Blood Agar Mean (SD)	Mac Conkey Agar Mean (SD)	Nutrient Agar Mean (SD)
Group A (Titanium abutment)	2.55 (1.3)	3.9 (1.7)	1.8 (0.6)
Group B (Cobalt Chromium)	81.9 (43.1)	93.8 (51.2)	65.4 (23.8)
Group C (Zirconia polished)	6.6 (2.6)	8.2 (3.7)	5.3 (1.7)
Group D (Zirconia glazed)	5.4 (2.1)	5.2 (1.8)	3.1 (2.4)
One way Anova F test	F = 37.8	F = 24.9	F = 20.1
P value, Significance	p < 0.001**	p < 0.001**	p =< 0.001**

www.jchr.org

JCHR (2024) 14(2), 3718-3725 | ISSN:2251-6727





<u>Graph 1: Overall intergroup comparison of mean antimicrobial activity associated with transmucosal components of four different implant materials respectively</u>

<u>Table 2: Pairwise intergroup comparison of mean antimicrobial activity associated with transmucosal components of four different implant materials respectively using Tukey's post hoc test</u>

	Blood Agar Mean (SD)	Mac Conkey Agar Mean (SD)	Nutrient Agar Mean (SD)
Group A (Titanium abutment) vs Group B (Cobalt Chromium)	p < 0.001**	p<0.001**	p<0.001**
Group A (Titanium abutment) vs Group C (Zirconia polished)	p = 0.143	p =0.084	p =0.065
Group A (Titanium abutment) vs Group D (Zirconia glazed)	p = 0.195	p = 0.472	p =0.245
Group B (Cobalt Chromium) vs Group C (Zirconia polished)	p<0.001**	p<0.001**	p<0.001**

www.jchr.org

JCHR (2024) 14(2), 3718-3725 | ISSN:2251-6727



Group B (Cobalt Chromium) vs Group D (Zirconia glazed)	p<0.001**	p<0.001**	p<0.001**
Group C (Zirconia polished) vs Group D (Zirconia glazed)	p =0.872	p = 0.751	p =0.617

DISCUSSION:

Recently, there has been a growing interest in mucointegration as the formation of an early and long standing soft tissues barrier seems essential for both the initial healing and long term implant survival. The mucointegration is of major importance to ensure good prognosis of implant and to prevent bacterial progression from the oral cavity on the implant surface.

The results showed least antimicromial activity with respect to titanium abutment, while Cobalt chromium showed high microbial activity. Glazed and polished zirconia did not show any significant difference as compared to titanium.

The risks for the development of peri-implant inflammation have been widely studied in the past. Once peri-implantitis is established, it does not respond predictably to treatment. Therefore, it appears that the best management of plaque-induced peri-implant inflammatory diseases is prevention. To avoid the development of peri-implantitis not only solid osseointegration of the implants but also a robust soft-tissue integration at the transmucosal region is mandatory, since it is the first barrier against a bacterial invasion.

As the colonisation of the oral microflora is of particular clinical significance, our in-vitro study is of importance as they show that the titanium abutments had little inhibitory effect on the microflora as compared to the other groups.

REFERENCES:

 Steigenga, J.T.; al-Shammari, K.F.; Nociti, F.H.; Misch, C.E.; Wang, H.L. Dental implant design and its relationship to long-term implant success. Implant. Dent. 2003, 12, 306–317.

- 2. Cervino G, Germanà A, Fiorillo L, D'Amico C, Abbate F, Cicciù M. Passant Connection Screw of Dental Implants: An In Vitro SEM Preliminary Study. Biomed Res Int. 2022 Apr 21;2022:9720488.
- 3.Thoma DS, Gil A, Hämmerle CHF, Jung RE. Management and prevention of soft tissue complications in implant dentistry. Periodontol 2000. 2022 Feb;88(1):116-129.
- Furuhashi A, Ayukawa Y, Atsuta I, Rakhmatia YD, Koyano K. Soft Tissue Interface with Various Kinds of Implant Abutment Materials. J Clin Med. 2021 May 28;10(11):2386.
- 5. Vinhas AS, Aroso C, Salazar F, López-Jarana P, Ríos-Santos JV, Herrero-Climent M. Review of the Mechanical Behavior of Different Implant-Abutment Connections. Int J Environ Res Public Health. 2020 Nov 23:17(22):8685.
- 6.Borie M, Lecloux G, Bosshardt D, Barrantes A, Haugen HJ, Lambert F, Bacevic M. Peri-implant soft tissue integration in humans - influence of materials: A study protocol for a randomised controlled trial and a pilot study results. Contemp Clin Trials Commun. 2020 Aug 15;19:100643.
- 7.Bagchi P, Alfawzan AA, Magar SM, Priya R, Kochhar AS, Agrawal S, AlMutairi FJ. An In vitro evaluation of effect of implant abutment on human gingival epithelial keratinocytes. Ann Afr Med. 2022 Jul-Sep;21(3):217-222.
- Brunello G, Brun P, Gardin C, Ferroni L, Bressan E, Meneghello R, Zavan B, Sivolella S. Biocompatibility and antibacterial properties of zirconium nitride coating on titanium abutments: An in vitro study. PLoS One. 2018 Jun 26;13(6):e0199591.
- 9. Trullenque-Eriksson, A.; Guisado-Moya, B. Retrospective long-term evaluation of dental implants in totally and partially edentulous patients.

www.jchr.org

JCHR (2024) 14(2), 3718-3725 | ISSN:2251-6727



- Part I: Survival and marginal bone loss. Implant. Dent. 2014, 23, 732–737.
- 10. Palmer, R.; Palmer, P.; Howe, L. Complications and maintenance. Br. Dent. J. 1999, 187, 653–658.
- 11. Petrie, C.S.; Williams, J.L. Comparative evaluation of implant designs: Influence of diameter, length, and taper on strains in the alveolar crest. A three-dimensional finite-element analysis. Clin. Oral Implant. Res. 2005, 16, 486–494.
- Vinhas, A.S.; Aroso, C.; Salazar, F.; Lopez-Jarana, P.; Rios-Santos, J.V.; Herrero-Climent, M. Review of the Mechanical Behavior of Different Implant-Abutment Connections. Int. J. Environ. Res. Public Health 2020, 17, 8685.
- 13. Sailer, I.; Zembic, A.; Jung, R.E.; Siegenthaler, D.; Holderegger, C.; Hammerle, C.H. Randomized controlled clinical trial of customized zirconia and titanium implant abutments for canine and posterior single-tooth implant reconstructions: Preliminary results at 1 year of function. Clin. Oral Implant. Res. 2009, 20, 219–225.
- 14. Zembic, A.; Sailer, I.; Jung, R.E.; Hammerle, C.H. Randomized-controlled clinical trial of customized zirconia and titanium implant abutments for singletooth implants in canine and posterior regions: 3-year results. Clin. Oral Implant. Res. 2009, 20, 802–808.
- 15. Lewis, S.G.; Llamas, D.; Avera, S. The UCLA abutment: A four-year review. J. Prosthet Dent. 1992, 67, 509–515.
- 16. Conejo J, Kobayashi T, Anadioti E, Blatz MB. Performance of CAD/CAM monolithic ceramic Implant-supported restorations bonded to titanium inserts: A systematic review. Eur J Oral Implantol. 2017;10 Suppl 1:139-146
- 17.Ikeda, H.; Yamaza, T.; Yoshinari, M.; Ohsaki, Y.; Ayukawa, Y.; Kido, M.A.; Inoue, T.; Shimono, M.; Koyano, K.; Tanaka, T. Ultrastructural and immunoelectron microscopic studies of the perimplant epithelium-implant (Ti-6Al-4V) interface of rat maxilla. J. Periodontol. 2000, 71, 961–97
- 18.Leonardi de Oliveira Rigotti R, Dias Corpa Tardelli J, Cândido Dos Reis A. Influence of antibacterial surface treatment on dental implants on cell viability: A systematic review. Heliyon. 2023 Feb 16;9(3):e13693.
- 19.Dias Corpa Tardelli J, Lima da Costa Valente M, Theodoro de Oliveira T, Cândido Dos Reis A.

- Influence of chemical composition on cell viability on titanium surfaces: A systematic review. J Prosthet Dent. 2021 Mar;125(3):421-425.
- 20.Park S., Kim H., Choi K.S., Ji M.K., Kim S., Gwon Y., Park C., Kim J., Lim H.P. Graphene-chitosan hybrid dental implants with enhanced antibacterial and cell-proliferation properties. Appl. Sci. 2020;10
- 21.Lindhe J., Meyle J. Peri-implant diseases: consensus report of the sixth European workshop on periodontology. J. Clin. Periodontol. 2008;35:282– 285. doi: 10.1111/j.1600-051X.2008.01283.x.
- 22.Calazans Neto JV, Kreve S, Valente MLDC, Reis ACD. Protein absorption on titanium surfaces treated with a high-power laser: A systematic review. J Prosthet Dent. 2022 Apr 10:S0022-3913(22)00178-0.
- 23. Van Oirschot BA, Meijer GJ, Bronkhorst EM, Närhi T, Jansen JA, van den Beucken JJ. Comparison of different surface modifications for titanium implants installed into the goat iliac crest. Clin Oral Implants Res. 2016 Feb;27(2):e57-67.
- 24.López-Valverde N, Flores-Fraile J, Ramírez JM, Sousa BM, Herrero-Hernández S, López-Valverde A. Bioactive Surfaces vs. Conventional Surfaces in Titanium Dental Implants: A Comparative Systematic Review. J Clin Med. 2020 Jun 29;9(7):2047.
- 25.Simões IG, Dos Reis AC, da Costa Valente ML. Analysis of the influence of surface treatment by high-power laser irradiation on the surface properties of titanium dental implants: A systematic review. J Prosthet Dent. 2023 Jun;129(6):863-870.
- 26.López-Valverde N, López-Valverde A, Aragoneses JM, Macedo de Sousa B, Rodrigues MJ, Ramírez JM. Systematic Review and Meta-Analysis of the Effectiveness of Calcium-Phosphate Coating on the Osseointegration of Titanium Implants. Materials (Basel). 2021 Jun 2;14(11):3015.
- 27.López-Valverde N, Macedo-de-Sousa B, López-Valverde A, Ramírez JM. Effectiveness of Antibacterial Surfaces in Osseointegration of Titanium Dental Implants: A Systematic Review. Antibiotics (Basel). 2021 Mar 28;10(4):360.
- 28. Wehner C, Lettner S, Moritz A, Andrukhov O, Rausch-Fan X. Effect of bisphosphonate treatment of titanium surfaces on alkaline phosphatase activity in osteoblasts: a systematic review and meta-analysis. BMC Oral Health. 2020 Apr 25;20(1):125.

www.jchr.org

JCHR (2024) 14(2), 3718-3725 | ISSN:2251-6727





Image 1 : Saliva Sample

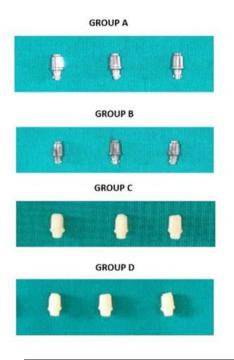


Image 2: Prosthetic Abutments



Image 3: Prosthetic Abutments immersed in saliva sample



Image 4: Incubation of samples in saliva

www.jchr.org JCHR (2024) 14(2), 3718-3725 | ISSN:2251-6727



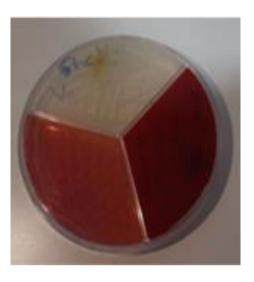
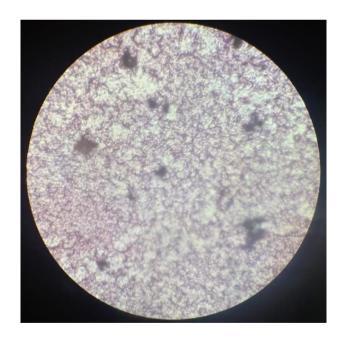


Image 5: Colonies formed after 72 hours of incubation



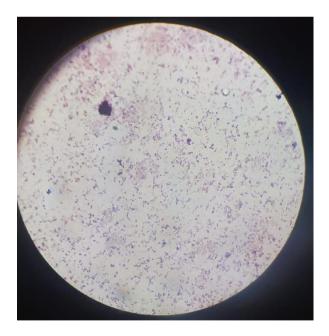


Image 6 : Microscopic view showing high mibrobes in cobalt chromium group and least in titanium group