www.jchr.org

JCHR (2023) 13(06), 3182-3195 | ISSN:2251-6727



## Advances in Plant-Isolated Indole Derivative-Based Niosomes: Applications in Drug Delivery and Cancer Treatment

Devinder Kumar Maheshwari <sup>1</sup>, S.Vigneswari <sup>2</sup>, Jameel Ahmed <sup>3</sup>, Sarakula Prasanthi <sup>4</sup>, Pampayya Thanthati <sup>5</sup>, Pranjali Bajrang Chole <sup>6</sup>, Abhisek Saha <sup>7</sup>, Mangala K J <sup>8</sup>

#### **Corresponding Author:**

Mangala K J, Assistant Professor, Government First Grade College, Ramanagara, Bm Road, Near Police Bhavana, Ramanagara District, Karnataka, India, Pin Code – 562159

(Received: 07 October 2023 Revised: 12 November Accepted: 06 December)

#### **KEYWORDS**

Niosomes, Indole derivatives, Drug delivery, Cancer treatment,

Targeted therapy

#### **ABSTRACT:**

Plant-isolated indole derivative-based niosomes represent a novel amalgamation of natural bioactive compounds and advanced drug delivery systems, holding immense promise in revolutionizing pharmaceutical interventions. These lipid-based vesicles, enriched with indole derivatives extracted from plants, offer a multifaceted approach to address critical challenges in drug delivery and cancer treatment. The integration of indole derivatives into niosomes enables targeted drug delivery, leveraging the compounds' inherent biological activities to achieve precision in therapeutic delivery while minimizing off-target effects. This targeted approach enhances drug efficacy and bioavailability while reducing systemic toxicity, marking a significant advancement in personalized medicine. In cancer treatment, these advanced delivery systems pave the way for transformative strategies by selectively delivering therapeutic agents to tumor sites, minimizing damage to healthy tissues, and potentially improving patient outcomes. The versatility of indole derivatives allows for tailored modifications of niosomes, optimizing drug stability, controlled release, and bioavailability. Moreover, the potential synergistic effects of these compounds with existing cancer treatments open avenues for combinatorial therapies and overcoming drug resistance. However, challenges persist in achieving reproducibility, scalability, and navigating regulatory pathways for clinical translation. Addressing these hurdles requires rigorous preclinical evaluations, establishing safety profiles, and optimizing formulation strategies. Despite these challenges, the convergence of plant-isolated indole derivative-based niosomes signifies a transformative

<sup>&</sup>lt;sup>1</sup>Assistant Professor, Guru Kashi University Talwandi Sabo Distt Bathinda Punjab, India, Pin Code -151302

<sup>&</sup>lt;sup>2</sup>Assistant Professor Of Chemistry, Seethalakshmi Achi College For Women, Pallathur-, Sivagangai Dt, India, Pin Code - 630107

<sup>&</sup>lt;sup>3</sup>Principal, Kandhar College Of Pharmacy, Kandhar, Dist Nanded

<sup>&</sup>lt;sup>4</sup>Assistant Professor, Department Of Chemistry, Adikavi Nannaya University, Tadepalligudem, Andhrapradesh, India, Pin Code -534101

<sup>&</sup>lt;sup>5</sup>Research Scholar-Full Time, Department Of Botany, Sri Krishnadevaraya University, Anantapuramu, Andhrapradesh, India, Pin Code -515003

<sup>&</sup>lt;sup>6</sup>Research Scholar, Christ (Deemed To Be University), Bangalore, Karnataka.

<sup>&</sup>lt;sup>7</sup>Associate Professor, Departmen In Chemistry, Tufanganj College (Affiliated To Cooch Behar Panchanan Barma University), Tufanganj, Cooch Behar, West Bengal, India, Pin Code-736160

<sup>&</sup>lt;sup>8</sup>Assistant Professor, Government First Grade College, Ramanagara, Bm Road, Near Police Bhavana, Ramanagara District, Karnataka, India, Pin Code – 562159

www.jchr.org

JCHR (2023) 13(06), 3182-3195 | ISSN:2251-6727



approach at the intersection of natural compounds and advanced drug delivery, offering tailored therapeutic solutions, enhanced drug efficacy, and improved patient care in the future of pharmaceutical interventions.

#### Introduction

Niosomes, lipid-based vesicles, have garnered substantial attention in pharmaceutical research due to their potential in drug delivery[1]. These microscopic structures, composed of non-ionic surfactants and cholesterol, resemble liposomes but offer advantages like enhanced stability and controlled drug release[2]. Their adaptability to encapsulate both hydrophilic and hydrophobic compounds makes them promising carriers for various therapeutics[3]. However, the use of niosomes has been further revolutionized by the integration of plant-isolated indole derivatives. Indole derivatives, extracted from natural sources like plants, exhibit diverse pharmacological properties, including anti-inflammatory, antimicrobial, and anticancer activities[4]. These compounds possess unique structural features that make them ideal candidates for pharmaceutical applications, particularly in enhancing drug delivery systems[5]. The integration of indole derivatives into niosomes has unlocked a new realm of possibilities in drug delivery and cancer treatment[6,7]. These derivatives serve as potent bioactive compounds capable of influencing the physicochemical properties of niosomes, thereby augmenting their therapeutic potential. Their inclusion not only enhances the stability and biocompatibility of niosomes but also contributes to targeted drug delivery, improved bioavailability, and reduced systemic toxicity[8]. The significance of indole derivative-based niosomes in drug delivery lies in their ability to overcome various challenges encountered in conventional drug administration. By utilizing these compounds, niosomes can selectively target specific cells or tissues, ensuring the efficient delivery of therapeutic agents[9]. This targeted approach minimizes off-target effects and maximizes drug efficacy, presenting a promising strategy for personalized medicine. Moreover, in the realm of cancer treatment, indole derivative-based niosomes have emerged as potential game-changers. The ability to encapsulate chemotherapeutic agents within these vesicles allows for precise delivery to cancer cells while sparing healthy tissues[10]. This targeted delivery minimizes adverse

effects and improves the therapeutic index of anticancer drugs, thereby potentially revolutionizing the landscape of cancer therapy[11]. The amalgamation of plantisolated indole derivatives with niosomes represents a convergence of natural bioactive compounds and advanced drug delivery systems[12]. This synergy not only harnesses the therapeutic potential of indole derivatives but also leverages the strengths of niosomes, creating a platform with multifaceted benefits for drug delivery and cancer treatment[13]. In essence, the incorporation of plant-isolated indole derivatives into niosomes marks a significant advancement in pharmaceutical research. It opens doors to innovative strategies for drug delivery, emphasizing precision, efficacy, and reduced toxicity[14]. The potential applications in cancer treatment hold promise for improving patient outcomes and transforming the landscape of oncology. As research progresses, the exploration of these indole derivative-based niosomes is expected to unveil further opportunities for addressing unmet medical needs and shaping the future of therapeutics[15].

#### Indole derivatives, compounds

Indole derivatives, compounds derived from the heterocyclic structure of indole, possess diverse chemical structures and pharmacological properties[16]. The indole ring, consisting of a benzene ring fused to a pyrrole ring, forms the core structure of these derivatives. Their versatile nature makes them integral in various biological processes and pharmaceutical applications[17].

#### **Characteristics of Indole Derivatives**

Indole derivatives are a class of organic compounds characterized by the presence of the indole moiety in their structure. This molecular arrangement imparts unique physicochemical and biological properties to these compounds. Indole derivatives can vary significantly in their substitution patterns, leading to a wide array of compounds with distinct properties and

www.jchr.org

JCHR (2023) 13(06), 3182-3195 | ISSN:2251-6727



functions[18]. The indole structure is inherently aromatic and serves as a scaffold for synthesizing compounds with diverse functionalities. Modifications to the indole nucleus result in alterations in the compound's solubility, reactivity, and pharmacological activities[19]. Their structural versatility enables the development of compounds with targeted biological activities, making them attractive candidates for pharmaceutical applications[20].

#### **Sources of Plant-Isolated Indole Derivatives**

Plant sources serve as abundant reservoirs of indole derivatives. Various plants naturally produce these compounds, which serve essential roles in plant defense signaling pathways, and mechanisms, regulation[21]. Examples of plants known to contain indole derivatives include Catharanthus roseus (source of vinblastine and vincristine used in cancer treatment), Brassica oleracea (rich in indole-3-carbinol), and Cinchona officinalis (source of quinine)[22]. Extraction and isolation techniques allow for the procurement of these compounds from plants. Bioactive indole derivatives are obtained through processes like solvent extraction, chromatography, and isolation from specific plant parts such as leaves, roots, or seeds[23]. These natural sources provide a sustainable and diverse repertoire of indole derivatives for pharmaceutical exploration[24].

# **Properties Contributing to Their Role in Drug Delivery**

The multifaceted properties of indole derivatives render them pivotal in drug delivery systems. These compounds boast inherent biological activities encompassing antimicrobial, anti-inflammatory, and anticancer effects, augmenting the therapeutic potential of medications upon their integration into drug delivery platforms[25,26]. The structural diversity inherent in indole derivatives enables the creation of a vast spectrum of compounds with varying pharmacological activities, facilitating tailored design of drug delivery systems to suit specific therapeutic requirements[27]. Their favorable biocompatibility and low toxicity profiles make indole derivatives prime candidates for integration into drug delivery systems intended for

biological applications. The presence of functional groups on the indole nucleus allows for facile modification, empowering the design of derivatives tailored to exhibit desired properties, such as enhanced solubility, targeted delivery, and controlled release[28]. Moreover, certain indole derivatives showcase an affinity for specific receptors or biological targets, empowering targeted drug delivery, thereby heightening the selectivity and efficacy of drug delivery systems[29]. Incorporating plant-isolated indole derivatives into drug delivery systems like niosomes capitalizes on these properties, elevating performance and functionalities of these delivery vehicles[30]. These derivatives function as bioactive components, endowing specific functionalities to the delivery systems, potentially amplifying drug stability, bioavailability, and targeted delivery to designated sites within the body[31]. In summation, the manifold characteristics and diverse sources of indole derivatives designate them as indispensable components in drug delivery systems. Their chemical adaptability, biological activities, and compatibility with biological systems position them as promising contenders for enhancing the precision and effectiveness of drug delivery mechanisms, fostering the advent of innovative therapeutic interventions[32,33].

#### **Niosomes**

Niosomes represent lipid-based vesicular systems composed of non-ionic surfactants, cholesterol, and sometimes other additives[33]. These vesicles self-assemble into bilayer structures, akin to liposomes, but with the key distinction of being non-ionic. The amphiphilic nature of the surfactants used in niosome formation allows them to encapsulate both hydrophilic and hydrophobic drugs within their aqueous core or lipid bilayers, providing a versatile drug delivery platform[34]. The structural composition of niosomes, comprising surfactants and cholesterol in varying ratios, influences their size, shape, and membrane properties, affecting their performance as drug carriers[35].

# A. Advantages over other drug delivery systems

Niosomes, distinct lipid-based vesicular systems composed of non-ionic surfactants and cholesterol, present a myriad of advantages over conventional drug

www.jchr.org

JCHR (2023) 13(06), 3182-3195 | ISSN:2251-6727



delivery systems[36]. Their non-ionic nature offers enhanced stability compared to ionic counterparts, mitigating issues related to aggregation and fusion, thereby ensuring improved shelf life and sustained drug release[37] The versatility of niosomes to encapsulate a wide range of drugs, irrespective of their solubility (hydrophilic or hydrophobic), expands their utility and applicability in pharmaceutical formulations. This capability not only broadens the spectrum of drugs that can be delivered but also facilitates simultaneous delivery of multiple agents[38]. Their biocompatibility and low production costs make them attractive for large-scale manufacturing, potentially reducing the overall cost of pharmaceutical products[39]. Additionally, the adaptability of niosomal membrane composition allows tailored modifications, enabling targeted delivery, controlled release, and improved bioavailability of encapsulated drugs[40]. The ability to fine-tune niosome properties, including size, surface charge, and membrane permeability, further accentuates their advantages, paving the way for customizable drug delivery systems that cater to specific therapeutic requirements[41].

# B. Challenges and limitations in niosomal drug delivery

Navigating the landscape of niosomal drug delivery reveals a panorama of challenges and limitations that underscore the complexity and evolution of these lipidbased vesicular systems[42]. One primary challenge lies achieving consistent reproducibility and standardization during manufacturing, given the batch-to-batch variability inherent niosome characteristics like size, shape, and encapsulation efficiency[43]. This variability can impede uniform drug delivery and compromise therapeutic outcomes. Stability concerns also loom large, as niosomes are prone to aggregation, size alterations, and drug leakage during prolonged storage, potentially affecting their efficacy and safety upon administration[44]. Contending with these stability issues is pivotal for ensuring reliable and long-term storage of niosomal formulations[45]. Moreover, the delicate balance between achieving controlled drug release and preventing premature or burst release poses a significant hurdle[46]. Tailoring niosomes to achieve precise kinetics in drug release, especially for drugs with diverse physicochemical properties, remains a challenge vet to be fully conquered[47]. Addressing these challenges necessitates innovations in formulation strategies, manufacturing techniques, and stabilization methods to enhance the reproducibility, stability, and control of niosomal drug delivery systems[48]. Another critical aspect involves the potential toxicity associated with certain surfactants used in niosome formulation, necessitating thorough toxicity assessments and the exploration of alternative, biocompatible materials [49]. Additionally, the intricate interplay between niosome properties and biological barriers within the body presents complexities in achieving targeted and efficient delivery. Overcoming these multifaceted challenges requires concerted efforts in research and development, aiming to refine formulation techniques, enhance stability profiles, and optimize drug release kinetics while ensuring safety and efficacy, thereby paving the way for the broader clinical translation of niosomal drug delivery systems[50].

# Synthesis and Characterization of Indole-Derivative Based Niosomes

# A. Methods of Incorporating Indole Derivatives into Niosomes

Integrating indole derivatives into niosomes involves diverse methodologies aiming to exploit the beneficial properties of these compounds within the vesicular structure[51]. One common approach is the coacervation method, where indole derivatives are coformulated with non-ionic surfactants and cholesterol, followed by vesicle formation through hydration. This method enables the encapsulation of indole derivatives within the niosomal bilayers or the aqueous core, depending on the derivative's solubility[52]. Another strategy involves the post-insertion technique, where pre-formed niosomes are incubated with indole derivatives, allowing for their insertion into the vesicular membrane. This approach offers precise control over the indole derivative concentration within the niosomes[53]. Furthermore, the thin-film hydration method, widely used for niosome preparation, can be modified to encapsulate indole derivatives by incorporating these compounds into the lipid film during vesicle formation. The choice of method depends on factors like the physicochemical properties of the indole derivative, desired encapsulation efficiency, and

www.jchr.org

JCHR (2023) 13(06), 3182-3195 | ISSN:2251-6727



the intended application of the indole-loaded niosomes[54].

# **B.** Characterization Techniques Used for Assessing Indole Derivative-Based Niosomes

Assessing the characteristics and performance of indole derivative-based niosomes necessitates a comprehensive array of characterization techniques. Morphological analysis using techniques such as Transmission Electron Microscopy (TEM) and Scanning Electron Microscopy (SEM) provides insights into the shape, size distribution, and surface morphology of the niosomes loaded with indole derivatives[55,56]. Dynamic Light Scattering (DLS) offers information on particle size distribution and polydispersity index, crucial for evaluating the uniformity and stability of these vesicles. Fourier Transform Infrared Spectroscopy (FTIR) aids in identifying chemical interactions between indole derivatives and niosomal components, elucidating their incorporation within the vesicles[57]. Differential Scanning Calorimetry (DSC) and X-ray Diffraction (XRD) provide data on the physical state and crystallinity of indole derivatives within the niosomal bilayers. Moreover, techniques like Entrapment Efficiency (EE) determination and Drug Release Studies enable quantification of the encapsulated indole derivatives and evaluation of their release kinetics from the niosomal carriers, respectively [58,59].

# C. Factors Influencing the Properties of These Niosomes

Several key factors intricately influence the properties and performance of indole derivative-loaded niosomes. The choice of non-ionic surfactants and their ratios significantly impacts the vesicle size, morphology, and encapsulation efficiency of indole derivatives [60]. The nature and concentration of cholesterol, an essential component in niosomal formulations, play a pivotal role in membrane fluidity and stability, thereby affecting the entrapment and release of indole derivatives[61]. The physicochemical properties of indole derivatives, including their solubility, molecular weight, and hydrophobicity, dictate their distribution within the niosomal bilayers and influence the overall stability of the vesicles[62]. The preparation method employed for niosome synthesis, whether it involves coacervation, thin-film hydration, or post-insertion, influences the loading efficiency and distribution of indole derivatives within the vesicles[63]. Environmental factors such as temperature, pH, and ionic strength during niosome preparation can also impact the properties of indole derivative-loaded niosomes, highlighting the need for meticulous optimization to attain desirable characteristics for specific applications. Understanding and manipulating these factors are critical for tailoring the properties of indole derivative-based niosomes to achieve optimal drug delivery performance[64].

# Applications in Drug Delivery A. Targeted Drug Delivery Using Indole DerivativeBased Niosomes

The integration of indole derivatives into niosomes has unlocked a realm of possibilities for targeted drug delivery, revolutionizing therapeutic precision[65]. Indole derivative-based niosomes offer a strategic approach to navigate the complexities of targeted drug delivery. These systems capitalize on the inherent biological activities and receptor affinity of indole derivatives to achieve site-specific drug delivery[66]. By functionalizing the niosomal surface or modifying the vesicular membrane with ligands or targeting indole derivative-based niosomes moieties, selectively recognize and bind to specific receptors or cellular markers overexpressed in diseased tissues[67]. This targeted approach facilitates the accumulation of drugs at the intended site while minimizing exposure to healthy tissues, reducing off-target effects, and enhancing therapeutic efficacy. Indole derivatives, acting as guiding elements, direct niosomes loaded with therapeutic agents to disease sites, ranging from cancer cells to inflamed tissues, offering a promising avenue for personalized medicine and tailored drug delivery strategies[68].

### B. Enhanced Bioavailability and Sustained Release

Indole derivative-based niosomes have emerged as facilitators of enhanced bioavailability and sustained drug release, addressing challenges encountered with conventional drug formulations. The encapsulation of drugs within niosomes, facilitated by indole derivatives, shields them from premature degradation metabolism, thereby improving their bioavailability[69,70]. The bilayer structure

www.jchr.org

JCHR (2023) 13(06), 3182-3195 | ISSN:2251-6727



niosomes, aided by the presence of indole derivatives, provides a protective environment that extends drug circulation time, leading to prolonged systemic exposure and increased therapeutic efficacy[71]. Additionally, the controlled and sustained release properties of indole derivative-based niosomes play a pivotal role in optimizing drug delivery kinetics. These vesicular systems enable the modulation of drug release rates, allowing for a sustained and controlled release profile over an extended period. The controlled release not only ensures a consistent therapeutic effect but also minimizes fluctuations in drug plasma levels, thereby improving patient compliance and reducing dosing frequency, marking a significant advancement in pharmaceutical formulation design[72,73].

# **C.** Overcoming Challenges in Delivering Specific Therapeutics

Indole derivative-based niosomes serve as robust platforms for overcoming challenges associated with delivering specific therapeutics, particularly those with inherent limitations in solubility, stability, or targeting requirements[74]. These versatile delivery systems excel in encapsulating diverse classes of drugs, including hydrophobic, hydrophilic, and poorly soluble compounds. Indole derivatives aid in improving the solubility and stability of these challenging drugs within the niosomal structure, thereby enhancing their delivery potential[75]. Furthermore, indole derivative-based niosomes facilitate the co-delivery of multiple drugs or combination therapies, addressing complexities in administering synergistic drug combinations enhanced therapeutic outcomes. By encapsulating and co-delivering drugs with different physicochemical properties, these niosomes enable precise control over drug ratios and release kinetics, ensuring synergistic effects and drug minimizing potential interactions[76,77]. Additionally, these delivery systems offer avenues for delivering biologics, nucleic acids, or gene-based therapies, overcoming the challenges associated with their administration and ensuring their targeted and efficient delivery to specific cells or tissues, heralding new horizons in precision medicine[78].

#### **Role in Cancer Treatment**

# A. Indole Derivative-Based Niosomes in Chemotherapy

The integration of indole derivative-based niosomes holds immense promise in reshaping chemotherapy paradigms[79]. These innovative delivery systems present a strategic avenue to optimize the efficacy of chemotherapeutic agents while mitigating their adverse effects. Indole derivatives play a pivotal role in enhancing the therapeutic index of chemotherapeutic drugs by encapsulating them within niosomes[80]. The encapsulation shields the drugs from premature degradation, ensuring their stability and preserving their pharmacological activity. Moreover, indole derivatives may potentiate the cytotoxic effects of chemotherapy drugs, enhancing their anti-cancer properties[81]. The controlled and targeted delivery of chemotherapeutic agents using indole derivative-based niosomes enables higher drug concentrations at tumor sites, augmenting their efficacy against cancer cells while minimizing exposure to healthy tissues, thus reducing systemic toxicity. This targeted approach, facilitated by indole derivatives, redefines the landscape of chemotherapy, offering the potential for increased treatment efficacy and reduced side effects, ultimately improving patient outcomes[82].

# B. Targeted Delivery to Cancer Cells and Reduced Systemic Toxicity

Indole derivative-based niosomes exhibit remarkable capabilities in achieving targeted drug delivery to cancer cells, a critical aspect in minimizing systemic associated with conventional toxicity chemotherapy[83]. Leveraging the inherent properties of indole derivatives, these niosomes can be engineered recognize specific biomarkers or receptors overexpressed on cancer cells. By functionalizing the niosomal surface or modifying the vesicular membrane with targeting ligands, indole derivative-based niosomes achieve selective recognition and binding to cancerous tissues while evading healthy cells[84,85]. This targeted delivery strategy ensures preferential accumulation of chemotherapeutic agents within tumor maximizing their concentration precisely where needed, thus enhancing their anti-cancer effects. Consequently, the reduced exposure of healthy tissues to potent cytotoxic drugs minimizes off-target effects and

www.jchr.org

JCHR (2023) 13(06), 3182-3195 | ISSN:2251-6727



systemic toxicity, ameliorating the debilitating side effects commonly associated with traditional chemotherapy[86]. The ability of indole derivative-based niosomes to navigate the complexities of cancer heterogeneity and target specific cell populations heralds a paradigm shift towards safer and more effective cancer treatments.

# C. Potential Synergistic Effects with Existing Cancer Treatments

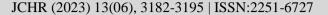
Indole derivative-based niosomes present an avenue for exploring synergistic effects when combined with existing cancer treatments, fostering a comprehensive approach to combat malignancies[87]. These versatile delivery systems offer the possibility of coencapsulating multiple therapeutic agents within the niosomal structure, including chemotherapeutic drugs, targeted therapies, or even immunotherapeutic agents. Indole derivatives aid in optimizing the co-delivery of

these agents, potentially eliciting synergistic effects that enhance therapeutic outcomes[88]. The controlled release kinetics and tailored delivery of drug combinations facilitated by indole derivative-based niosomes allow for precise modulation of drug ratios and schedules, maximizing the synergistic potential between different treatment modalities. Furthermore, these niosomes may potentiate the effectiveness of existing cancer treatments by overcoming drug resistance mechanisms or enhancing the cellular uptake of therapeutics[89,90]. The integration of indole derivatives into niosomal delivery platforms offers a promising avenue to explore synergistic interactions among diverse treatment modalities, heralding a new era of combinatorial cancer therapies aimed at improving patient responses and overcoming treatment resistance[91]. Anticancer drug-loaded formulations, in-vivo models, their characterization, and the outcome shown in table 1.

Table 1. Anticancer drug-loaded niosomal formulations, in-vivo models, their characterization, and the outcome.

Study	Characterization	Outcome	Route of	Ref
			Administration	
Co-delivery of	Size: $260.37 \pm 6.58$	Enhancing the stability of	Orally and	[19]
hydrophobic natural	nm, PDI: $0.42 \pm 0.03$ ,	curcumin and QC and their	Subcutaneous	
products	Zeta potential: -34.97	pharmacological efficacy.	injection	
	± 1.5 mv, EE	Better anti-inflammatory		
	Curcumin: 98.85 ±	impact		
	0.55 %, EE QC: 93.13			
	± 1.22 %			
Synthesized and	Size: 278 ± 5 nm, EE:	A significant antitumor effect	Injected	[17]
characterization of	$87 \pm 3 \%$ , Zeta	of WA-niosomes in-vivo was	intraperitoneally	
anticancer niosomal	Potential: -41.72 ±	discovered as a prototype of	into the flanks of	
withaferin-A	6.01 mV, PDI: 0.419 ±	cancer	the test animal	
formulation	0.073			
Curcumin entrapped	Size: 249.83 ± 6.38	The anti-inflammatory effect	Orally and	[20]
hyaluronan containing	nm, EE: $98.28 \pm 0.278$	of the hyaluronan containing	subcutaneous	
niosomes	% (w/w), PDI: 0.36 ±	niosomes was higher than free	injection	
	0.04, Zeta potential:	curcumin		
	$-34.83 \pm 0.5 \text{ mv}$			
Treatment of breast	Size: $325.5 \pm 9.53$ nm,	Significant antitumor effect	Intra-tumour	[19]
cancer with	EE: 91.18 ± 0.72 %	shown compared to TMX	injection	
engineered novel pH-	with slow drug release			
sensitive Triaryl-(Z)-	of 45.41 ± 1.20 %			
olefin niosomes	within 8 h.			

www.jchr.org





containing hydrogel				
Smart stimuli- responsive biofunctionalized niosomal nanocarriers for programmed release of bioactive compounds into cancer cells	Size: 163.27 nm, Zeta potential: -0.71 mV	Delivery to cancer models caused a higher tumor inhibition rate than in other groups.	Subcutaneous injection	[19]
Delivery of vinblastine-containing niosomes resulted in potent cytotoxicity on tumor cells	Size: 234.3 ± 11.4 nm, Zeta potential: -34.6 ± 4.2 mV, EE: 99.92 ± 1.6 %	In animal model, PnVB exhibited stronger tumor inhibitory effect and longer life time in comparison to free VB	Administered intravenously (tail vein) and inoculated subcutaneously into the right flank	[20]
Paclitaxel (PTX)- loaded pH-responsive niosomes modified with ergosterol were developed	Size: 240 nm, EE: 77 %	Encapsulating PTX in niosomal formulation developed its therapeutic efficacy	Intraperitoneally injected	[18]
Delivery of melittin- loaded niosomes for breast cancer treatment	Size: 121.4 nm, PDI: 0.211, EE: 79.32 %	Melittin-loaded niosome enhanced targeting, encapsulation efficiency, PDI, and release rate and shows a high anticancer effect on cell lines	Intraperitoneally injected	[16]
Prepared and characterized in-vitro and in-vivo niosomal formulation loaded with Galangin on chemically induced hepatocellular carcinoma	Size: 173.7–355.6 nm, EE: 45.13 %–77.69 %, Drug loading capacity (DL%): 9.02 %–16.72 %	Histopathological and immunohistochemical examinations revealed that GAL-loaded niosomes allowed a meaningful decrease in MCM3 immunostained hepatocytes and liver tumor lesions with few liver adenomas	Subcutaneous injection	[14]

# Future Perspectives and Challenges A. Emerging Trends and Future Directions in This Field

The amalgamation of indole derivative-based niosomes in drug delivery heralds a future brimming with innovation and transformative trends[92]. One emerging trend involves the refinement and diversification of indole derivatives to tailor their properties for enhanced compatibility with niosomal carriers[93]. Researchers are exploring chemical modifications and structural

optimizations of indole derivatives to fine-tune their interactions within niosomal structures, optimizing drug loading, stability, and release kinetics[94]. Additionally, the integration of advanced nanotechnology and nanomedicine principles into the design of indole derivative-based niosomes opens avenues for engineering sophisticated delivery systems[95]. These include the development of stimuli-responsive niosomes that can trigger drug release in response to specific environmental cues within the body, leading to ondemand drug delivery. Furthermore, the exploration of

www.jchr.org

JCHR (2023) 13(06), 3182-3195 | ISSN:2251-6727



multifunctionalized niosomes, combining targeting ligands, imaging agents, and therapeutic payloads, paves the way for theranostic applications, enabling simultaneous diagnosis and treatment[96]. The convergence of these trends positions the field for significant strides in precision medicine, personalized therapeutics, and innovative drug delivery systems.

# B. Unexplored Potential and Areas for Further Research

Despite significant advancements, the realm of indole derivative-based niosomes harbors unexplored territories ripe for further exploration and scientific inquiry[97]. One intriguing avenue lies in delving deeper into the synergistic interactions between indole derivatives and niosomes to harness their combined potential for diverse applications. Exploring the biophysical interactions between indole derivatives and niosomal components at the molecular level could unveil novel insights into formulation design and optimization[98]. Moreover, expanding the scope of indole derivatives to encompass a broader range of naturally occurring compounds or synthetically modified derivatives presents an intriguing area for exploration[99]. Investigating the pharmacokinetics and pharmacodynamics of indole-loaded niosomes in intricate biological systems, such as animal models or human trials, is pivotal for comprehensive understanding and clinical translation[100]. Additionally, exploring the potential of these advanced delivery systems in addressing challenges associated with drug resistance mechanisms in various diseases, especially in cancer, opens new vistas for therapeutic interventions. Embracing these uncharted realms through interdisciplinary collaborations and innovative research methodologies holds the key to unlocking the full potential of indole derivative-based niosomes[101]

# C. Addressing Limitations and Challenges for Clinical Translation

The journey from laboratory innovation to clinical application demands concerted efforts to overcome challenges and address limitations inherent in indole derivative-based niosomes. One pressing challenge is ensuring the reproducibility and scalability of niosome manufacturing processes to meet regulatory standards

and facilitate clinical translation[99]. Developing standardized protocols and robust quality control measures to ensure consistency in niosome formulation and performance is crucial. Moreover, bridging the gap between benchside innovation and bedside application requires comprehensive preclinical studies to elucidate safety profiles, toxicity, and pharmacokinetics[50]. Addressing concerns regarding long-term stability, storage conditions, and shelf-life stability of indole derivative-based niosomes is paramount for their viability[14]. Furthermore. clinical navigating regulatory pathways and securing approval for these advanced delivery systems necessitates aligning with regulatory frameworks and demonstrating their safety and efficacy through rigorous clinical trials[10]. Collaboration between academia, industry, regulatory bodies is essential to surmounting these challenges and expediting the translation of indole derivative-based niosomes from experimental concepts to clinically viable therapeutic solutions[16].

#### Conclusion

Plant-isolated indole derivative-based represent a groundbreaking convergence of natural bioactive compounds and advanced drug delivery systems. These innovative vesicular carriers, fortified by the inclusion of indole derivatives, hold immense promise in revolutionizing drug delivery and cancer treatment landscapes. Their significance lies in offering multifaceted solutions to critical challenges encountered in conventional drug delivery. By leveraging the inherent biological activities and versatile properties of indole derivatives, these niosomes enable targeted drug delivery, enhancing therapeutic precision and efficacy while minimizing off-target effects. The potential impact of these advanced delivery systems transcends boundaries, envisaging a paradigm shift in drug delivery by enhancing bioavailability, achieving sustained release, and overcoming limitations associated with specific therapeutics. In the realm of cancer treatment, indole derivative-based niosomes emerge as gamechangers, orchestrating targeted delivery to cancer cells while reducing systemic toxicity, thus promising improved patient outcomes. The amalgamation of these compounds in niosomal carriers holds the potential to usher in synergistic effects with existing cancer treatments, amplifying therapeutic outcomes and

#### www.jchr.org

JCHR (2023) 13(06), 3182-3195 | ISSN:2251-6727



addressing drug resistance. As these advancements continue to unfold, their translation from bench to bedside demands strategic focus on overcoming challenges related to reproducibility, scalability, and regulatory pathways. The journey towards clinical translation necessitates comprehensive preclinical evaluations to establish safety profiles pharmacokinetics while navigating regulatory frameworks for eventual approval. In closing, the convergence of plant-isolated indole derivative-based niosomes epitomizes innovation at the intersection of nature-inspired compounds and cutting-edge delivery systems, poised to redefine the future of drug delivery and cancer therapeutics, promising tailored treatments, improved efficacy, and enhanced patient care.

#### References

- [1] Moghimi, S.M.; Kissel, T.J. Particulate nanomedicines. Adv. Drug Deliv. Rev. 2006, 58(14), 1451–1455.
- [2] Miller, R.P.; et al. Mechanisms of cisplatin nephrotoxicity. Toxins 2010, 2(11), 2490–2518.
- [3] Grigorian, A.; O'Brien, C.B. Hepatotoxicity secondary to chemotherapy. J. Clin. Transl. Hepatol. 2014, 2(2), 95.
- [4] Omoti, A.E.; Omoti, C.E. Ocular toxicity of systemic anticancer chemotherapy. Pharm. Pract. 2006, 4(2), 55.
- [5] Lindenberg, M.; et al. Classification of orally administered drugs on the World Health Organization Model list of Essential Medicines according to the biopharmaceutics classification system. Eur. J. Pharm. BioPharm. 2004, 58(2), 265–278.
- [6] Hauss, D.J. Oral Lipid-Based Formulations: Enhancing the Bioavailability of Poorly Water-Soluble Drugs; CRC Press: Boca Raton, 2007.
- [7] Naseroleslami, M.; et al. Simvastatin-loaded nanoniosomes confer cardioprotection against myocardial ischemia/reperfusion injury. Drug Deliv. Transl. Res. 2022, 12(6), 1423–1432.
- [8] Barani, M.; et al. A new formulation of hydrophobin-coated niosome as a drug carrier to cancer cells. Mater. Sci. Eng. C 2020, 113, 110975.
- [9] Choi, Y.H.; Han, H.-K.J. Nanomedicines: current status and future perspectives in aspect of drug

- delivery and pharmacokinetics. J. Pharm. Investig. 2018, 48(1), 43–60.
- [10] Rajaee Behbahani, S.; et al. Red elemental nanoparticles mediated substantial selenium in differentiation. variations growth, tissue metabolism, gene transcription, epigenetic cytosine DNA methylation, and callogenesis in bittermelon (Momordica charantia); an in vitro experiment. PLoS ONE 2020, 15(7), e0235556.
- [11] Sotoodehnia-Korani, S.; et al. Selenium nanoparticles induced variations in growth, morphology, anatomy, biochemistry, gene expression, and epigenetic DNA methylation in Capsicum annuum; an in vitro study. Environ. Pollut. 2020, 265, 114727.
- [12] Bourbour, M.; et al. Evaluation of anti-cancer and anti-metastatic effects of folate-PEGylated niosomes for co-delivery of letrozole and ascorbic acid on breast cancer cells. Mol. Syst. Des. Eng. 2022, 7, 1102.
- [13] Akbarzadeh, I.; et al. The optimized formulation of tamoxifen-loaded niosomes efficiently induced apoptosis and cell cycle arrest in breast cancer cells. AAPS PharmSciTech. 2022, 23(1), 1–13.
- [14] Mahale, N.; et al. Niosomes: novel sustained release nonionic stable vesicular systems—an overview. Adv. Colloid Interface Sci. 2012, 183, 46–54.
- [15] Rai, A.; et al. Niosomes: an approach to current drug delivery-A review. Int. J. Adv. Pharm. 2017, 6(2), 41–48.
- [16] Moghanloo, M.; et al. Differential physiology and expression of phenylalanine ammonia lyase (PAL) and universal stress protein (USP) in the endangered species Astragalus fridae following seed priming with cold plasma and manipulation of culture medium with silica nanoparticles. 3 Biotech 2019, 9(7), 1–13
- [17] Sahrayi, H.; et al. Co-delivery of letrozole and cyclophosphamide via folic acid-decorated nanoniosomes for breast cancer therapy: synergic effect, augmentation of cytotoxicity, and apoptosis gene expression. Pharmaceuticals 2021, 15(1), 6.
- [18] Abarzadeh, I.; et al. Development of a novel niosomal formulation for Gabapentin. Iran J Colorectal Res. 2021, 9(4), 149–157.
- [19] Akbarzadeh, I.; et al. Folic acid-functionalized niosomal nanoparticles for selective dual-drug

#### www.jchr.org



- delivery into breast cancer cells: an in-vitro investigation. Adv. Powder Technol. 2020, 31(9), 4064–4071.
- [20] Akbarzadeh, I.; et al. Optimization, physicochemical characterization, and antimicrobial activity of a novel simvastatin nanoniosomal gel against E. coli and S. aureus. Chem. Phys. Lipids. 2021, 234, 105019.
- [21] Moghanloo, M.; et al. Seed priming with cold plasma and supplementation of culture medium with silicon nanoparticle modified growth, physiology, and anatomy in Astragalus fridae as an endangered species. Acta Physiol. Plant. 2019, 41(4), 1–13.
- [22] Manosroi, A.; et al. Entrapment enhancement of peptide drugs in niosomes. J. Microencapsul. 2010, 27(3), 272–280
- [23] Muller, J.M.; et al. VIP as a cell-growth and differentiation neuromodulator role in neurodevelopment. Mol. Neurobiol. 1995, 10(2–3), 115–134.
- [24] Kumar, G.P.; Rajeshwarrao, P. Nonionic surfactant vesicular systems for effective drug delivery—an overview. Acta Pharm. Sin. B. 2011, 1(4), 208–219.
- [25] Weng, Y.; et al. Nanotechnology-based strategies for treatment of ocular disease. Acta Pharm. Sin. B. 2017, 7(3), 281–291.
- [26] Barzadeh, I.; et al. Niosomal formulation for coadministration of hydrophobic anticancer drugs into MCF-7 cancer cells. Arch. Adv. Biosci. 2020, 11(2), 1–9.
- [27] Ghafelehbashi, R.; et al. Preparation, physicochemical properties, in vitro evaluation and release behavior of cephalexin-loaded niosomes. Int. J. Pharm. 2019, 569, 118580.
- [28] Hedayati Ch, M.; et al. Niosome-encapsulated tobramycin reduced antibiotic resistance and enhanced antibacterial activity against multidrugresistant clinical strains of Pseudomonas aeruginosa. J. Biomed. Mater. Res. Part A. 2021, 109(6), 966–980.
- [29] Akbarzadeh, I.; et al. Preparation, optimization and in-vitro evaluation of curcumin-loaded Niosome@ calcium alginate nanocarrier as a new approach for breast cancer treatment. Biology. 2021, 10(3), 173.

- [30] Akbarzadeh, I.; et al. Optimized doxycyclineloaded niosomal formulation for treatment of infection-associated prostate cancer: an in-vitro investigation. J. Drug Deliv. Sci. Technol. 2020, 57, 101715.
- [31] Moghaddam, F.D.; et al. Delivery of melittinloaded niosomes for breast cancer treatment: an in vitro and in vivo evaluation of anti-cancer effect. Cancer Nanotechnol. 2021, 12(1), 1–35.
- [32] Khanam, N.; et al. Recent trends in drug delivery by niosomes: a review. Asian J. Pharm. Res. Dev. 2013, 1, 115–122.
- [33] Sadeghi, S.; et al. Design and physicochemical characterization of lysozyme loaded niosomal formulations as a new controlled delivery system. Pharm. Chem. J. 2020, 53(10), 921–930.
- [34] Shirzad, M.; et al. The role of polyethylene glycol size in chemical spectra, cytotoxicity, and release of PEGylated nanoliposomal cisplatin. Assay Drug Dev. 2019, 17(5), 231–239.
- [35] Kazi, K.M.; et al. Niosome: a future of targeted drug delivery systems. J. Adv. Pharm. Technol. Res. 2010, 1(4), 374.
- [36] Targhi, A.A.; et al. Synergistic effect of curcumin-Cu and curcumin-Ag nanoparticle loaded niosome: enhanced antibacterial and anti-biofilm activities. Bioorg. Chem. 2021, 115, 105116.
- [37] Moghtaderi, M.; et al. Enhanced antibacterial activity of Echinacea angustifolia extract against multidrug-resistant Klebsiella pneumoniae through niosome encapsulation. Nanomaterials 2021, 11(6), 1573.
- [38] Onoue, S.; Yamada, S.; Chan, H.-K.J. Nanodrugs: pharmacokinetics and safety. Int. J. Nanomed. 2014, 9, 1025.
- [39] Baillie, A.J.; et al. The preparation and properties of niosomes–non-ionic surfactant vesicles. J. Pharm. Pharmacol. 1985, 37(12), 863–868.
- [40] Jin, C.; et al. Application of nanotechnology in cancer diagnosis and therapy-a mini-review. Int. J. Med. Sci. 2020, 17(18), 2964.
- [41] Din, F.U.; Shah, S.U. Proniosomes derived niosomes: recent advancements in drug delivery and targeting. Drug Deliv. 2017, 24, 56.
- [42] Akbarzadeh, I.; et al. Niosomal delivery of simvastatin to MDA-MB-231 cancer cells. Drug Dev. Ind. Pharm. 2020, 46(9), 1535–1549.

#### www.jchr.org



- [43] Naseroleslami, M.; et al. Simvastatin-loaded nanoniosomes confer cardioprotection against myocardial ischemia/reperfusion injury. Drug Deliv. Transl. Res. 2021, 12, 1423.
- [44] Moghassemi, S.; Hadjizadeh, A. Nano-niosomes as nanoscale drug delivery systems: an illustrated review. J. Control. Release 2014, 185, 22–36.
- [45] Shi, C.; et al. A drug-specific nanocarrier design for efficient anticancer therapy. Nat. Commun. 2015, 6(1), 1–14.
- [46] Mirzaie, A.; et al. Preparation and optimization of ciprofloxacin encapsulated niosomes: a new approach for enhanced antibacterial activity, biofilm inhibition and reduced antibiotic resistance in ciprofloxacin-resistant methicillin-resistance Staphylococcus aureus. Bioorg. Chem. 2020, 103, 104231.
- [47] Iranbakhsh, A.; Ardebili, Z.O.; Ardebili, N.O. Synthesis and characterization of zinc oxide nanoparticles and their impact on plants. Plant Responses to Nanomaterials: Recent Interventions, and Physiological and Biochemical Responses, 2021, 33–93.
- [48] Khan, R.; Irchhaiya, R.J. Niosomes: a potential tool for novel drug delivery. J. Pharm. Investig. 2016, 46(3), 195–204.
- [49] Jamshidifar, E.; et al. Super magnetic niosomal nanocarrier as a new approach for treatment of breast cancer: a case study on SK-BR-3 and MDA-MB-231 cell lines. Int. J. Mol. Sci. 2021, 22(15), 7948.
- [50] Ge, X.; et al. Advances of non-ionic surfactant vesicles (niosomes) and their application in drug delivery. Pharmaceutics 2019, 11(2), 55.
- [51] Marianecci, C.; et al. Niosomes from 80s to present: the state of the art. Adv. Colloid Interface Sci. 2014, 205, 187–206.
- [52] Jiao, J. Polyoxyethylated nonionic surfactants and their applications in topical ocular drug delivery. Adv. Drug Deliv. Rev. 2008, 60(15), 1663–1673.
- [53] Biswal, S.; et al. Vesicles of non-ionic surfactants (niosomes) and drug delivery potential. Int. J. Pharm. Sci. Nanotechnol. 2008, 1(1), 1–8.
- [54] Shahiwala, A.; Misra, A.J. Studies in topical application of niosomally entrapped nimesulide. J. Pharm. Pharm. Sci. 2002, 5(3), 220–225.
- [55] Pardakhty, A.; Varshosaz, J.; Rouholamini, A.J. In vitro study of polyoxyethylene alkyl ether

- niosomes for delivery of insulin. Int. J. Pharm. 2007, 328(2), 130–141.
- [56] Mohanty, A.; Dey, J.J. Enantioselectivity of vesicle-forming chiral surfactants in capillary electrophoresis: role of the surfactant headgroup structure. J. Chromatogr. A. 2006, 1128(1–2), 259–266.
- [57] Sharma, S. K.; Bhatt, P.; Asdaq, S. M. B.; Alshammari, M. K.; Alanazi, A.; Alrasheedi, N. S.; Alrashdi, B. A.; Alyami, S. S.; Alhazmi, B. H.; Alam, P.; Sharma, P.; Tomar, R.; Arora, M. K.; Imran, M. Combined Therapy with Ivermectin and Doxycycline Can Effectively Alleviate the Cytokine Storm of COVID-19 Infection amid Vaccination Drive: A Narrative Review. J. Infect. Public Health 2022, 15 (5), 566–572. <a href="https://doi.org/10.1016/j.jiph.2022.03.014">https://doi.org/10.1016/j.jiph.2022.03.014</a>.
- [58] Chand, S. P.; Debnath, S.; Rahimi, M.; Ashraf, M. S.; Bhatt, P.; Rahin, S. A. Contextualization of Trait Nexus and Gene Action for Quantitative and Qualitative Characteristics in Indian Mustard. J. Food Qual. 2022, 2022, 1–24. https://doi.org/10.1155/2022/4387318.
- [59] Singh, S.; Pankaj, B.; Nagarajan, K.; P. Singh, N.; Bala, V. Blockchain with Cloud for Handling Healthcare Data: A Privacy-Friendly Platform. Mater. Today 2022, 62, 5021–5026. https://doi.org/10.1016/j.matpr.2022.04.910.
- [60] Ahamed, S.; Bhatt, P.; Sultanuddin, S. J.; Walia, R.; Haque, M. A.; InayathAhamed, S. B. An Intelligent IoT Enabled Health Care Surveillance Using Machine Learning. In 2022 International Conference on Advances in Computing, Communication and Applied Informatics (ACCAI); IEEE, 2022.
- [61] Bhatt, P.; Kumar, V.; Goel, R.; Sharma, S. K.; Kaushik, S.; Sharma, S.; Shrivastava, A.; Tesema, M. Structural Modifications and Strategies for Native Starch for Applications in Advanced Drug Delivery. Biomed Res. Int. 2022, 2022, 1–14. <a href="https://doi.org/10.1155/2022/2188940">https://doi.org/10.1155/2022/2188940</a>.
- [62] Bhatt, P.; Singh, S.; Kumar Sharma, S.; Rabiu, S. Development and Characterization of Fast Dissolving Buccal Strip of Frovatriptan Succinate Monoydrate for Buccal Delivery. Int. J. Pharm. Investig. 2021, 11 (1), 69–75. <a href="https://doi.org/10.5530/ijpi.2021.1.13">https://doi.org/10.5530/ijpi.2021.1.13</a>.

#### www.jchr.org



- [63] Al-Snafi, A. E.; Singh, S.; Bhatt, P.; Kumar, V. A Review on Prescription and Non-Prescription Appetite Suppressants and Evidence-Based Method to Treat Overweight and Obesity. GSC biol. pharm. sci. 2022, 19 (3), 148–155. https://doi.org/10.30574/gscbps.2022.19.3.0231.
- [64] Singh, S.; Bhatt, P.; Alfuraiji, N.; Thuwaini, M. M.; Snafi, A. E. Cardiovascular Comorbidity of COVID-19 Disease: A Review. WJPMR 2022, 8 (4), 216–225.
- [65] Malik, M. K.; Bhatt, P.; Kumar, T.; Singh, J.; Kumar, V.; Faruk, A.; Fuloria, S.; Fuloria, N. K.; Subrimanyan, V.; Kumar, S. Significance of Chemically Derivatized Starch as Drug Carrier in Developing Novel Drug Delivery Devices. Nat. Prod. J. 2022, 12. <a href="https://doi.org/10.2174/2210315512666220819112">https://doi.org/10.2174/2210315512666220819112</a> 334.
- [66] Pankaj. Anti-Cancer Cyclodextrin Nanocapsules Based Formulation Development for Lung Chemotherapy. J. Pharm. Res. Int. 2021, 54–63. https://doi.org/10.9734/jpri/2020/v32i3931024.
- [67] Pankaj. Cyclodextrin Modified Block Polymer for Oral Chemotherapy. J. Pharm. Res. Int. 2021, 21–29. https://doi.org/10.9734/jpri/2020/v32i3831009.
- [68] Singh, S.; Kumar Sharma, S.; Mehrotra, P.; Bhatt, P.; Kaurav, M. Blockchain Technology for Efficient Data Management in Healthcare System: Opportunity, Challenges and Future Perspectives. Mater. Today 2022, 62, 5042–5046. <a href="https://doi.org/10.1016/j.matpr.2022.04.998">https://doi.org/10.1016/j.matpr.2022.04.998</a>.
- [69] Malik, M. K.; Bhatt, P.; Singh, J.; Kaushik, R. D.; Sharma, G.; Kumar, V. Preclinical Safety Assessment of Chemically Cross-Linked Modified Mandua Starch: Acute and Sub-Acute Oral Toxicity Studies in Swiss Albino Mice. ACS Omega 2022, 7 (40), 35506–35514. https://doi.org/10.1021/acsomega.2c01309.
- [70] Kaur, T.; Singh, S. Controlled Release of Bi-Layered Malvidin Tablets Using 3D Printing Techniques. J. Pharm. Res. Int. 2021, 70–78. <a href="https://doi.org/10.9734/jpri/2020/v32i4031034">https://doi.org/10.9734/jpri/2020/v32i4031034</a>.
- [71] Bhatt, P.; Singh, S.; Sharma, S. K.; Kumar, V. Blockchain Technology Applications for Improving Quality of Electronic Healthcare System. In Blockchain for Healthcare Systems; CRC Press: Boca Raton, 2021; pp 97–113.

- [72] Singh, S.; Bhatt, P.; Sharma, S. K.; Rabiu, S. Digital Transformation in Healthcare: Innovation and Technologies. In Blockchain for Healthcare Systems; CRC Press: Boca Raton, 2021; pp 61–79.
- [73] Malik, M. K.; Kumar, V.; Singh, J.; Bhatt, P.; Dixit, R.; Kumar, S. Phosphorylation of Alkali Extracted Mandua Starch by STPP/STMP for Improving Digestion Resistibility. ACS Omega 2023, 8 (13), 11750–11767. <a href="https://doi.org/10.1021/acsomega.2c05783">https://doi.org/10.1021/acsomega.2c05783</a>.
- [74] Bhatt, P.; Kumar, A.; Shukla, R. Nanorobots Recent and Future Advances in Cancer or Dentistry Therapy- A Review. Am. J. PharmTech Res. 2019, 9 (3), 321–331. https://doi.org/10.46624/ajptr.2019.v9.i3.027.
- [75] Sharma, S. K.; Bhatt, P. Controlled Release of Bi-Layered EGCG Tablets Using 3D Printing Techniques. J. Pharm. Res. Int. 2021, 5–13. https://doi.org/10.9734/jpri/2020/v32i3931019.
- [76] Kumar, A.; Bhatt, P.; Mishra, N. Irritable Bowel Syndrome with Reference of Alosetron Hydrochloride and Excipient Profile Used in the Manufacturing of Alosetron Tablet-A Review. J. Chem. Pharm. Sci. 2019, 12 (03), 71–78. https://doi.org/10.30558/jchps.20191203002.
- [77] Bhatt, P.; Kumar, V.; Subramaniyan, V.; Nagarajan, K.; Sekar, M.; Chinni, S. V.; Ramachawolran, G. Plasma Modification Techniques for Natural Polymer-Based Drug Delivery Systems. Pharmaceutics 2023, 15 (8). <a href="https://doi.org/10.3390/pharmaceutics15082066">https://doi.org/10.3390/pharmaceutics15082066</a>.
- [78] Bhatt, P.; Kumar, V.; Rastogi, H.; Malik, M. K.; Dixit, R.; Garg, S.; Kapoor, G.; Singh, S. Functional and Tableting Properties of Alkali-Isolated and Phosphorylated Barnyard Millet (Echinochloa Esculenta) Starch. ACS Omega 2023, 8 (33), 30294–30305. https://doi.org/10.1021/acsomega.3c03158.
- [79] Encyclopedia of Green Materials; Baskar, C., Ramakrishna, S., Daniela La Rosa, A., Eds.; Springer Nature Singapore: Singapore, 2022.
- [80] Bhatt, P.; Singh, S.; Kumar, V.; Nagarajan, K.; Mishra, S. K.; Kumar Dixit, P.; Kumar, V.; Kumar, S. Artificial Intelligence in Pharmaceutical Industry: Revolutionizing Drug Development and Delivery. The Chinese Journal of Artificial Intelligence 2023, 02.

www.jchr.org



- https://doi.org/10.2174/0129503752250813231124 092946.
- [81] Sahu, K. K.; Kaurav, M.; Bhatt, P.; Minz, S.; Pradhan, M.; Khan, J.; Sahu, R. K.; Yadav, K. Utility of Nanomaterials in Wound Management. In Nanotechnological Aspects for Next-Generation Wound Management; Elsevier, 2024; pp 101–130.
- [82] Bhaskaran, S.; Panigrahi, L.J. Formulation and evaluation of niosomes using different non-ionic surfactants. Indian J. Pharm. Sci. 2002, 64(1), 63.
- [83] Rogerson, A.; et al. The distribution of doxorubicin in mice following administration in niosomes. J. Pharm. Pharmacol. 1988, 40(5), 337–342.
- [84] Yadavar-Nikravesh, M-S.; et al. Construction and characterization of a novel tenofovir-loaded pegylated niosome conjugated with tat peptide for evaluation of its cytotoxicity and anti-HIV effects. Adv. Powder Technol. 2021, 32(9), 3161–3173.
- [85] Sahu, A.K.; Mishra, J.; Mishra, A.K. Introducing Tween-curcumin niosomes: preparation, characterization and microenvironment study. Soft Matter 2020, 16(7), 1779–1791.
- [86] Uchegbu, I.F.; Florence, A.T. Non-ionic surfactant vesicles (niosomes): physical and pharmaceutical chemistry. Adv. Colloid Interface Sci. 1995, 58(1), 1–55.
- [87] Yoshioka, T.; Sternberg, B.; Florence, A.T. Preparation and properties of vesicles (niosomes) of sorbitan monoesters (Span 20, 40, 60 and 80) and a sorbitan triester (Span 85). Int. J. Pharm. 1994, 105(1), 1–6.
- [88] Mokhtar, M.; et al. Effect of some formulation parameters on flurbiprofen encapsulation and release rates of niosomes prepared from proniosomes. Int. J. Pharm. 2008, 361(1–2), 104–111.
- [89] Obeid, M.A.; et al. The effects of hydration media on the characteristics of non-ionic surfactant vesicles (NISV) prepared by microfluidics. Int. J. Pharm. 2017, 516(1–2), 52–60.
- [90] Sahin, N.O. Niosomes as nanocarrier systems. In: Mozafari, M.R., editor. Nanomaterials and Nanosystems for Biomedical Applications. Dordrecht: Springer; 2007, p. 67–81.
- [91] Marianecci, C.; et al. Non-ionic surfactant vesicles in pulmonary glucocorticoid delivery: characterization and interaction with human lung

- fibroblasts. J. Control. Release. 2010, 147(1), 127–135.
- [92] Mizrahy, S.; et al. Hyaluronan-coated nanoparticles: the influence of the molecular weight on CD44-hyaluronan interactions and on the immune response. J. Control. Release. 2011, 156(2), 231–238.
- [93] Liu, T.; et al. Structure behaviors of hemoglobin in PEG 6000/Tween 80/Span 80/H2O niosome system. Colloids Surf A. 2007, 293(1–3), 255– 261.
- [94] Pouyani, T.; Prestwich, G.D. Functionalized derivatives of hyaluronic acid oligosaccharides: drug carriers and novel biomaterials. Bioconjug. Chem. 1994, 5(4), 339–347.
- [95] Kaur, D.; Kumar, S.J. Niosomes: present scenario and future aspects. J. Drug Deliv. Ther. 2018, 8(5), 35–43
- [96] Ong, S.G.M.; et al. Evaluation of extrusion technique for nanosizing liposomes. Pharmaceutics. 2016, 8(4), 36.
- [97] Jain, S.; et al. Mannosylated niosomes as adjuvant-carrier system for oral genetic immunization against hepatitis B. Immunol. Lett. 2005, 101(1), 41–49.
- [98] Devaraj, G.N.; et al. Release studies on niosomes containing fatty alcohols as bilayer stabilizers instead of cholesterol. J. Colloid Interfaces Sci. 2002, 251(2), 360–365.
- [99] Baillie, A.; et al. The preparation and properties of niosomes—non-ionic surfactant vesicles. J. Pharm. Pharmacol. 1985, 37(12), 863–868.
- [100] Khan, D.H.; et al. Process optimization of ecological probe sonication technique for production of rifampicin loaded niosomes. J. Drug Deliv. Sci. Technol. 2019, 50, 27–33.
- [101] Chen, S.; et al. Recent advances in non-ionic surfactant vesicles (niosomes): fabrication, characterization, pharmaceutical and cosmetic applications. Eur. J. Pharm. Biopharm. 2019, 144, 18–39.