www.jchr.org

JCHR (2023) 13(6), 653-658 | ISSN:2251-6727



# Study on the Buccal Epithelial Cells in Beedi Smokers and Non Smokers in Dharmapuri District (Tamilnadu)

<sup>1</sup> Rathika R, <sup>2</sup> Mahmoodah Parveen K, <sup>3</sup> Srinivasan K, <sup>4</sup> Murali M, <sup>5</sup> Suganthi P

<sup>1</sup>Associate Professor, PG & Research Department of Zoology, Government Arts College, Affiliated to Periyar University, Dharmapuri – 636705, Tamil Nadu.

<sup>2</sup> Assistant Professor, PG & Research Department of Chemistry, Jamal Mohamed College (Autonomous), Affiliated to Bharathidasan University, Tiruchirappalli-620020, Tamil Nadu

<sup>3</sup> Guest Lecturer, PG & Research Department of Zoology, Government Arts College, Affiliated to Periyar University, Dharmapuri – 636705, Tamil Nadu

<sup>4</sup> Guest Lecturer. Department of Zoology, Bharat Ratna Puratchi Thalaivar Dr. MGR Government Arts and Science College, Affiliated to Periyar University, Palacode-636808, Tamil Nadu

<sup>5</sup> Research Associate, KIRND Institute of Research and Development Pvt. Ltd., Tiruchirappalli-620020, Tamil Nadu.

(Reco	eived: 07 October 2023	Revised: 12 November	Accepted: 06 December)	
KEYWORDS Buccal smear, Beedi smokers, Micronucleus, Tobacco	<ul> <li>ABSTRACT: Introduction: Even though it is commonly recognised that tobacco smoke contains carcinogens, findings in the scientific literature that relate smoking behaviours to the frequency of micronuclei are not quite clear-cut. Objectives: A study was carried out to compare and assess the prevalence of micronuclei in chronic beedi smokers and non-smokers in the Dharmapuri district of Tamil Nadu.</li> <li>Methods: The study had one hundred male subjects who were recruited at random. 50 participants were the chronic beedi smokers and the other 50 were non-smokers. The mean age of the smokers was 44.48±4.2, while the mean age of the controls was 42.11±4.9. Buccal smears were collected and used for micronucleus analysis.</li> </ul>			
	Results: In both controls existence of micronucle (p>0.05). Subjects who group based on the frequ	and subjects, there was a significant correct i (p>0.05) and between tobacco use an smoke tendu leaves frequently had a hig ency of habit and age.	lation between radiation exposure and the d the frequency of micronucleated cells her incidence of MN within the smokers	
	Conclusion: This study a have been accustomed to of micronuclei, ionising the contributing aspects improve the specificity, research.	attempted to quantify the cytogenetic dama o consuming tendu leaves. Although tobac radiation and individual habits also influer are the smear site, sample size, and use cytogenetic changes such as pyknosis, ka	age to the oral mucosa in individuals who co is a major contributor to the formation nee the frequency of micronuclei. Some of of nuclear specific dye. Furthermore, to ryolysis, etc., can be added in subsequent	

#### 1. Introduction

The intricate blend of more than 4,000 chemical components found in cigarette smoke easily combines to create additional reactive chemicals that harm DNA and have cytogenetic effects.<sup>1, 2</sup> Low-grade tobacco is hand-rolled into tendu (*Diospyros melanoxylon*) leaves and knotted with cotton thread to create the traditional beedi (Bidi) cigarette.3. Even though it is commonly

recognised that tobacco smoke contains carcinogens, findings in the scientific literature that relate smoking behaviours to the frequency of micronuclei are not quite clear-cut.4

Numerous assays have been suggested as possible biomarkers in biomonitoring investigations; however, these techniques are usually time-consuming and laborintensive, requiring highly skilled personnel to

www.jchr.org

JCHR (2023) 13(6), 653-658 | ISSN:2251-6727



appropriately read and interpret slides. To this end, the application of the micronucleus test to uncultured exfoliated cells generated a great lot of enthusiasm.2, 5, 6 Buccal epithelial cells have the benefit of quick and simple sampling in addition to offering an alternate source.5

Acentric fragments or entire chromosomes that are absent from the daughter cells' major nuclei give rise to the micronucleus. Both drugs that alter the spindle machinery (aneugens) and those that promote chromosomal break age (clastogens) can induce the creation of micronuclei. Micronuclei can arise due to a variety of endogenous and external causes. Radiation, chemicals, and the infiltration of microorganisms are examples of exogenous influences. Genetic flaws, pathological alterations, a lack of vital nutrients (like folic acid), and wounds brought on by harmful metabolic products (like reactive oxygen species) are examples of endogenous causes.7-19 Thus, a study was carried out to assess and compare the prevalence of micronuclei in chronic beedi smokers and non-smokers in Dharmapuri District, Tamil Nadu.

#### 2. Objectives

Both short-term and long-term effects of smoking are detrimental to the immune system and respiratory system. It is a significant risk factor for lung conditions such as asthma, ARDS, idiopathic pulmonary fibrosis (IPF), chronic obstructive pulmonary disease (COPD), and cancer. The harmful ingredient that adds flavour to tobacco products is nicotine. The main factor influencing immunity and lung health is also nicotine. It increases the inflammatory response of neutrophils and macrophages, but it also inhibits both of these cell types' ability to phagocytose other species. Nicotine also has a significant impact on the functions of epithelial cells, and it inhibits both the proliferation of lymphocytes and cytotoxic action.

An essential part of lung defence is played by respiratory epithelial cells. They provide a barrier that keeps pathogenic and dangerous substances out. Furthermore, mucus and antimicrobial peptides are secreted by epithelial cells to aid in host defence. An integral aspect of lung health is the preservation of epithelial cell health. Chronic nicotine exposure has been linked to an increase in goblet cell hyperplasia and effects on mucociliary transport. Nicotine's enhancement of lung inflammation and prevention of apoptosis is the primary source of respiratory epithelium damage and cancer-causing survival.

#### 3. Methods

#### Sample collection

The study included one hundred participants: Fifty chronic beedi smokers and fifty non-smokers who served as the control group. Following a thorough mouth rinse, samples were taken from the buccal mucosa using a wooden spatula and spread out on sterile glass slides and transported in a wooden box to the laboratory for additional processing in a matter of three to four hours.

#### Sample processing

For the purpose of clearing debris and fixing the cells, slides were fixated in a freshly made, refrigerated fixative (3 methanol:1 glacial acetic acid) for 20 to 25 minutes. Slides were hydrolysis (1N HCl) at  $60^{\circ}$ C, these were air-dried and then cleaned with distilled water before being air-dried again. After staining the slides for 25 minutes at 40°C with 2% aceto-orcein, they were cleaned with ethanol and water. A binocular microscope was used to examine the DPX mounted slides at a magnification of  $40 \times$ .

#### **Micronucleus Scoring**

Micronuclei (MN) were detected in 2,000 cells at a magnification of  $40\times$ . Their presence was verified by an additional scorer at random and by magnification of  $100\times$  (oil immersion).

#### **Statistical Analysis**

Data were compiled and ready for interoperability with the statistical programme Statistical Package for the Social Sciences (SPSS) version 19.0 after being entered into Microsoft Excel using a Windows 7 computer. The mean, medium median, Pearson chi square, p value, and cross tabulations of the frequency and existence of micronuclei in habit users are examples of descriptive and inferential statistics. The values of mean, variance, standard error, and standard deviation were computed.

#### 4. Results

Following the collection of baseline data from each respondent, a variety of factors were examined,

www.jchr.org

JCHR (2023) 13(6), 653-658 | ISSN:2251-6727



including age, education, occupation, dietary habits, brushing habits, frequency of brushing, smoking habits, length of smoking, and frequency of smoking. The buccal mucosa scrapings of the control and smoker groups were used to compute the Micronuclei (MN) cells. For every subject, the total MN cells were also determined. Figure 1A showed the definite squamous shaped buccal epithelial cells collected from the nonsmokers i.e., control group. Figure 1B depicted the presence of nucleus (N) and binucleated cells (BN) which were responsible for the formation of micronuclei. Age of the subjects were categorized into two types as (i) <30 and (ii) <30 years. In non-smokers (control) group, 31 individuals belong to category I (<30 years) and 19 individuals belongs to category II (>30years). In smokers group, 35 individuals belong to category I (<30 years) and 15 individuals belongs to category II (>30years) (Figure 2). The mean age of the smokers was 44.48±4.2, while the mean age of the controls was 42.11±4.9 were observed.

# Figure 1. Microscopic observation of buccal epithelial cells of non-smoker and smoker subject



**Figure 2.** Age distribution of non-smoker (A) and smoker (B) group.



The subjects tobacco (beedi) smoking habits and the total number of MN cells were cross-tabulated, and the results demonstrated a significant correlation. The Pearson chi-square (6.99) was used, with degrees of freedom (df=2) and p<0.05 (Table 1). Age of respondents (smokers/controls) was cross-tabulated, and the results showed significant connection (p<0.01) as the exposure of beedi smoke increased the incidence of MN in beedi smoker group than control i.e. non-smoker group. In the smokers group, a cross-tabulation of the micronucleated cell count from the buccal mucosa scrapings revealed the increased incidence of micronuclei (p<0.05) the connection was determined to be significant. Among

 
 Table 1. Mean±SD frequency of smoking habits and MN scoring among the testing subjects.

Frequency of smoking (per day)	Control group (non- smoker)	Smoker (Beedi) group	Micronuclei scoring for Smoker group
0	50	NA	NA
<5	NA	3.57±0.99e	1.35±0.78 <sup>e</sup>
6-10	NA	$28.84{\pm}1.67^{a}$	3.46±0.23 <sup>d</sup>
11-15	NA	11.68±1.55°	5.03±0.47°
16-20	NA	13.85±1.87 <sup>b</sup>	5.99±0.97 <sup>a,b,c</sup>
>20	NA	8.45±0.53 <sup>d</sup>	6.24±0.37 <sup>a</sup>
			Chi square – 6.99*

NA-not applicable; Subscripts indicated the homogenous subset between the test groups; \*P<0.05

smokers and controls, the frequency of habit also significantly affected outcomes (Table 1). In smokers and controls, different frequency groups (0, <5, 6 to 10, 11 to 15, 16 to 20, 21 to 25, >25) were compared. Four degrees of freedom were evaluated using the Pearson chi-square test (6.93). There was a noteworthy correlation observed between the occurrence of micronuclei and frequency habit.

www.jchr.org

JCHR (2023) 13(6), 653-658 | ISSN:2251-6727



#### 5. Discussion

A wide range of exploratory and mechanistic studies have made considerable use of the micronucleus (MN) test in an effort to comprehend the fundamental mechanisms behind genotoxicity. Eight factors such as dietary habits, climatic shifts, and physical and chemical genotoxins, can affect genetic damage. In actively proliferating tissues, MN can also be observed following irradiation which offers a practical and trustworthy index for loss and breakage of chromosomes.<sup>20–22</sup>

Human saliva has been shown to contain nitrosamines (TSNA) peculiar to tobacco use. They induce chromatid and chromosomal abnormalities, which raise the rates of MN. They are strong carcinogenic agents.1. Alcohol drinkers who smoked three or more packs of cigarettes a day had eight times more MNd mucosa cells than those who smoked one to two packs 23. Tobacco and alcohol combine to create chromosomal abnormalities that lead to the development of MNi. A different study by Stich et al. (6) also found a strong correlation between reverse cigar smokers and controls because the tobacco's TSNA acts as a strong mutagenic and clastogenic agent.

As TSNA, which are strong carcinogens that cause MN development, are present in beedi smoke, which contains tobacco, a study by Suhas et al. [3] found a substantial correlation between smoking and the frequency of MN in buccal mucosa cells. Angilieri et al. on the other hand, showed that there was no difference in MN frequency in exfoliated buccal mucosa cells and tongue lateral borders between smokers and nonsmokers. Because tobacco has clastogenic and aneugenic effects on buccal mucosa cells, our study found a significant relationship between the number of MN cells in subjects and controls who smoked. However, there was no significant correlation (P=0.14) between the total MNd cells obtained from smokers and controls.

The findings are consistent with the research conducted by Nersesyan et al., which found that staining techniques have a significant impact on the micronuclei tests in exfoliated oral mucosa cells of smokers and nonsmokers. Using non-DNA-specific stains such as Giemsa, smokers had higher micronuclei rates; however, no discernible changes were observed when using Feulgen, AO, etc.24

Nevertheless, aceto-orcein stain, which mostly reacts with histone proteins in the nucleus, was employed as the nuclear stain in our investigation. However, when the interaction with occupational exposure is taken into consideration, heavy smokers were the only group showing a significant increase in genotoxic injury as measured by the micronucleus assay in lymphocytes. Overall, smokers do not experience an increase in MN frequency in lymphocytes, according to Bonassi et al. When compared to nonsmokers, there was a slight drop in MN frequency in current smokers with frequency ratio (FR)=0.97 and in former smokers FR=0.96 with 95% confidence limit.

This is so that cytogenetic changes caused by tobacco carcinogens cannot directly affect lymphocytes.4 Between smokers and controls, the frequency of habit had a significant (p=0.00014) bearing. Because of the corresponding rise in the potentially carcinogenic environment, the number of micronucleated cells in the buccal mucosa grows as the frequency of habit increases, that is, as the number of beedis smoked per day increases.

Bloching et al. <sup>25</sup> showed a highly significant correlation between increasing tobacco consumption (daily cigarettes and pack per years) and a higher MN count because of increased resultant cytogenetic damage. The amount and duration of tobacco abuse is directly correlated with increased exposure to carcinogenic contents in tobacco.<sup>25</sup> Gabriel et al could not detect dose effect, because there was no significant difference in the micronucleus frequency between those who smoked 20 cigarettes per day ( $9.3\pm0.4$  and  $10.1\pm0.8$  respectively) as only a small number of subject (n=8) showed increased habit frequency. However, a higher sample size was required to confirm the test results.<sup>26</sup>

#### Acknowledgment

Authors are thankful to the authorities at PG and Research Department of Zoology, Government Arts College, Dharmapuri for the support during this study.

www.jchr.org

JCHR (2023) 13(6), 653-658 | ISSN:2251-6727



- 1. Husgafvel-Pursiainen K. Genotoxicity of environmental tobacco smoke: a review. Mutat Res 2004 Nov;567(2-3): 427-445.
- Angelieri F, de Cássia Gonçalves Moleirinho T, Carlin V, Oshima CT, Ribeiro DA. Biomonitoring of oral epithelial cells in smokers and non-smokers submitted to panoramic X-ray: Comparison between buccal mucosa and lateral border of the tongue. Clin Oral Investig 2010 Dec;14(6):669-674.
- Suhas S, Ganapathy KS, Gayatridevi M, Ramesh C. Application of the micronucleus test to exfoliated epithelial cells from the oral cavity of beedi smokers, a high-risk group for oral cancer. Mutat Res 2004 Jul;561(1-2):15-21.
- Bonassi S, Neri M, Lando C, Ceppi M, Lin Y-P, Chang WP, Holland N, Kirsch-Volders M, Zeiger E, Fenech M. Effect of smoking habit on the frequency of micronuclei in human lymphocytes: results from the Human MicroNucleus project. Mutat Res 2003 Mar;543(2):155-166.
- Tolbert PE, Shy CM, Allen JW. Micronuclei and other nuclear anomalies in buccal smears: a field test in snuff user. Am J Epidemiol 1991; 134(8):840-850.
- Stich HF, Parida BB, Brunneman KD. Localized formation of micronuclei in the oral mucosa and tobacco-specific nitrosamines in the saliva of "reverse" smokers, khaini-tobacco chewers and gudakhu users. Int J Cancer 1992, 50(2): 172-176.
- Majer BJ, Laky B, Knasmuller S, Kassie F. Use of the micronucleus assay with exfoliated epithelial cells as a biomarker for monitoring individuals at elevated risk of genetic damage and in chemoprevention trials. Mutat Res 2001, 489(2-3): 147-172.
- Elhajouji A, Lukamowicz M, Cammerer Z, Kirsch-Volders M. Potential thresholds for genotoxic effects by micronucleus scoring. Mutagenesis 2011, 26(1):199-204.
- Crott JW, Mashiyama ST, Ames BN, Fenech M. The effect of folic acid deficiency and MTHFR C677T Polymorphism on

Journal of Consider Loadsk Rass Particular Ras

chromosome damage in human lymphocytes in vitro. Cancer Epidemiol Biomarkers Prev 2001, 10(10):1089-1096.

- Ban S, Shinohara T, Hirai Y, Moritaku Y, Cologne JB, MacPhee DG. Chromosomal instability in BRCA1- or BRCA2-defective human cancer cells detected by spontaneous micronucleus assay. Mutat Res 2011, 474(1-2):15-23.
- Petrozzi L, Lucetti C, Scarpato R, Gambaccini G, Trippi F, Bernardini S, Del Dotto P, Migliore L, Bonuccelli U. Cytogenetic alterations in lymphocytes of Alzheimers's disease and Parkinson's disease patients. Neurol Sci 2002 23(Suppl 2): S97-S98.
- 12. Mateuca R, Lombaert N, Aka PV, Decordier I, Kirsch Volders M. Chromosomal changes: induction, detection methods and applicability in human biomonitoring. Biochimie 2006 Nov;88(11):1515-1531.
- 13. Editorial. The use of the in vitro micronucleus assay to detect and assess the aneugenic activity of chemicals. Mutat Res 2006 Aug;607(1):5-8.
- Lorge E, Thybaud V, Aardema MJ, Oliver J, Wakata A, Lorenzon G, Marzin D. SFTG international collaborative study on in vitro micronucleus test I. General conditions and overall conclusions of the study. Mutat Res 2006 Aug;607(1):13-36.
- Grieshaber SS, Grieshaber NA, Miller N, Hackstadt T. Chlamydia trachomatis causes centrosomal defects resulting in chromosomal segregation abnormalities. Traffic 2007 7(8):940-949.
- 16. Vral A, Fenech M, Thierens H. The micronucleus assay as a biological dosimeter of in vivo ionising radiation exposure. Mutagenesis 2011 Jan;26(1):11-17.
- Bull CF, Beetstra-Hill S, Benassi-Evans BJ, Crott JW, Kimura M, Teo T, Wu J, Fenech MF. Application and adaptation of the in vitro micronucleus assay for the assessment of nutritional requirements of cells for DNA damage prevention. Mutagenesis 2011 Jan;26(1):193-197.

www.jchr.org

JCHR (2023) 13(6), 653-658 | ISSN:2251-6727



- Huang Y, Fenech M, Shi Q. Micronucleus formation detected by live-cell imaging. Mutagenesis 2011 Jan;26(1):133-138.
- 19. Srivastava AK, Srivastava PK, Al-Khedhairy AA, Musarrat J, Shukla Y. Allethrin-induced genotoxicity and oxidative stress in Swiss albino mice. Mutat Res 2012 Aug; 747(1):22-28.
- 20. Fenech M. The cytokinesis block micronucleus technique and its application to genotoxicity studies in human populations. Environ Health Perspect (Suppl) 1993 Oct;101(3):101-107.
- 21. Evans HJ, Neary GJ, Williamson FS. The relative biological efficiency of single doses of fast neutrons and gamma-rays on Vicia faba roots and the effect of oxygen. Int J Rad Biol Relat Stud Phys Chem Med1959 Jul;3:216-229.
- 22. Fenech F. The in vitro micronucleus technique. Mutat Res 2000 Nov;455(1-2):81-95.
- 23. Stich HF, Rosin MP. Quantitating the synergistic effect of smoking and alcohol consumption with the micronucleus test on human buccal mucosa cells. Int J Cancer 1983 Mar;31(3):305-308.
- 24. Nersesyan A, Kundi M, Atefie K, Schulte-Hermann R. Effect of staining procedures on the results of micronucleus assays with exfoliated oral mucosa cells. Cancer Epidemiol Biomarkers Prev 2006 Oct;15(10):1835-1840.
- 25. Bloching M, Hofmann A, Lautenschlager Ch, Berghaus A, Grummt T. Exfoliative cytology of normal buccal mucosa to predict the relative risk of cancer in the upper aerodigestive tract using the MN-assay. Oral Oncol 2000 Nov;36(6):550-555.
- 26. Gabriel HE, Crott JW, Ghandour H, Dallal GE, Choi S-W, Keyes MK, Jang H, Liu Z, Nadeau M, Johnston A, et al. Chronic cigarette smoking is associated with diminished folate status, altered folate form distribution, and increased genetic damage in the buccal mucosa of healthy adults. Am J Clin Nutr 2006 Apr;83(4):835-841.