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ORIGINAL ARTICLE

Oil pollution

Germination

Stem length

Vicia ervilia

Anatomy

Changes Germination, Growth and Anatomy Vicia ervilia in Response to Light Crude Oil Stress

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ABSTRACT: Thepetroleum concentrations are contaminant important sources in the environment. Hydrocarbon's contaminants depend on their type and concentration can cause variable toxicity in **KEYWORDS** soils, on the other hand, different kinds of plants also response concentration of contaminant differently, because of the diversity of physiological and morphological characteristics. In this study the effect of different concentrations of light crude oil was investigated on stem length, germination and anatomical of Vicia ervilia. A factorial experiment was used with a randomized complete block design (RCBD) with three replications. The results showed that the germination and stem lengths decreased significantly ($p \le 0.05$) with increasing concentrations of petroleum. Anatomical studies the changes of parenchyma, vascular bundles, epidermal and increase crack showed.

INTRODUCTION

Increasing industrial productions are along with increasing pollutant in the environment [1]. Petroleum pollutant is an unavoidable consequence of population increasing and industrialization process. Soil pollution petroleum concentrations is an by important environmental problem [2]. Petroleum hydrocarbons are harmful for the alive and no alive components due to toxicity effects [3]. Pollutant in soil can insert into food chain [4] and this is dangerous for the health of human and other animals [5]. Petroleum concentrations can affects on vegetation growth through stimulate the soil bacteria, consume soil nutrients and reduce soil oxygen.

Oil pollutions make disturbances in vegetations [6]. The signs of pollution in vegetation are numerous [6]. Petroleum cause chemically disturbances in metabolism and growth of vegetation [7], reduction of vegetation growth [8], autumn of areal organs (leaves), variations of pile, chlorophyll destruction, delay in flowering [9] and cell death [10]. In studies which were conducted by researchers, the performance of vegetation was studied in polluted soil by petroleum. The results showed that petroleum in some vegetations cause to decrease germination and growth (11, 3 and 12), delay or lack of germination [13, 14], increase root length and improve the vegetation growth in polluted soil [15, 16 and 17].

In previous studies, the performance of vegetation was studied in polluted soil by petroleum. The obtained results showed that petroleum in some vegetations cause the following effects: decrease of germination and growth (11, 3 and 12), delay or lack of germination [13, 14], increase of root length and improve the vegetation growth in polluted soil [15, 16 and 17]. Results of several researches showed that the petroleum concentrations produce different degree of toxicity according to the type and their concentration. Mentioned items show that the petroleum pollution caused disturbances that are morphological, physiological and anatomy variations in vegetations. The anatomic properties of vegetations are very important, but a little attention has been paid to the effects of petroleum on vegetation anatomy which have grown in polluted soil. Therefore, in this study the effects of different concentrations of light crude oilon germination, stem length and anatomy of Vicia ervilia was investigated in polluted soil.

MATERIALS AND METHODS

Soil was prepared from the adjacent agricultural lands. Soil was polluted by different concentration (1, 2, 3 and 4% W/W %) of petroleum and a soil without pollution was considered as control. Manual method was used to mix soil and petroleum [18]. Seeds with germination potential were planted in vessels contained by polluted and unpolluted soil at the depth of 1.5 cm. three replications were prepared from each concentration. All vessels were placed in free location with even temperature and environmental conditions. Control treatment was for the vegetation grown in unpolluted soil.

In this study the germination and stem height were measured. The germination rate in each vessel was calculated according to the number of planted seeds and following equation:

$$G = \frac{n}{N} \times 100$$

Where in this equation N is the number of planted seeds, n is the number of germinated seeds and G is the germination percentage [19]. The length of stem from the soil surface to top of the stem was measured using ruler in cm [20]. The lower part of stem was sampled for the anatomy experiments [10]. Vegetation texture was placed in solution of glycerin and ethanol 70%, because vegetation texture can be protected in these consolidators in long term [10]. The common blade was used to prepare sections from vegetation; the sample coloring was done using duplicated Carmen Stain. Prepared samples were assessed under the light Moticmicroscope and IXUS 220 HS canon camera was used to image samples. Excel was used to design diagrams and SPSS software was used to compare means by Duncan test at probability level of 0.05.

RESULTS

The results of this study showed that the existence of light crude oil at the different concentrations had effects on germination of plants and the germination percentage decreased with increasing petroleum in soil. At concentration of 4% it was observed a delay in germination. 1005 germination was detected in control treatment. Germination at the concentrations of 1, 2, 3 and 4% were 77.7, 33.3, 22.2 and 72.2%, respectively (Figure 1).



Figure 1. Germination of *Vicia ervilia* (Based on the counting of the number of germination per day)

Statistical analysis of data showed the significant difference among treatments. In comparison to control and contaminant treatments, it was not observed a significant difference between the control and treatments of 1 and 4% (Figure 2).



subscript shows that there isno significant difference at probability level of 5%)

The height of stem was measured every 5 days to determine the difference in growth variations. The plant in initial days had more height growth at concentration of 3% as compared to other contaminant concentrations. But at the end of growth period it showed significant reduction compared to treatment of 1 and 2% (Figure 3).



Figure 3. measuring stem height of *Vicia ervilia* (Every 5 day in growth period)

The results of the statistical analysis of the mean of stem height and investigation of the effects of concentration on stem height showed that there was significant difference among the treatments at probability level of 5%, but there was no significant difference among the control and treatments of 1 and 2% (Figure 4).



Figure 4.mean height of *Vicia ervilia*stem in cm(Columns with same subscript shows that there isno significant difference at probability level of 5%)

The stem of the vegetation grown in contaminant soil was different with the stem of vegetation grown in noncontaminant soil. These variations are shown in Figures 5 and 6.



Figure 5. Increasing of pile in contaminant sample compared to control (Left sample is control and right sample is contaminant sample) e: Epidermal, ph: Phloem, X: Xylem, co: Collenchyma, p: Parenchyma, Pi: pile



Figure 6. comparison of the cross section of Vicia ervilia and Variation in contaminant sample (right) and control (left) e: Epidermal, ph: Phloem, X: Xylem, co: Collenchyma, p: Parenchyma

Anatomical studies showed the changes of parenchyma, vascular bundles, epidermal and increase crack. As it was shown in Figure 5, control sample of *Vicia ervilia* has determined section and shape, in contaminant samples, it can be observed and an increase in swell and variations in cross section shape. In Figure 6 (down figure) control sample had large parenchyma cells and regular epidermal but in contaminant samples the size of Parenchyma cells decreased but its frequency increased. Moreover the irregularity in epidermal size can be observed. Phloem vascular of contaminant

samples was irregular and compacted as compared to control.

DISCUSSION

Germination is the most important vegetative stage of plants [21]. Germination rate was observed in all the concentrations of petroleum but the germination rate decreased with increasing petroleum concentration in soil which this was in agreement with the findings of Njoku and colleagues, and Kirk and colleagues [22, 23]. Decreasing of germination was happened due to reducing oxygen and moisture [24], poor accessibility nutrient for plant [25] and increasing toxicity materials of petroleum compositions in soil [26]. Delay in germination at 4% concentration was in agreement with the findings of Kisic and colleagues, which reported that petroleum compositions provided a soft layer around the seed and prevented from access to oxygen and water [27]. This caused delaying or lacking germination in polluted soil by oil. The mean of stem length of Vicia erviliain three replications from control to oil concentrations of 4% were respectively equal to 14.26, 13.69, 12.26, 10.03 and 6.53. The stem length was significantly decreased with increasing the concentrations of light crude oil (P<0.05). Reduction of vegetation growth by contaminant soil was proved by several researchers [28, 29 and 30]. Daryabeigizand and colleagues found that the light crude oil compounds hadn't significant negative effects on germination but it had significant effects during the vegetative stages [31]. This result showed that the oil by its toxicity effects prevented the natural growth of plant. Mendelssohn and Hester reported that the decreasing of stem length was due to physical, chemical and biological variations by petroleum compounds in soil [32]. Moreover reduction of stem length can be due to decreasing accessible nutrients for plants [33] or poor ventilation of soil which caused that the growth prevented and its length decreased [34]. According to the reports and results of statistical investigations of different concentrations of oil as compared to control showed that the toxicity compounds of light crude oil had negative effect on germination and growth of Vicia ervilia and the severity of effects increased with increasing the concentration of petroleum in soil. Species had degrees of sensitivity based on their morphological and physiological properties; therefore it is important to investigate the anatomical properties of plants. The observed variations were due to the effects of petroleum compounds in soil

grown plants. Vicia erviliais dicotyledonous species which have 3 epidermal layers, parenchyma and vascular bundles in the structure of stem. In contaminant treatment the pile increased. Increasing pile can be a tool of sourcing more petroleum compounds. Moreover an irregularity was observed in vascular bundles. Xylem and phloem vascular were compacted [35] and decreased by increasing the concentration of petroleum which this was not in agreement with the finding of Omosun and colleagues, [10]. The response of vegetation to decrease nutrients and nitrogen of soil reduced xylem and phloem vascular. Thus, reduction of vascular bundles led to decrease transporting petroleum into plant. Decreasing of the vegetation growth showed the disturbances in performance of xylem and phloem vascular in providing water and nutrient under the stress of soil contaminant by petroleum. This result was in agreement with the finding of Agbogidi and Eshegbeyi, and Agbogidi and Ofuoku [34, 36]. Increasing of parenchyma was in agreement with the findings of Gill and colleagues, [6]. This change has been occurred to produce stability in response to stress and/or preventing from the entering petroleum compounds in to cells. The cross section Vicia erviliahas changed. Nogueira and colleagues, in their research showed the changing of cross section [37]. Irregularity in epidermal was due to the disturbances in plant morphology. Epidermal thickness increased slowly which this was in agreement with the finding of Sharma and colleagues, and Gill and colleagues [38, 6]. They found the epidermal variations in vegetation grown in contaminant soil. Epidermal variations was occurred due to the preventing the reduction of inter tissue water [38]. In Table 1 observations of the present study are given compared to other studies.

The parameters Studied	Observations in this study	Correspondence	Nonconcurrence
Germination	Germination rate of was observed in all the concentrations of petroleum but the germination rate decreased with increasing petroleum concentration in soil.	Njoku <i>et al.</i> , 2011; Kirk <i>et al.</i> , 2002	_
Delaygermination	Delay in germination was observed at a concentration of 4 percent.	Kisic <i>et al.</i> , 2009	_
Averagestem length	The stem length was significantly decreased with increasing the concentrations of light crude oil (P<0.05).	Okonokhua <i>at el.</i> , 2007; Shahriari <i>et al.</i> , 2007; Ogboghodo <i>et al.</i> , 2003	_
Anatomical	The pile was increased in contaminant treatments.	_	_
	Irregularity was observed in vascular bundles.	_	_
	Xylem and phloem vascular were compacted by increasing the concentration of petroleum.	Kofidiset al., 2008	_
	Xylem and phloem vascular were decreased by increasing the concentration of petroleum.	_	Omosunet al., 2008
	Decreasing of the vegetation growth shows the disturbances in performance of xylem and phloem vascular in providing water and nutrient under the stress of soil contaminant by petroleum.	Gill et al., 1992	_
	Increasedparenchymaltissue.	Nogueiraet al., 2011	_
	The cross section Vicia ervilia has changed.	Sharma et al.,1980; Gill et al., 1992	_

Table 1. Observations of the present study comparison with other studies

CONCLUSIONS

In this study the effect of different concentrations of light crude oil was investigated on stem length, germination and anatomical of Vicia ervilia. The results showed that the oil had negative effect on germination and growth of Vicia ervilia. Germination and stem decreased significantly lengths with increasing concentrations of petroleum. The results of anatomical studies showed that the species changed its structure in response to petroleum contaminant which these variations were adaption mechanisms in response to contaminant location. Vicia ervilia can be generated in all petroleum concentrations and the growth continued in period, so it can be described that the plant had relative resistance against soil with low pollution. In recent years many countries which produce petroleum suggested Phytoremediation practices. In this process some plans can be generated in contaminant soil to petroleum.

REFERENCES

1. Uaboi-Egbenni P. O., Okolie P. N., Adejuyitan O. E., Sobande A.O., Akinyemi O., 2009. Effect of industrial effluents on the growth and anatomical structures of Abelmoschusesculentus (okra). African Journal of Biotechnology. 8(14): 3251-3260.

2. US EPA, 2000. Introduction to phytoremediation, EPA/600/R99/107, Environmental Protection Agency, USA.

3. Bamidele J.F., 2010. Threats to sustainable utilization of coastal wetlands in Nigeria. Journal of Nigerian Environmental Society. 5(3):217-225.

4. Morelli I. S., Del Panno M. T., De Antoni G. L., Painceira M. T., 2005. Laboratory study on the bioremediation of petrochemical sludge contaminated soil. International Bioremediation & Biodegradation. 55, 271-278.

5. Khan A.G., 2005. Role of soilmicrobes in the rhizospheres of plants growing on trace metal

contaminated soils in phytoremediation. Journal of Trace Elements in Medicine and Biology. 18, 355-364.

6. Gill L.S., Nyawuame H.G.K, Ehikhametalor A.O,
1992. Effect of crude oil on the growth andanatomical features of *Chromolaenaodorata* (L) K, e R. Newsletter.
6, 1-6.

7. Meudec A., Poupart N., Dussauze J., Deslandes E., 2007. Relationship between heavy fuel oil phytotoxicity and polycyclic aromatic hydrocarbon contamination in *Salicorniafragilis*. Journal of Science of the Total Environment. 381, 146-156.

8. Sharifi M., Sadeghi Y., Akbarpour M., 2007. Germination and growth of six plant species on contaminated soil with spent oil. International Journal of Environmental Science Technology. 4, 463-470.

9. Alkio M., Tabuchi T.M., Wang X., Colon Carmona A., 2005. Stress responses to polycyclic aromatic hydrocarbons in *Arabidopsis* include growth inhibition and hypersensitive response-like symptoms. Journal of Experimental Botany. 56(421): 2983–2994.

10. Omosun G., Markson A.A., Mbanasor O., 2008. Growth and anatomy of AmaranthusHybridus as affected by diferrent crude oil concentrations. AmericanEurasian Journal of Scientific Research. 3(1): 70-74.

11. Eze C.N., Maduka J.N., Ogbonna J.C., Eze E.A., 2013. Effects of Bonny light crude oil contamination on the germination, shoot growth and rhizobacterial flora of Vignaunguiculata and Arachis hypogea grown in sandy loam soil. Scientific Research and Essays. 8(2): 99 -107.

12. Lin Q., Mendelsshohn I. A., 2009. Potential of restoration and phytoremediation with Juncusroemerianus for diesel-contaminated coastal wetlands. Ecological Engineering. 35, 85-91.

13. Bona C., Mendonça de Rezende I., de Oliveira G.,2011. Antonio de Souza, L, Effect of Soil Contaminatedby Diesel Oil on the Germination of Seeds and theGrowth of Schinusterebinthifolius Raddi

(Anacardiaceae) .Seedlings An International journal. 54, 1379-1387.

 Kulakow P.A., Schwab A.P., Banks M.K., 2000.
 Screening plant species for growth in weathered petroleum hydrocarbon contaminated sediments.
 International Journal of Phytoremediation. 2, 297-317.

15. Ogbo E.M., 2009. Effect of diesel fuel contamination on seed germination of four crop plants-Arachis hypogea, Vignaunguiculata, Sorghum bicolor and Zea mays. Journal Biotechnology. 8(2): 250-253.

16. Njoku K.L., Akinola M.O., Taiwo B.G., 2009. Effect of gasoline diesel fuel mixture on the germination and the growth of *Vignaunguiculata* (Cowpea). African Journal of Environmental Science and Technology. 3(12): 466-471.

 Merkl N., Schultze-Kraft R., Infante C., 2005.
 Assessment of tropical grasses and legumes for phytoremediation of petroleumcontaminated soils.
 Water, air, soil pollution. 165, 195-209.

Nageswara Rao C.V., Afzal M., Malallah G., Kurian M., Gulshan S., 2007. Hydrocarbon Uptake by Roots of *Vicia faba* (Fabaceae). Environ Monit Assess. 132, 439–443.

 Sangabriel W., Ferrera Cerrato R., TrejoAguilar D., Mendonza Lopez M.R., Cruz Sanches J.S., Lopez Ortiz C., 2006. Internacional de ContaminacionAmbiental. 22, 63–73.

20. Vwioko D.E., Fashemi D.S., 2005. Growth response of Ricinuscommunis L (Castor Oil) in spent lubricating oil polluted soil. Journal of Applied Sciences & Environmental Management. 9(2): 73-79.

21. Banks M.K., Schultz K.E., 2005. Comparison of plants for germination toxicity tests in petroleumcontaminated soils. Water, Air, and Soil Pollution. 167, 211-219.

22. Njoku K.L., Akinola M.O., Oshodin O.R., 2011. Phytotoxicity Assay of Crude Oil Using Different Accessions of *Sorghum bicolor*. World Applied Sciences Journal. 15(1): 38-46. 23. Kirk J.L., Klironomos J.N., Lee H., Trevors J.T., 2002. Phytotoxicity assay to assess plant species for phytoremediation of petroleum contaminated soil. Bioremediation Journal. 6, 57-63.

24. Agbogidi O.M., Ejemete O.R., 2005. An assessment of the effects of crude oil pollution on soil properties, germination and growth of *Gambayaalbida* (L.). Uniswa. Journal of Agricultural Science Technology. 8(2): 148-155.

25. Nwadinigwe A.O., Uzodimma N.S., 2005. Effects of petroleum spills on the germination and growth of groundnut (*ArachishypogaeaL.*). Journal Biology res Biotechnology. 3(2): 101-105.

26. Kathi S., Khan B., 2011. Phytoremediation approaches to PAH contaminated soil. Indian Journal of Science and Technology. 4(1): 56-63.

27. Kisic I., Mesic M., Basic F., Brkic V., Mesic M., Durn G., 2009. The effect of drilling fl uids and crude oil on some chemical characteristics of soil and crops. Geoderma. 149, 209–216.

28. Okonokhua B.O., Ikhajiagbe B., Anoliefo G.O., Emede T., 2007. The Effects of Spent Engine Oil on Soil Properties and Growth of Maize (*Zea mays L.*). Journal of Applied Science and Environmental Management. 11(3): 147–152.

29. Shahriari M.H., Savaghebi Firoozabadi G., Azizi M., Kalantari F., Minai Tehrani D., 2007. Study of growth and germination of *Medicago sativa* (Alfalfa) in light crude oil contaminated soil. Research Journal of Agriculture and Biological Sciences. 3(1): 46–51.

30. Ogboghodo I.A., Iruaga E.K Osemwota I.O., Chokor J.U., 2003. An assessment of the effects of crude oil pollution on soil proprrties, germination and of maize (Zea *mays*) using two crude types- forcados light and escravos light, Nigeria.

Daryabeigizand A., Nabibidhendi G., Mehrdadi N.,
 2010. Phytoremediation of total petroleum hydrocarbons

(TPHs) using plant species in Iran. Turk Journal Agriculture (TUBİTAK). 34: 429-438.

32. Hester M.W., Mendelssohn I.A., 2000. Longterm recovery of a Louisiana brackish marsh plant community form oil spill impact: vegetation response and mitigation effects of marsh surface elevation. Marine Environmental Research. 49, 339-347.

33. Daniel-Kalio L.A., Pepple S.F., 2006. Effect of bonny light crude oil pollution of soil on the growth of dayflower (*CommelinabenghalensisL.*) in the Niger Delta, Nigeria. Journal of Applied Sciences & Environmental Management. 10 (2): 111-114.

34. Agbogidi O.M., Eshegbeyi O.F., 2006. Performance of Dacryodesedulis (Don. G. Lam. H.J.) Seeds and seedlings in a crude oil contaminated soil. Journal of Sustainable Forestry. 22 (3/4): 114.

35. Kofids G., Giannakoula A., Ilias I., 2008. Growth, anatomy and chlorophyll fluorescence of coriander plants (*Coriadrumsativum L.*) treated with prohexadionecalcium and daminozine. Acta Biologica Cracoviensia Series Botanica. 50(2): 55–62.

36. Agbogidi O.M., Ofuoku A.U., 2005. Response of sour sop (Annonamuricata Linn) to crude oil levels. Journal of Sustainable Tropical Agricultural Research. 16, 98-102.

37. Nogueira L., Inckot R., de Oliveira Santos G., de Souza L., Bona C., 2011. Phytotoxicity of petroleum-contaminated soil and Bioremediated soil on Allophylusedulis. Rodriguesia. 62(3): 459-466.

38. Sharma G.K., Chandler C., Salemi L., 1980. Environmental pollution and leaf cutivular variation in kadzu (PuererialobataWilld.). Annals of Botany. 45, 77-80.