



Green Preparation and Characterization of Silver Nanoparticles from Aloe Vera and Neem Leaf Extracts and Studying their Antimicrobial Activity against Oral Pathogens

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Abstract

The use of toxic compounds in conventional nanoparticle production processes can cause environmental damage. Green synthesis will therefore be a necessary process to minimize the production of these compounds. Previous studies have shown that silver nanoparticles have potent antibacterial properties. Silver was used to create a nanoparticle because it induces cell wall breaking due to intracellular material leakage, which makes it an efficient antibacterial agent. *Aim* : Aim of the study was to test the antimicrobial properties of silver nanoparticles mixed with aloe and neem leaf extracts against oral pathogens. *Material and methods*: Antibacterial activity of the corresponding nanoparticles against the strains of *Enterococcus faecalis*, *Streptococcus mutans* and *Candida albicans*. For this experiment, MHA agar was used to identify the zone of inhibition. The zone of inhibition was tested after the incubation period. *Result* : With increasing concentrations, increased values are seen for all three bacteria. *Conclusion*: Within the limits of the study it can be concluded that silver nanoparticles have good antimicrobial properties and further can be incorporated in dentistry either as the drug delivery systems or can be used to coating materials to improve their properties.

1. INTRODUCTION

An expanding field of nanoscience and nanotechnology is the application of materials structures at the nanoscale, which typically ranges from 1 to 100 nanometers (nm). In the fields of catalysis, biology, biomedical science, and water treatment, nanomaterials may offer answers to technical and environmental problems(1) The surface-to-volume ratios of nanoparticles are quite high. When a large surface area is required in science, this attribute can be used. As an illustration, several nanoparticles have really shown to be effective catalysts in the catalytic sector(2). The nanoparticles also exhibit antibacterial

properties. Silver nanoparticles have been given credit for these crucial characteristics silver nanoparticles(AgNPs)(2,3). AgNPs have therefore drawn a lot of attention in nano-biotechnological research due to their distinct physical, chemical, and biological characteristics as well as their practicality(1,2). These nanoparticles can be used in the fields of electronics, optics, engineering and even medicine and dentistry(4). Silver nanoparticles can also be employed for a variety of significant applications, such as optical receptors for biolabeling, intercalation



materials for batteries, and spectrally selective coatings for solar energy absorption(5),(6)

Since the fourteenth century, bacteria have been employed as infection-causing agents. The first antibacterial drug was created in 1910 and was called salvarsan. Following that, antimicrobials including macrolides, nalidixic acid, and chloramphenicol were routinely used(7). Silver nanoparticles can also be employed for a variety of significant applications, such as optical receptors for biolabeling, intercalation materials for batteries, and spectrally selective coatings for solar energy absorption(8). The antibacterial and inhibitory properties of AgNPs are well recognised. Pathogenic bacteria have begun to develop resistance to antimicrobial treatments, nonetheless, in recent years. This has been extensively researched and represents a significant problem for the health care sector. Concern over the rise of antibiotic resistance has grown as bacteria quickly develop adaptive defenses against common antibiotics. Bacteria are pathogens that have the capacity to cause death and spread infectious diseases(5,9).

Since then, numerous efforts have been made to curb the emergence of these resistant strains, including the development of new antibiotic drugs with greater chemical diversity, the identification of antibiotic-producing bacteria, and the discovery of additional antibiotics from previously unknown natural sources. However, these advancements were insufficient to offset the rapidly increasing number of resistant bacterial species(7,10).

Finding a simple, cost-effective, eco-friendly alternative method for the synthesis of Ag NPs is crucial to overcoming these types of challenges(11)(12). Microorganisms and plants are non-toxic reproducible biological resources that are safe for people and the environment, and they may be a better alternative to physical and chemical approaches for the synthesis of Ag NPs(13). In recent times, reports of fungus, bacteria, and plants acting as reducing agents in the production of Ag NPs have surfaced. Plants are a better choice than microorganisms for biosynthesis(11). As a result, this avoids the expansive procedure of cell culture and its maintenance(14).

Silver is an essential trace element for the complete body development and health to be achieved. Because of the fact that this is the least poisonous form of elemental Ag, the nanoparticle form of silver has garnered a lot of attention.

This study was hence conducted to study the antimicrobial efficacy of an extract containing aloe vera and neem leaves.

2. MATERIALS AND METHODS

2.1. Preparation of the aloe vera and neem extract

Commercially available organic preparations of aloe leaf vera leaf powder was procured and used. Fresh neem leaves were obtained and washed multiple times with distilled water and left to dry. A sterilized knife was used to cut the air-dried neem leaf into fine pieces. These pieces were then ground using a mortar and pestle into a powder. 1g of this extract was added to 100ml of distilled water to achieve a 1 M concentration of the aloe vera and neem extract. The Whatmann No. 1 filter paper was used to filter the boiling extract. The filtrates were kept at 5°C for upcoming tests.

2.2 Synthesis of the silver nanoparticle

For the bioreduction procedure, aloe vera and neem leaf extract were employed as aqueous extracts. 0.1mM silver nitrate was dissolved in 100ml of distilled water and stirred for a short period of time to facilitate the biosynthesis of AgNPS. 5ml of filtered neem leaf extract and aloe vera extract was then added drop by drop. For 72 hours, the solution mixture was stirred in a magnetic stirrer at 650–800 rpm. A twin beam UV visible spectrophotometer was used to constantly record the colour changes in the reaction mixture at various wavelengths between 25 and 650 nm. The created neem leaf extract and aloe vera extract-mediated silver nanoparticles underwent a 10-minute centrifugation at 8000 rpm. The reaction colour gradually changed to a yellowish brown colloidal suspension, seemingly indicating that the reduction of Ag⁺ ions into metallic Ag NPs was aided by the phytochemicals present in aloe vera and neem leaf extract.

After being cleaned with ethanol and twice-deionized water, the collected Ag NPs were dried at 100 °C for two hours. For the 10 and 15 ml extract volumes, the same process was repeated.

Hourly UV-vis spectrometric data were collected to track the formation of silver nano composites. Centrifuging the resulting mixture produced silver nanoparticles.

TEM analysis was also done to ascertain the size of the silver nanoparticles.

2.3 Determination of the antimicrobial activity

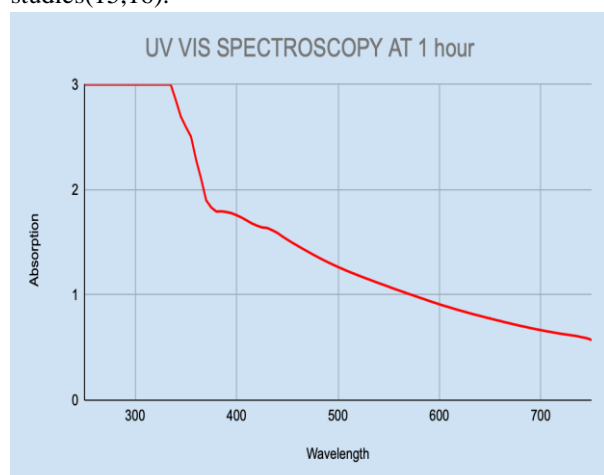


The antibacterial activity of the corresponding nanoparticles against the strains of *Enterococcus faecalis*, *Streptococcus mutans* and *Candida albicans* was being tested. Muller Hinton Agar was employed in this experiment to locate the zone of inhibition. The agar was prepared and sterilised for 45 minutes at 120 lbs. The medium was placed into sterile plates and allowed to harden. After the wells were cut with the well cutter, the test organisms were swabbed. Concentration was added to several nanoparticle plates, which were then maintained at 37 ° C for 24 hours. After the incubation time, the zone of inhibition was evaluated.

3. RESULTS

3.1 UV-vis absorption study

The figure shows the colloidal solution of leaf extract and aqueous silver nitrate's UV-vis absorption spectra readings. The silver salt solution was colourless at first and then changed to brown when aloe vera and neem leaf extract was added to the reaction mixture. The production of Ag NPs in the aqueous medium is firmly confirmed by this change in colour of the silver solution, which is clearly visible. A larger concentration of Ag NPs with a deep brown solution results from an increase in the reduction rate of Ag⁺ ions to Ag caused by an increase in extract volume. In Figure 1, the wavelength is seen between 370-380 nm as in accordance with various studies (15,16).



3.3 Antimicrobial Activity

Figure1: The wavelengths are noted between 370-380nm.

3.2 Surface morphological study

From the TEM analysis, the synthesised Ag NPs' size and form were examined, and the results are presented in Fig. 3a. In Figure 2, according to the TEM image, the produced Ag NPs are uniformly disseminated, primarily spherical in shape, and range in size from 20-30 nm. This number can be correlated with the other studies that were done and is in line with the results. Aloe vera and neem leaf extract contains stabilising phytochemicals that might be responsible for the well-dispersion of AgNPs.

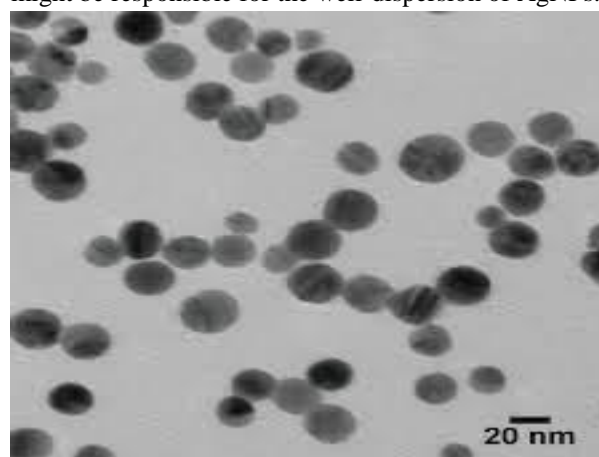


FIGURE 2: SEM analysis revealed the size of the particle at 20nm.



MICRO ORGANISM CONC.	ZONE OF INHIBITION			
	25μ	50μ	100μ	CONTROL
<i>S. Mutans</i>	20	22	24	30
<i>E. Faecalis</i>	12	14	16	28
<i>C. Albicans</i>	11	14	18	24

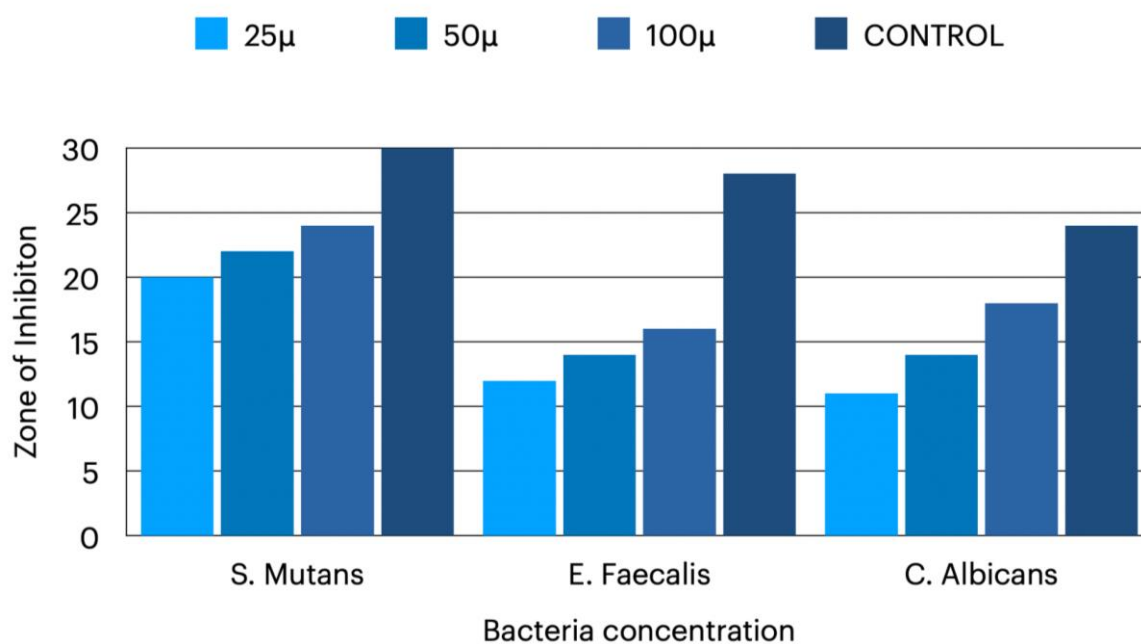
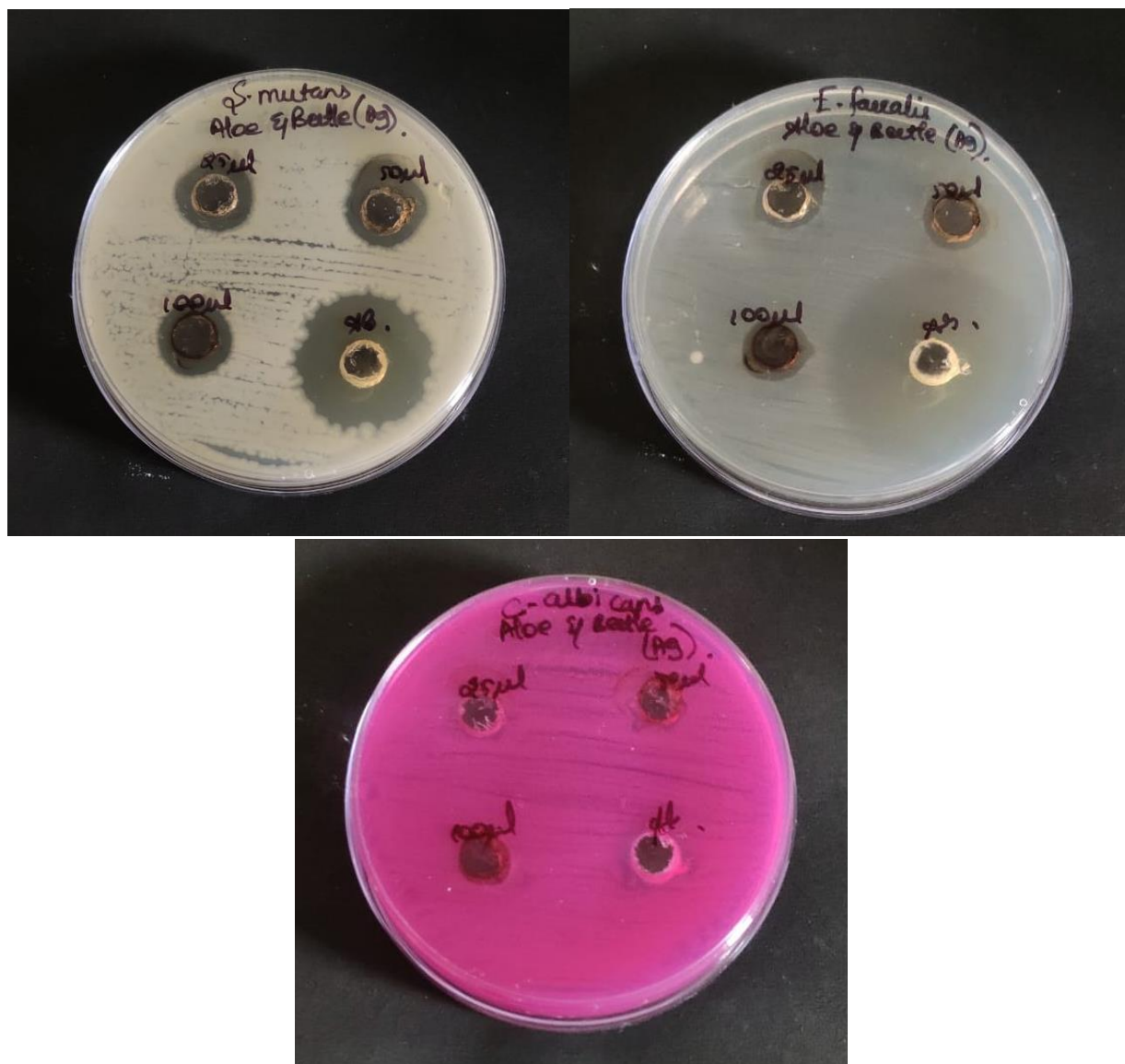


Figure 3 shows the ZOI of *S. Mutans*, Figure 4 shows the ZOI of *E. Faecalis* and Figure 5 shows the ZOI of *C. Albicans*.



At 100 μ , the concentrations of *S. Mutans*, *E. Faecalis* and *C. Albicans* were 24,16 and 18.

DISCUSSION

One of the most common mechanism of action is that AgNPs can build up on the cell wall and membrane, resulting in visibly noticeable morphological alterations such as cytoplasmic shrinkage, membrane separation, many electron dense pits, and ultimately ruptured membrane(17,18) Gram positive and gram negative bacteria have different cell wall compositions, Ag NPs showed greater growth inhibition of gram-positive bacteria(19). Ag NPs generated from varying extract volumes result in variations in the zone of inhibition, which is due to different particle sizes(20)(21). Because of their increased surface area, researchers found that

tiny Ag NPs have superior antibacterial action. Small (nm) sized produced Ag NPs adhere to bacteria's cell wall membrane and even penetrate the bacterium themselves(22). As a result, it hinders the system's normal operation, disrupts the bacteria's respiration mechanism, and ultimately kills them(23). Size of the silver nanoparticles played a very important in the antibacterial properties. In a study conducted by Choi and Hu AgNPs synthesised between 9-21nm showed best antibacterial properties(24,25). Agnihotri et al also concluded that from 100nm to 20nm the best antibacterial properties were seen with the smallest sizes(24). Monroes et al goes on to say that smaller than



10nm sizes can also be very effective in destroying the bacteria(26). Helmlinger et al. have conducted more in-depth research on the shape-effect on antibacterial activities, synthesising five distinct AgNP kinds with remarkably homogeneous size and shape. He came to the conclusion that, in line with the size-dependent antibacterial activity, the antibacterial activity of variously shaped AgNPs is mostly caused by the production of Ag⁺ ions on the surface as a result of dissolution(26,27). AgNPs' antibacterial activity was also discovered to be influenced by their form. In order to test their antibacterial efficacy against *E. coli* in solution and on agar plates, Pal et al. produced spherical, rod-shaped AgNPs and truncated triangular Ag nanoplates(28,29). Truncated triangular silver nanoplates have the highest percentage of exposed facets when compared to silver nanospheres and nanorods. This facilitates direct interaction of the nanoplates with bacterial surfaces, which improves surface binding, cell uptake, and effectively kills bacteria, although both can be effective in killing bacteria(28). Silver nanoparticles' well-known and applied broad-spectrum antibacterial capabilities are employed in wound healing. AgNPs come in a wide variety of sizes, shapes, and surface chemistries; therefore, a thorough understanding of how these structural parameters affect their antibacterial activities can aid in extending the range of biological applications for AgNPs(30).

4. CONCLUSION

Within the limits of this study, we can conclude that silver nanoparticles with the aloe vera and neem leaf extract can be useful against commonly seen oral pathogens. The readings against *S. Mutans* were high at all three concentrations and it was similar to the control. Adequate readings were seen in the case of *E. Faecalis* and *C. Albicans* too. A good antimicrobial activity has been displayed. There is great scope for future research to be done in this field.

5. CONFLICT OF INTEREST

The author claims that there is no conflict of interest.

6. ACKNOWLEDGEMENT

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REFERENCES

1. Alarcon EI, Griffith M, Udekwu KI. Silver Nanoparticle Applications: In the Fabrication and Design of Medical and Biosensing Devices. Springer; 2015. 146 p.
2. Okafor F, Janen A, Kukhtareva T, Edwards V, Curley M. Green Synthesis of Silver Nanoparticles, Their Characterization, Application and Antibacterial Activity. Int J Environ Res Public Health. 2013 Oct 21;10(10):5221–38.
3. Nowack B. Nanosilver Revisited Downstream [Internet]. Vol. 330, Science. 2010. p. 1054–5. Available from: <http://dx.doi.org/10.1126/science.1198074>
4. Website [Internet]. Available from: <https://doi.org/10.47750/jptcp.2022.962>
5. Benn TM, Westerhoff P. Nanoparticle Silver Released into Water from Commercially Available Sock Fabrics [Internet]. Vol. 42, Environmental Science & Technology. 2008. p. 4133–9. Available from: <http://dx.doi.org/10.1021/es7032718>
6. Website [Internet]. Available from: Nanoparticles Inspire Plasmonic Solar Cell. Available online: <http://images.iop.org/objects/ntw/news/8/3/48/pdf.pdf> (accessed on 21 March 2009).
7. Morens DM, Folkers GK, Fauci AS. Emerging infections: a perpetual challenge [Internet]. Vol. 8, The Lancet Infectious Diseases. 2008. p. 710–9. Available from: [http://dx.doi.org/10.1016/s1473-3099\(08\)70256-1](http://dx.doi.org/10.1016/s1473-3099(08)70256-1)
8. Bonsak J, Mayandi J, Thøgersen A, Marstein ES, Mahalingam U. Chemical synthesis of silver nanoparticles for solar cell applications [Internet]. Vol. 8, physica status solidi c. 2011. p. 924–7. Available from: <http://dx.doi.org/10.1002/pssc.201000275>
9. Mackevica A, Olsson ME, Hansen SF. Silver nanoparticle release from commercially available plastic food containers into food simulants [Internet]. Vol. 18, Journal of Nanoparticle Research. 2016. Available from: <http://dx.doi.org/10.1007/s11051-015-3313-x>
10. Li X, Schirmer K, Bernard L, Sigg L, Pillai S, Behra R. Silver nanoparticle toxicity and association with the alga *Euglena gracilis* [Internet]. Vol. 2, Environmental Science: Nano. 2015. p. 594–602. Available from: <http://dx.doi.org/10.1039/c5en00093a>



11. Rajeshkumar S, Agarwal H, Venkat Kumar S, Lakshmi T. Brassica oleracea Mediated Synthesis of Zinc Oxide Nanoparticles and its Antibacterial Activity against Pathogenic Bacteria [Internet]. Vol. 30, Asian Journal of Chemistry. 2018. p. 2711–5. Available from: <http://dx.doi.org/10.14233/ajchem.2018.21562>
12. Nasim I, Rajesh Kumar S, Vishnupriya V, Jabin Z. Cytotoxicity and anti-microbial analysis of silver and graphene oxide bio nanoparticles. Bioinformation. 2020 Nov 30;16(11):831–6.
13. M G, Gomathi M, Prakasam A, Rajkumar PV, Rajeshkumar S, Chandrasekaran R, et al. Phyllanthus reticulatus mediated synthesis and characterization of silver nanoparticles and its antibacterial activity against gram positive and gram negative pathogens [Internet]. Vol. 10, International Journal of Research in Pharmaceutical Sciences. 2019. p. 3099–106. Available from: <http://dx.doi.org/10.26452/ijrps.v10i4.1603>
14. P S, Santhanam P, Rajeshkumar S, Lakshmi T, Roy A. Antifungal activity of neem and Aloe vera formulation mediated zirconium oxide nanoparticles [Internet]. Vol. 10, International Journal of Research in Pharmaceutical Sciences. 2019. p. 2864–8. Available from: <http://dx.doi.org/10.26452/ijrps.v10i4.1565>
15. Website [Internet]. Available from: Anandalakshmi, K., Venugobal, J. & Ramasamy, V. Characterization of silver nanoparticles by green synthesis method using Pedalium murex leaf extract and their antibacterial activity. Appl Nanosci 6, 399–408 (2016). <https://doi.org/10.1007/s13204-015-0449-z>
16. Anandalakshmi K, Venugobal J, Ramasamy V. Characterization of silver nanoparticles by green synthesis method using Pedalium murex leaf extract and their antibacterial activity [Internet]. Vol. 6, Applied Nanoscience. 2016. p. 399–408. Available from: <http://dx.doi.org/10.1007/s13204-015-0449-z>
17. Ponarulselvam S, Panneerselvam C, Murugan K, Aarthi N, Kalimuthu K, Thangamani S. Synthesis of silver nanoparticles using leaves of Catharanthus roseus Linn. G. Don and their antiplasmodial activities [Internet]. Vol. 2, Asian Pacific Journal of Tropical Biomedicine. 2012. p. 574–80. Available from: [http://dx.doi.org/10.1016/s2221-1691\(12\)60100-2](http://dx.doi.org/10.1016/s2221-1691(12)60100-2)
18. Kim JS, Kuk E, Yu KN, Kim JH, Park SJ, Lee HJ, et al. Antimicrobial effects of silver nanoparticles [Internet]. Vol. 3, Nanomedicine: Nanotechnology, Biology and Medicine. 2007. p. 95–101. Available from: <http://dx.doi.org/10.1016/j.nano.2006.12.001>
19. Sneka, Santhakumar P. Antibacterial Activity of Selenium Nanoparticles extracted from Capparis decidua against Escherichia coli and Lactobacillus Species. J Adv Pharm Technol Res. 2021 Aug 6;4452–4.
20. Shilpa R. Synthesis of Nickel Oxide Nanoparticles by Electrochemical Method, Characterization and Photo degradation of Acetic Acid and Study of Antibacterial Activity of Synthesized Nickel Oxide Nanoparticles [Internet]. Vol. 7, International Journal for Research in Applied Science and Engineering Technology. 2019. p. 71–9. Available from: <http://dx.doi.org/10.22214/ijraset.2019.9011>
21. Anna Thomas A, Varghese RM, Rajeshkumar S. Antimicrobial effects of copper nanoparticles with green tea and neem formulation. Bioinformation. 2022 Mar 31;18(3):284–8.
22. Zhang XF, Liu ZG, Shen W, Gurunathan S. Silver Nanoparticles: Synthesis, Characterization, Properties, Applications, and Therapeutic Approaches. Int J Mol Sci [Internet]. 2016 Sep 13;17(9). Available from: <http://dx.doi.org/10.3390/ijms17091534>
23. Soliman H, Elsayed A, Dyaa A. Antimicrobial activity of silver nanoparticles biosynthesised by Rhodotorula sp. strain ATL72 [Internet]. Vol. 5, Egyptian Journal of Basic and Applied Sciences. 2018. p. 228–33. Available from: <http://dx.doi.org/10.1016/j.ejbas.2018.05.005>
24. Agnihotri S, Mukherji S, Mukherji S. Size-controlled silver nanoparticles synthesized over the range 5–100 nm using the same protocol and their antibacterial efficacy [Internet]. Vol. 4, RSC Adv. 2014. p. 3974–83. Available from: <http://dx.doi.org/10.1039/c3ra44507k>
25. Choi O, Hu Z. Size Dependent and Reactive Oxygen Species Related Nanosilver Toxicity to Nitrifying Bacteria [Internet]. Vol. 42, Environmental Science & Technology. 2008. p. 4583–8. Available from: <http://dx.doi.org/10.1021/es703238h>
26. Morones JR, Elechiguerra JL, Camacho A, Holt K,



- Kouri JB, Ramírez JT, et al. The bactericidal effect of silver nanoparticles [Internet]. Vol. 16, Nanotechnology. 2005. p. 2346–53. Available from: <http://dx.doi.org/10.1088/0957-4484/16/10/059>
27. Helmlinger J, Sengstock C, Groß-Heitfeld C, Mayer C, Schildhauer TA, Köller M, et al. Silver nanoparticles with different size and shape: equal cytotoxicity, but different antibacterial effects [Internet]. Vol. 6, RSC Advances. 2016. p. 18490–501. Available from: <http://dx.doi.org/10.1039/c5ra27836h>
28. Pal S, Tak YK, Song JM. Does the Antibacterial Activity of Silver Nanoparticles Depend on the Shape of the Nanoparticle? A Study of the Gram-Negative Bacterium *Escherichia coli* [Internet]. Vol. 73, Applied and Environmental Microbiology. 2007. p. 1712–20. Available from: <http://dx.doi.org/10.1128/aem.02218-06>
29. Nasim I, Jabin Z, Kumar SR, Vishnupriya V. Green synthesis of calcium hydroxide-coated silver nanoparticles using and Linn. leaf extracts: An antimicrobial and cytotoxic activity. J Conserv Dent. 2022 Aug 2;25(4):369–74.
30. Tang S, Zheng J. Antibacterial Activity of Silver Nanoparticles: Structural Effects [Internet]. Vol. 7, Advanced Healthcare Materials. 2018. p. 1701503. Available from: <http://dx.doi.org/10.1002/adhm.201701503>